

A Chiroptical/Chemical Strategy for Configurational Assignments of Acyclic 1,3-Skipped Polyols: Model 1,2,4,6-Tetrols

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In the following we present a microscale strategy for determining the relative and absolute configurations of acyclic 1,3-polyols up to 1,2,4,6-tetrols; extension of the same principle allows one to apply the method to pentols and longer 1,3-polyols.

The 1,3-polyol systems are widely distributed in nature, particularly in the skipped-polyol polyene macrolides which are very important as antifungal and antiviral agents. However, due to difficulties associated with configurational assignments, out of the >200 polyene macrolides isolated, most of which are amorphous, the planar structures of only ca. 40 have been determined.¹ Furthermore, the number of members to which full or partial stereochemistry has been assigned is less than 10:¹ amphotericin B (X-ray),² roxatracin (X-ray),³ mycoticin (degradation and partial synthesis),⁴ nystatin (degradation and spectroscopy),⁵ and lienomycin (degradation and spectroscopy).⁶

Two reiterative methods have been published recently to assign configurations to the skipped polyols. According to Oishi and co-workers,⁷ the steps consisting of lactonization between 1-COOH and 5-OH to 3-hydroxy- δ -lactone \rightarrow NMR \rightarrow dehydration to δ -enolactone \rightarrow NMR give configurations at C-3/C-5/C-7; repetition of these steps after oxidative removal of C-1 to C-4 gives C-7/C-9/C-11 configurations. In the method of Mori et al.,⁸ the difference CD between 1-hydroxy-3,5,...perbenzoate and the corresponding allylic perbenzoate obtained upon 1,2-dehydration gives the absolute configuration at C-3 based on the sign of acyclic allylic benzoate CD;⁹ the starting hydroxy perbenzoate is degraded to 3-hydroxy-5,7,...perbenzoate, and CD measurements are repeated for the C-5 configuration.

The present method employs the bichromophoric¹⁰ exciton chirality method¹¹ which was used in the microscale configura-

tional determination of 1,2-polyols¹² (up to hexols, including 1-aminopentols). A combination of CD spectra of 1-anthroylated *p*-methoxycinnamates of the 1,3-polyols and the diastereoselective spiroketalization reaction of Oku and co-workers¹³ has led to the following procedure.

The four possible diastereomeric tetrols 1–4 with established configurations, synthesized from (*S*)-(-)-malic acid,¹⁴ served as models to develop the procedure. The absolute configuration at C-2 is first determined, thus decreasing the number of possible configurations in an unknown 1,2,4,6-tetrol from eight to four. This is achieved by selective derivatization with two different exciton coupling chromophores (Scheme I): C-1 anthroylation (λ_{max} 252 nm, ϵ 140 000) with 9-anthroylimidazole followed by per-*p*-methoxycinnamoylation (λ_{max} 306 nm, ϵ 23 400). The clear Cotton effect (CE)¹⁵ at 252 nm results primarily from exciton coupling between the 1-Anth/2-Cinn chromophores;^{10a,b,12} the contributions from the 1-Anth/4-Cinn and 1-Anth/6-Cinn couplets to the 252-nm band are much weaker and can be ignored. Thus a positive 252-nm CE in the 1-Anth-2,4,6-tricinnamate derivative is diagnostic for an *S*-configuration at C-2, its sign being independent of configurations at remaining chiral centers, and *vice versa*.

A strong bisignate CD around 300 nm, namely, a strong positive CE at 280 nm and negative CE at 320 nm (negative exciton coupling) is characteristic of two of the four possible structures, 2,4-*syn*-4,6-*anti* (1a) and 2,4-*anti*-4,6-*syn* (2a) (Figure 1a). This group will be denoted as "S" (for strong). An acyclic *anti*-1,3-dibenzoate adopts a planar zigzag form in its most stable conformer and exhibits a typical CD exciton couplet corresponding to the sign of the screw sense between the two gauche oriented chromophores.¹⁶ However, in the most stable conformer of the *syn* analog, which is also zigzag, the transition moments of the acylate chromophores are parallel and hence show negligible coupling.¹⁶ Thus, in 1a and 2a, the strong couplet arises from the coupling of the 4,6- and 2,4-*anti*-cinnamates, respectively; the 2,4-cinnamates in 1a and 4,6-cinnamates in 2a are *syn* and hence do not couple. In contrast, a weak CD in the 280–320-nm region is characteristic for configurations 2,4-*syn*-4,6-*syn* (3a) and 2,4-*anti*-4,6-*anti* (4a), denoted by group "W" (for weak) (Figure 1b). The coupling is weak in 3a because the 2,4,6-cinnamates are all *syn*, whereas in 4a the 2,4-*anti* and 4,6-*anti* contributions cancel out.

The differentiation between 1 and 2 of group S and 3 and 4 of group W becomes possible by using *l*-menthone, a highly diastereoselective reagent. According to Oku and co-workers,¹³ *l*-menthone selectively spiroketalizes 1,3-*syn*-diols at -78 °C while leaving the 1,3-*anti*-diols unchanged. Both tetrols 1 and 2 reacted with *l*-menthone to give spiroketal derivatives 1b and 2b in agreement with their 2,4- and 4,6-*syn* configurations, respectively (Scheme I). With 1b, cinnamoylation followed by deketalization gave 1c, which exhibited only a weak CD throughout the region 220–360 nm because of the remoteness of the 1-Anth and 6-Cinn chromophores; the same two-step reaction applied to 2b gave 2c, which shows the strong positive 252-nm CE (Figure 1c). This differentiates 1 and 2 in group S. Treatment of tetrols 3 and 4 (group W) with *l*-menthone under the same conditions readily distinguished the two. Namely, 3 yielded two spiroketals 3b and

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(1) Omura, S.; Tanaka, H. In *Macrolide Antibiotics: Chemistry, Biology and Practice*; Omura, S., Ed.; Academic Press: New York, 1984; pp 351–401.

(2) Mechliniski, W.; Schaffner, C. P.; Ganis, P.; Avitabile, G. *Tetrahedron Lett.* 1970, 3873–3876.

(3) Maehr, H.; Yang, R.; Hong, L.-N.; Liu, C.-M.; Hatada, M. H.; Todaro, L. J. *J. Org. Chem.* 1989, 54, 3816–3819.

(4) (a) Schreiber, S. L.; Goulet, M. T. *Tetrahedron Lett.* 1987, 28, 6001–6004. (b) Schreiber, S. L.; Goulet, M. T.; Sammakia, T. *Tetrahedron Lett.* 1987, 28, 6005–6008. (c) Schreiber, S. L.; Goulet, M. T. *J. Am. Chem. Soc.* 1987, 109, 8120–8122.

(5) (a) Lancelin, J. M.; Beau, J. M. *Tetrahedron Lett.* 1989, 30, 4521–4524. (b) Prandi, J.; Beau, J. M. *Tetrahedron Lett.* 1989, 30, 4517–4520. (c) Nicolaou, K. C.; Ahn, K. H. *Tetrahedron Lett.* 1989, 30, 1217–1220.

(6) Pawlak, J.; Nakanishi, K.; Iwashita, T.; Borowski, E. *J. Org. Chem.* 1987, 52, 2896–2901.

(7) (a) Nakata, T.; Noriaki, H.; Nakashima, K.; Oishi, T. *Chem. Pharm. Bull.* 1987, 35, 4355–4358. (b) Oishi, T. *Pure Appl. Chem.* 1989, 61, 427–430.

(8) Mori, Y.; Kohchi, Y.; Suzuki, M.; Furukawa, H. *J. Am. Chem. Soc.* 1992, 114, 3557–3559.

(9) Gonnella, N. C.; Nakanishi, K.; Martin, V. S.; Sharpless, K. B. *J. Am. Chem. Soc.* 1982, 104, 3775–3776.

(10) (a) Wiesler, W. T.; Nakanishi, K. *J. Am. Chem. Soc.* 1989, 111, 9205–9213. (b) Wiesler, W. T.; Nakanishi, K. *J. Am. Chem. Soc.* 1990, 112, 5574–5583. (c) Wiesler, W. T.; Berova, N.; Ojika, M.; Meyers, H. V.; Chang, M.; Zhou, P.; Lo, L.-C.; Niwa, M.; Takeda, R.; Nakanishi, K. *Helv. Chim. Acta* 1990, 73, 509–551.

(11) Harada, N.; Nakanishi, K. *Circular Dichroic Spectroscopy—Exciton Coupling in Organic Stereochemistry*; University Science Books: Mill Valley, CA, 1983.

(12) (a) Zhou, P.; Berova, N.; Nakanishi, K.; Rohmer, M. *J. Chem. Soc., Chem. Commun.* 1991, 256–258. (b) Zhou, P.; Berova, N.; Nakanishi, K.; Knani, M.; Rohmer, M. *J. Am. Chem. Soc.* 1991, 113, 4040–4042. (c) Zhou, P.; Berova, N.; Wiesler, W. T.; Nakanishi, K. *Tetrahedron*, in press.

(13) (a) Harada, T.; Wada, I.; Oku, A. *J. Org. Chem.* 1989, 54, 2599–2605. (b) Harada, T.; Kurokawa, H.; Kagamihara, Y.; Tanaka, S.; Inoue, A.; Oku, A. *J. Org. Chem.* 1992, 57, 1412–1421.

(14) Manuscript in preparation.

(15) CD and UV spectra were recorded in acetonitrile on JASCO J-720 and Perkin-Elmer Lambda 4B UV/VIS spectrometers, respectively. In-house-developed software was employed for CD and UV data manipulation.

(16) Harada, N.; Saito, A.; Ono, H.; Gawronski, J.; Gawronska, K.; Sugioka, T.; Uda, H.; Kuriki, T. *J. Am. Chem. Soc.* 1991, 113, 3842–3850.

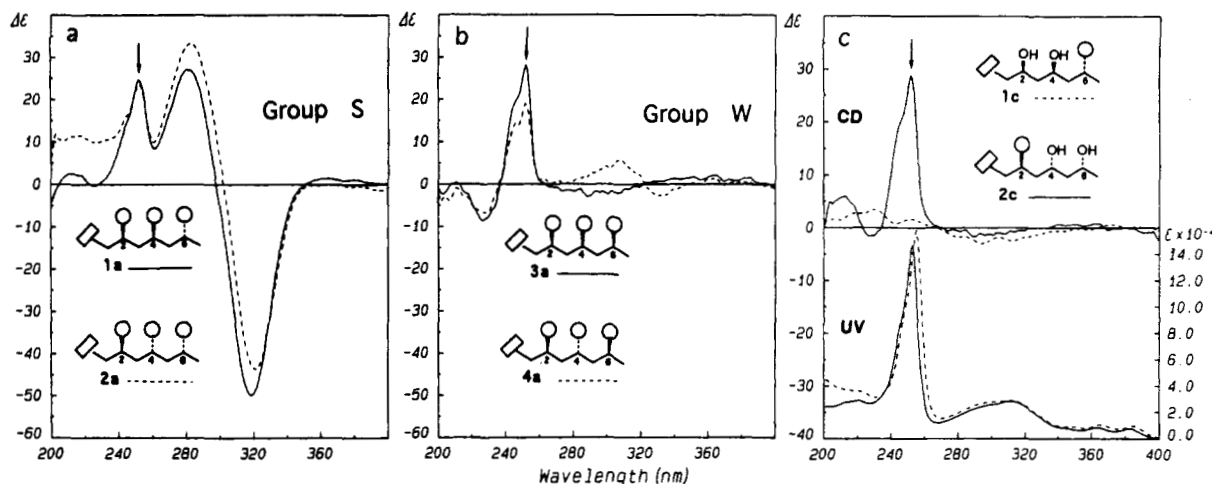
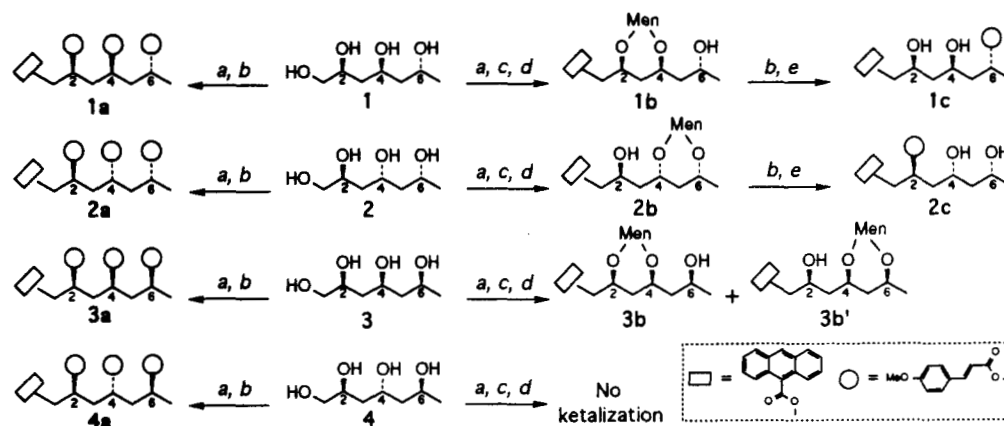


Figure 1. CD (λ_{ext} nm/ $\Delta\epsilon$) and UV (λ_{max} nm/ ϵ) spectra of derivatized tetrols in acetonitrile. (a) CD of **1a**: 252 ($\Delta\epsilon$ +24.7); 281 (+27.2); 319 (−49.8). CD of **2a**: 252 (+23.8); 283 (+33.3); 321 (−43.5). (b) CD of **3a**: 252 (+28.2); 289 (−3.0); 363 (+2.2). CD of **4a**: 252 (+19.0); 307 (+5.7); 330 (−2.7). (c) CD of **1c**: 252 (+1.6); 294 (−2.9); 311 (−2.4). CD of **2c**: 252 (+28.8); 296 (−1.4). UV of **1c**: 254 (ϵ 158 000); 309 (29 000). **2c**: 252 (147 000); 310 (28 000).

Scheme I. Chromophoric Derivatization and Ketalization of Tetrols 1–4^a



^a (a) Anthroylation; (b) *p*-methoxycinnamoylation; (c) TMSCl, Et₃N; (d) *l*-menthone, TMSOTf, −78 °C; (e) MeOH, H⁺.

3b', whereas no reaction occurred with tetrol **4** having 2,4-*anti*-4,6-*anti*-configurations.

A typical procedure is as follows; although it was not attempted to minimize the amount of sample, the scale can readily be reduced to ca. one-tenth.¹⁷

(a) Peracylation of Tetrol, 1 → 1a. Treatment of tetrol **1** (10 mg) with 9-anthroyltetrazole^{12b} (2 equiv) and DBU (2 equiv) in CH₂Cl₂ at room temperature (rt) for 3 h followed by silica gel chromatography gave the 1-anthroate,¹⁷ 53% yield; further reaction with *p*-methoxycinnamoylimidazole^{12b} (5 equiv) and DBU (4 equiv) in CH₃CN at rt for 4 h furnished **1a** in 85% yield.

(b) Spiroketalization, 1 → 1b. The tris-TMS derivative (11 mg) of the 1-anthroate of **1**, prepared by stirring of the anthroate with TMSCl (6 equiv) and excess Et₃N in dry CH₂Cl₂ (81%), was condensed with freshly distilled *l*-menthone (2 equiv) in THF at −78 °C for 14 h in the presence of TMSOTf (0.5 equiv). After the reaction was quenched by addition of pyridine at −40 °C, methanol was added at 0 °C and the reaction mixture was stirred for 1 h, 0 °C, to yield **1b** (92%) after silica gel chromatography.

(c) Cinnamoylation and Deketalization, 1b → 1c. Cinnamoylation of spiroketal **1b** (9 mg) under conditions described above with 1.5 equiv of *p*-methoxycinnamoylimidazole^{12b} and 1.5 equiv of DBU furnished monocinnamoyl spiroketal (77%). Depro-

tection of **1b** in MeOH with a trace of CH₃COCl, rt, 2 h, yielded monocinnamoylated derivatives **1c** (65%).

(d) Separation of Ketals 3b and 3b'. The two were readily separable on silica gel column, **3b'** (40%) and **3b** (10%) being eluted, respectively, by 14% and 36% ether in hexane.

Acyclic 1,3-polyols with the α -glycol-terminal-CH₂-CHOH-CH₂OH are readily obtained from most polyene macrolides upon hydrolysis/reduction of the terminal -COOH coupled with ozonolysis. In the present method, the C-2 absolute configuration is determined from the 252-nm CD of the 1-anthroate per-*p*-methoxycinnamate; furthermore, depending on the *syn* or *anti* arrangements of the cinnamates, the 260–340-nm region shows a strong couplet or no couplet. Final determination of configurations is achieved by a spiroketalization step and, if necessary, cinnamoylation and deketalization. An advantage of the present method is that, unlike the method developed for 1,2-polyols,^{10a,b,12} reference CD curves are not necessary. This communication is restricted to a description of the strategic principle rather than an application to a real case since none of the macrolides with established structures^{1–5} possesses the tetrol moiety discussed above. However, extension of the strategy to acyclic pentols and mixed 1,2/1,3-polyol systems will be reported shortly with applications to real cases.

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(17) The purity and identity of all samples were checked by MS and ¹H NMR.