

THE FOUR CHROMOSES FROM CHROMOMYCIN A₃

M.Miyamoto, Y.Kawamatsu, M.Shinohara

Takeda Chemical Industries, Juso, Osaka, Japan

K.Nakanishi, Y.Nakadaira and N.S.Bhacca*

Department of Chemistry, Tohoku University, Sendai, Japan.

(Received 25 May 1964; in revised form 6 July 1964)

Chromomycin is the name given to a group of cancerostatic antibiotics produced by Streptomyces griseus No.7, and is a mixture of several closely related compounds¹⁾. The principal constituent, which is commercially available, has been designated chromomycin A₃¹⁾. The isolation of closely related compounds has been reported by several groups: e.g., aureolic acid²⁾, aburamycin³⁾, LA-7017⁴⁾, M5-18903⁵⁾, NSCA-649⁶⁾, mithramycin⁷⁾, olivomycin⁸⁾.

These antibiotics apparently belong to a new class of compounds, but their structures remain unclarified. This and following communications^{9),10)} deal with the structural elucidation of chromomycin A₃, C₅₁H₇₂O₂₃.

Chromomycin A₃ can be hydrolysed to the four 2,6-dideoxy-sugars, chromoses A¹¹⁾, B, C and D, and the aglycone chromomycinone⁹⁾ by boiling in 50% aqueous acetic acid. Chromose A has been established¹¹⁾ to belong to the D-series and its

* On leave from Varian Associates, Palo Alto, California.

structure along with those of chromosomes B, C and D is shown in FIG.1.

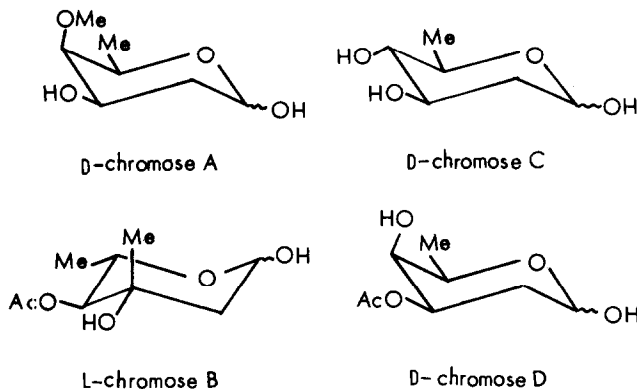


FIG.1.

2,6-Dideoxy-3-C-methyl-4-O-acetyl-L-arabo-hexopyranose (L-chromose B), $C_9H_{16}O_5$, is an oil* with $\nu_{cm^{-1}}^{CHCl_3}$ 1733(OAc), and $[\alpha]_D^{21} = -26^\circ$ (c = 1,1 in H_2O , immediately after preparation), and -24° after 40 min. or longer. Hydrolysis of chromose B with potassium carbonate gave acetic acid and deacetylchromose B, $C_7H_{14}O_4$, m.p. 109° , $[\alpha]_D^{22} = -15^\circ$ (c = 1.0 in H_2O , immediately after preparation), $[\alpha]_D^{22} = -22^\circ$ (after 1 hour and longer).

FIG.2 shows the 100 mc spectrum, including spin-decoupling data of deacetylchromose B dissolved in D_2O containing a small amount of sodium 2,2-dimethyl-2-silapentane-5-sulfonate the

* The analyses of all compounds described in this communication agreed with the molecular formulae although some could not be obtained crystalline.

latter acting as internal reference*.

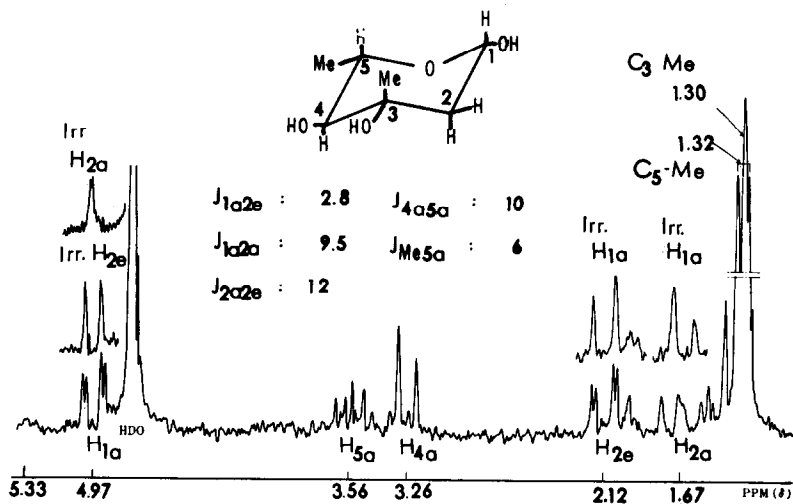


FIG.2 NMR spectrum and decoupled traces of deacetylchromose B in D_2O , ppm from DSS.

The NMR spectrum was measured immediately after dissolving the sample in D_2O . The tall peaks at 1.30 and 1.32 are due to $C_3\text{-Me}$ and $C_5\text{-Me}$ respectively. The latter is coupled with a spin coupling constant of 6 cps to its adjacent proton whose resonance appears at 3.56. H_5 is further coupled to H_4 with a characteristic axial-axial coupling constant of 10 cps.

The nonequivalent protons at C_2 resonate at 1.67 and 2.12, and exhibit a geminal coupling constant of 12 cps. $C_2\text{-H}_a$ and $C_2\text{-H}_e$ are further coupled to $C_1\text{-H}$ with coupling constants of 9.5(axial-axial) and 2.8(axial-equatorial) cps. The peaks

* See Preface II in N.S.Bhacca, D.P.Hollis, L.F.Johnson and E.A.Pier. "NMR Spectra Catalog" Volume 2. Varian Associates 1963.

corresponding to C₁ is located at 4.97 and the width of resonance pattern is 13 cps. However, after three days, the intensity of the quartet at 4.97 is reduced and a triplet (5 cps wide) of corresponding intensity is observed at 5.33. This indicates that axial-axial and axial-equatorial couplings of 9.5 and 2.8 has now been supplemented by a pattern which has axial-equatorial and equatorial-equatorial couplings of 2.5 cps. Thus anomerization takes place at C₁.

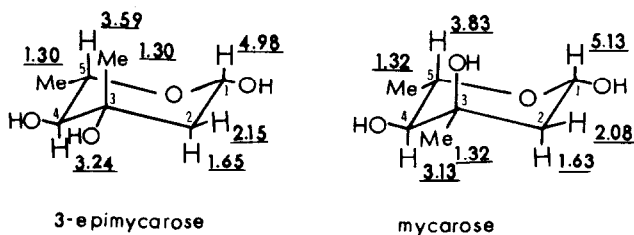
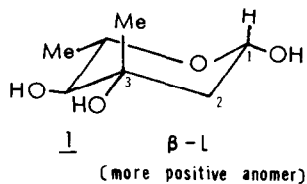


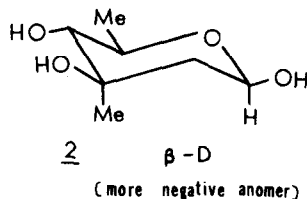
FIG.3 The chemical shifts¹²⁾ (in ppm) calculated using sodium 2,2-dimethyl-2-silapentane-5-sulfonate as internal reference.

The NMR spectrum of deacetylchromose B is very similar to that of mycarose¹²⁾ and 3-epimycarose¹²⁾ which are quite similar to each other (see FIG.2 and 3). Comparisons of FIG.2 and 3 indicate that deacetylchromose B has the 3-epimycarose structure, namely 1, or 2. The negative direction of mutarotation in deacetylchromose B (-15° to -22°) is accompanied by the change in NMR signals of the anomeric C₁-proton (axial to equatorial). Thus deacetylchromose B belongs to the L-series and is the yet unknown¹³⁾ 3-epi-L-mycarose.

Chromose B has an acetate group. The attachment of this function can be readily determined by comparing the NMR spectra



----- mirror

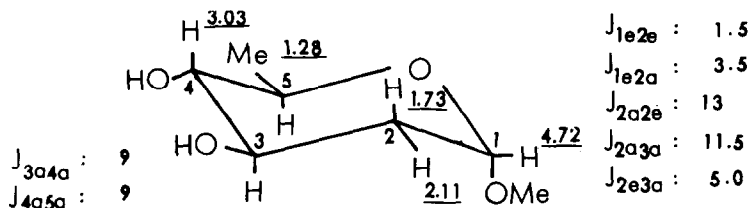


of deacetylchromose B with that of chromose B. The signals corresponding to the proton at C_4 shift down from 3.26 to 4.63 due to the replacement of hydroxyl by a more electronegative acetoxyl group. Therefore the acetate group must be located at C_4 in the equatorial position. This is further substantiated by the fact that L-chromose B itself does not consume periodic acid.

2,6-Dideoxy-D-arabo-hexopyranose(D-chromose C), the sugar eluted from the cellulose powder column was an oil that analysed for $C_6H_{12}O_4$; the $[\alpha]_D^{21}$ value of $+22^\circ$ ($c = 1.4$, in H_2O) remained constant, and thus the oil is already an equilibrium mixture of anomers. Its 60 mc NMR spectrum in D_2O solution showed two quartets of equal intensity at 5.33 ppm ($J: 1.5, 4$ cps) and 4.90 ppm ($J: 2, 10$ cps), which could be assigned to the equatorial and axial C_1 -H, respectively. Periodic oxidation of the oil yielded acetaldehyde and formic acid. Treatment of the oil with 10% HCl-MeOH gave methyl chromoside C, $C_7H_{14}O_4$,

b.p. 1.0 126-129°, $[\alpha]_D^{23} = +87^\circ$ (c = 1.2 in H₂O).

The 60 mc spectrum of methyl chromoside C, dissolved in CDCl₃ containing a trace amount of tetramethylsilane, showed the following signals:



The resonance at 4.72 was assigned to the proton at C₁. It shows two small coupling constants (3.5 and 1.5 cps) to its adjacent methylene group protons. This establishes equatorial configuration for H₁. Since signals at C₄ exhibit two 9 cps couplings, all three protons at C₃, C₄ and C₅ must be axial. This leads to a 2-deoxyrhamnose structure for the sugar. The rotation of the anomeric mixture of 2-deoxy-L-rhamnose¹⁴⁾ is $[\alpha]_D^{14} = -18.2^\circ$, which is opposite that of the present sugar. Accordingly chromose C belongs to the D-series (see FIG.1).

2,6-Dideoxy-3-O-acetyl-D-lyxo-hexopyranose(D-chromose D), C₈H₁₄O₅, was obtained in the form of crystals, m.p. 128°, $[\alpha]_D^{23} = +87^\circ$ (c = 1.5 in H₂O), $\nu_{\text{cm}^{-1}}^{\text{CHCl}_3} 1715(\text{OAc})$, $\delta_{\text{ppm}}^{\text{D}_2\text{O}} 2.13(\text{OAc})$, 1.21(sec-Me).

Chromose D consumes one mole of periodic acid. Upon treatment with potassium carbonate it gives acetic acid and deacetylchromose D, C₆H₁₂O₄, $[\alpha]_D^{20} = +53^\circ$ (c = 1.2 in H₂O); the latter is identical with demethyl D-chromose A (demethylation

with boron trichloride¹⁵⁾). The structure of this sugar is thus also established (see FIG.1).

We are indebted to Professor R.U.Lemieux, Alberta, Canada, for valuable discussions.

REFERENCES

- 1) M.Shibata, K.Tanabe, Y.Hamada, K.Nakazawa, A.Miyake, H.Hitomi, M.Miyamoto and K.Mizuno, J.Antibiotics, Ser. B 13, 1 (1960)
K.Mizuno, J.Antibiotics, Ser. A 16, 22 (1963)
- 2) J.E.Philip and J.R.Schenck, Antibiot. and Chemoth., 3, 1218 (1953)
- 3) H.Nishimura, R.Kimura, K.Tawara, K.Sasaki, N.Nakajima, N.Shimaoka, S.Okamoto, M.Shinohara and J.Isono, J.Antibiotics, Ser. A 10, 205 (1957)
- 4) P.Sensi, A.M.Greco and H.Pagani, Antibiot. and Chemoth., 8, 241 (1958)
- 5) R.M.Gale, M.H.Hoehn and M.H.Cormick, Antibiot. Ann., 1958/1959, 489 (1959)
- 6) H.Schemitz, B.Heinmann, J.Lein and I.R.Hooper, Antibiot. and Chemoth., 10, 740 (1960)
- 7) K.V.Rao, W.R.Cullen and B.O.Sobin, Antibiot. and Chemoth., 12, 182 (1962)
- 8) M.G.Brazhnikova, E.B.Kruglyak, I.N.Kovsharova, N.V.Konstantinova and V.V.Proshlyakova, Antibiotiki, 7, 39 (1962)
- 9) M.Miyamoto, K.Morita, Y.Kawamatsu, S.Noguchi, R.Marumoto, K.Tanaka, S.Tatsuoka, K.Nakanishi, Y.Nakadaira, and N.S.Bhacca, Tetrahedron Letters, this issue.
- 10) M.Miyamoto, K.Morita, Y.Kawamatsu, M.Sasai, A.Nohara, K.Tanaka, S.Tatsuoka, K.Nakanishi, Y.Nakadaira and N.S.Bhacca, Tetrahedron Letters, this issue.
- 11) M.Miyamoto, Y.Kawamatsu, M.Shinohara, Y.Asahi, Y.Nakadaira, H.Kakisawa, K.Nakanishi and N.S.Bhacca, Tetrahedron Letters, 693 (1963)
- 12) W.Hofheinz, H.Griesebach and H.Friebolin, Tetrahedron, 18, 1265 (1962)
- 13) F.Korte, U.Claussen and K.Gohrig, Tetrahedron, 18, 1257 (1962)
D.M.Lemal, P.D.Pacht and R.B.Woodward, Tetrahedron, 18, 1275 (1962)
- 14) B.Iselin and T.Reichstein, Helv. Chim. Acta, 27, 1146 (1944)
- 15) S.Allen, T.G.Bonner, E.J.Bourne and N.M.Saville, Chem. and Ind., 630 (1958)