

Targeting Dependency on the GPX4 Lipid Peroxide Repair Pathway for Cancer Therapy

Hengrui Liu,[†][®] Stuart L. Schreiber,^{*,‡,§}[®] and Brent R. Stockwell^{*,†,||}[®]

[†]Department of Chemistry, Columbia University, New York, New York 10027, United States

[‡]Department of Chemistry and Chemical Biology, Harvard University, Cambridge, Massachusetts 02138, United States

[§]Broad Institute, Cambridge, Massachusetts 02142, United States

Department of Biological Sciences, Columbia University, 550 West 120th Street, MC 4846, New York, New York 10027, United States

ells are dependent on their lipid composition for establishing and modulating membrane structural integrity, morphology, metabolism, and other cellular functions. For example, among the thousands of lipid species that compose eukaryotic cell membranes, the abundance and localization of polyunsaturated fatty acid (PUFA)-containing phospholipids (PUFA-PLs) are major factors in determining the fluidity of cell membranes.¹ Because the *cis* conformation of double bonds in PUFAs hinders efficient stacking of these fatty acid tails, elevated levels of PUFA-PLs contribute to increasing membrane fluidity and thinning.

PUFAs are, however, susceptible to lipid peroxidation via reaction with molecular oxygen at bis-allylic positions, catalyzed by lipoxygenases or through non-enzymatic mechanisms. Oxygenated PUFAs can serve as lipid signals, regulating inflammatory processes, for example. However, peroxy-PUFAs are prone to decompose into reactive species, which can damage biomolecules, such as proteins and nucleic acids. Thus, cells depend on a critical network of proteins to repair PUFA-PL peroxides. A key protein at the center of this repair network is the selenoprotein glutathione peroxidase 4 (GPX4); it is the only peroxidase in mammals capable of reducing phospholipid hydroperoxides within cell membranes.

When GPX4 activity is compromised, lipid peroxidation can cause ferroptosis, an oxidative iron-dependent form of nonapoptotic cell death. Ferroptosis has been implicated in degenerative diseases; for example, ferroptosis inhibitors are protective in models of Parkinson's, Huntington's and Alzheimer's diseases.² On the other hand, ferroptosis induced by exogenous agents is selectively lethal toward tumor cells that are addicted to GPX4 repair activity, which has suggested that inducing ferroptosis may be a beneficial approach to treating some cancers.²

Cancer cells from tissues of diverse origins have been screened for their sensitivity to ferroptosis-inducing compounds.³ Ferroptosis inducers, including GPX4 inhibitors, were found to selectively target cancers with a mesenchymal or otherwise drug-resistant signature. Consistent with the mesenchymal state being associated with drug resistance, another study on persister cancer cells, which are proposed to escape from conventional cytotoxic treatment through a dormant state and then revive to cause tumor relapse, revealed a similar selective dependency on the GPX4 pathway.⁴ Indeed, GPX4 inhibitors were among the compounds most selectively lethal to such persister cells.

Examination of persister cells revealed upregulation of mesenchymal markers and downregulation of epithelial markers. This common mesenchymal character partially explained the extension of GPX4 dependency from highmesenchymal cancers to persister cells; the underlying mechanisms of GPX4 dependency remain to be further illuminated (Figure 1).



Figure 1. Cancer cells undergo EMT to acquire resistance to apoptosis induction and chemotherapy. EMT requires an elevated level of PUFA-PLs, which inevitably makes cancer cells vulnerable to lipid peroxidation and ferroptosis induced by GPX4 inhibition.

Overexpression of mesenchymal state genes is associated with epithelial-mesenchymal transition (EMT). Because EMT increases the motility of tumor cells and enables the invasion of a primary tumor to distant sites, EMT is part of the process of metastasis. EMT also renders cancer cells resistant to apoptosis and chemotherapy.⁵ EMT requires plasma membrane remodeling to increase fluidity, which is associated with an elevated level of biosynthesis of PUFA-PLs. Given that PUFAs are more susceptible to peroxidation than saturated or monounsaturated fatty acid PLs are, cells in an EMT state should be more dependent on GPX4 to control lipid peroxides. Therefore, cancer cells undergoing EMT that acquire resistance to apoptosis induction may inevitably become vulnerable to lipid peroxidation and ferroptosis induced by GPX4 inhibition. The necessity of the EMT regulator and lipogenic factor ZEB1 for GPX4 dependency and sensitivity to fatty acid desaturase-2

Received: March 13, 2018

(FADS2) activation support this theory.³ The disabled antioxidant program in persister cells might be a result of this overwhelming lipid oxidation, considering that recycling of GPX4 consumes reduced glutathione (GSH) and NADPH.⁴ Additionally, therapy-resistant cancer cells with a high-mesenchymal state due to a mesenchymal origin may exhibit similar genetic alterations and protein expression, explaining their vulnerability to ferroptosis.

THERAPEUTIC POSSIBILITIES

As cancer cells can evolve into a high-mesenchymal or related drug-resistant state and become resistant to apoptosis induction, one may be able to selectively target such resistant cancer cells through ferroptosis induction. The most effective compounds in this context might be direct GPX4 inhibitors. For example, *in vivo* xenografts of GPX4-knockout high-mesenchymal therapy-resistant melanoma regressed after the cessation of ferrostatin-1 (a lipophilic antioxidant that suppresses the effect of loss of GPX4) and did not relapse after dabrafenib and trametinib treatment had been ceased, while GPX4 wild-type xenografts continued to grow in both experiments.³

However, because the *gpx4* gene is essential for embryonic development and some adult tissue homeostasis in mice, there is a concern about the potential toxicity of the therapeutic use of GPX4 inhibitors. However, GPX4 inhibitors were selectively lethal to persister cancer cells, with a minimal effect on parental cells and nontransformed cells, suggesting that the addiction to GPX4 may create a suitable therapeutic window. Despite this, the therapeutic window of the GPX4 inhibitor remains to be examined, such as its potential adverse impact on the nervous system and kidney in particular.

In addition to GPX4 direct inhibitors, persister cells might also be vulnerable to other ferroptosis inducers. For example, erastin and its derivatives, which inhibit system xc- and deplete the intracellular GPX4 cofactor GSH, are also selectively lethal to persister cancer cells, with minimal effects on parental cells. Additionally, high-mesenchymal state cancers are sensitive to statins, which can inhibit the biosynthesis of GPX4 and deplete mevalonate-derived coenzyme Q_{10} . Although tumors might develop other resistance mechanisms for compounds such as erastin and statins, the high potential therapeutic index may be useful: for example, knockout of the system xc- gene *slc7A11* has minimal effects in mice, suggesting that normal tissues do not require system xc- activity, at least in mice.

Dependency on GPX4 may thus derive from elevated levels of synthesis and usage of PUFA-PLs by some cancer cells, particularly the most drug-resistant and aggressive types. Further experiments that aimed to examine this lipid metabolism pathway may reveal additional therapeutic targets with different profiles. The GPX4 pathway thus represents a new frontier in targeting some of the most intractable cancers.

AUTHOR INFORMATION

Corresponding Authors

*E-mail: bstockwell@columbia.edu. *E-mail: stuart_schreiber@harvard.edu.

ORCID 0

Hengrui Liu: 0000-0002-5214-1572 Stuart L. Schreiber: 0000-0003-1922-7558 Brent R. Stockwell: 0000-0002-3532-3868

Funding

The research of the authors is supported by the National Cancer Institute and the National Institutes of Health (Grants R35CA209896 and P01CA087497 to B.R.S. and Grants GM038627 and U01CA176152 to S.L.S.).

Notes

The authors declare no competing financial interest.

REFERENCES

(1) Agmon, E., and Stockwell, B. R. (2017) Lipid homeostasis and regulated cell death. *Curr. Opin. Chem. Biol.* 39, 83–89.

(2) Stockwell, B. R., et al. (2017) *Ferroptosis*: A Regulated Cell Death Nexus Linking Metabolism, Redox Biology, and Disease. *Cell 171*, 273–285.

(3) Viswanathan, V. S., et al. (2017) Dependency of a therapyresistant state of cancer cells on a lipid peroxidase pathway. *Nature 547*, 453–457.

(4) Hangauer, M. J., et al. (2017) Drug-tolerant persister cancer cells are vulnerable to GPX4 inhibition. *Nature* 551, 247–250.

(5) Fischer, K. R., et al. (2015) Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* 527, 472–476.