



Canonical computations of cerebral cortex

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The idea that there is a fundamental cortical circuit that performs canonical computations remains compelling though far from proven. Here we review evidence for two canonical operations within sensory cortical areas: a feedforward computation of selectivity; and a recurrent computation of gain in which, given sufficiently strong external input, perhaps from multiple sources, intracortical input largely, but not completely, cancels this external input. This operation leads to many characteristic cortical nonlinearities in integrating multiple stimuli. The cortical computation must combine such local processing with hierarchical processing across areas. We point to important changes in moving from sensory cortex to motor and frontal cortex and the possibility of substantial differences between cortex in rodents vs. species with columnar organization of selectivity.

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A cortical computational unit?

The cerebral cortex performs a wide range of cognitive tasks in mammals — sensory, motor, and everything in between. Yet it processes these diverse tasks with what appears to be a remarkably uniform, primarily six-layer architecture, albeit with significant differences in details across species and cortical areas [1,2,3,4–10,11,12–14]. Dense connectivity is generally restricted to horizontal extents (parallel to the layers) of a few hundred microns [e.g. 15]. This has long suggested the idea that a piece of six-layer cortex with a surface area on the order of a square millimeter constitutes a fundamental cortical ‘processing unit’ [e.g. 16,17]. The cortex varies in surface area by a factor of 10000 across a set of 37 mammalian species, while thickness (the distance across the layers) varies only by a

factor of 10 over the same species [18], suggesting that the most salient evolutionary change in cortex has been enormous multiplication of the number of ‘units’ [e.g. 14].

The nature of this six-layer computation remains unclear. The classic picture [e.g. 19,20] is that feedforward input to a given area, which either comes from thalamus or from ‘lower’ cortical areas, comes dominantly into layer 4 (L4); L4 projects strongly to layers 2/3 (L2/3); L2/3 provides feedforward input to L4 of ‘higher’ cortical areas, and also projects to L5; L5 provides the only output from cortex other than feedback to thalamus, and also projects to L6; L6 projects up to layers 2 through 4, completing a loop through the layers; and L6, and more generally layers other than L4, provide feedback to areas projecting feedforward input to the given area. Layer 1, which contains inhibitory neurons and dendrites of excitatory neurons from lower layers, is thought to integrate various forms of modulatory input from higher cortical and thalamic areas. Several findings challenge this picture [21,22,23], including notably that in mouse primary somatosensory cortex (S1), L5 response to whisker movement is directly driven by thalamic inputs independently of layers 2–4 [21]. The picture is being filled out by detailed characterization of pathways involving subtypes of excitatory [e.g. 11,15,24,25] and inhibitory [e.g. 15,26,27,28,29,30,31,32–35] neurons in mice.

The idea of a cortical computational unit is sketchy and not without controversy, but it motivates the idea that there are fundamental or ‘canonical’ computations performed by cortical circuits [36,37] that, when fed appropriate inputs and arranged in appropriate hierarchies, might go some ways towards accounting for properties of mammalian intelligence. This idea was at least loosely supported by findings that, when visual inputs were fed to primary auditory cortex (A1), that cortex developed many functional properties normally seen in visual cortex [38]. Some support also comes from studies of artificial neural networks arranged in a hierarchy of layers, far removed from biological details but motivated by these basic ideas. These ‘deep’ networks have recently made great breakthroughs in training machine intelligence [39]. Such networks trained to identify objects in visual images are designed so that, within a given layer, they perform the same operations at each point in visual space. The properties found across layers in these trained networks resemble in some detail those seen across the hierarchically arranged cortical areas in the ventral visual stream, which is thought to perform object recognition [40]. These are hardly proofs, but at least show that hierarchical repetition of basic distributed processing units can both achieve

impressive aspects of intelligence and do so in ways broadly consistent with cortical representations.

Invariant recognition: hierarchical organization and local processing

Neurons in primary sensory cortices respond to stimuli in localized regions of the sensory periphery (the neuron's 'receptive field'). A critical computational task of sensory cortex is to integrate the discrete, localized information carried by individual neurons into unified percepts of objects in the world. Objects are recognized in a manner invariant to almost all particular details of the incoming stimulus, for example, invariant to rotations, occlusions, or changes of luminance, position, distance, apparent size, or contexts — yet of course dependent on those details [41]. This invariant recognition has, until the recent advent of 'deep' neural networks [39[•]], been difficult for machines to replicate, as it is not easily captured by rules. A similar interaction between high-level objects and varying low-level implementations is seen in motor planning: for example, an intention to point to an object might be instantiated with various fingers of either hand and various arm configurations, or a foot, or a laser pointer. We might speculate that this difficult-to-characterize, dependent-yet-independent relationship between cognitive elements at higher and lower levels reflects something general about many cognitive processes and thus about cortical computation.

How is invariant recognition achieved? While we do not know, we can recognize two elements of the computation from studies of the ventral stream in vision. First, there is a hierarchy of cortical areas, in which neurons in 'higher', more anterior areas of the hierarchy are sensitive to the presence of more complex aspects of objects across larger regions of sensory space and with greater ability to recognize these structures in ways invariant to position or pose [19,41]. Action control may similarly be hierarchically organized from posterior to anterior across areas of motor and frontal cortex [42,43]. Second, within any given area, neurons are selective for relevant features of the stimulus, but responses are modulated by other influences. This modulation tends to modify the gain rather than selectivity of responses, whether evoked by additional stimuli within the receptive field [37], context or surrounding stimuli outside the receptive field [44], or top-down influences such as attention [45] (however, attention to stimulus features can boost the gain of responses to the attended features and thus alter a neuron's stimulus selectivity [46]). The role of this dynamic gain modulation in the computations of invariant recognition is not currently understood.

Canonical local computations

Here we review ideas for these two canonical computations within a sensory cortical area: computation of selectivity and computation of gain.

Feedforward computation of selectivity

In L4 of primary sensory cortex in several modalities and species, the selectivity of neuronal responses (the relative response strength across different stimuli) is primarily established by the pattern of feedforward connections the neurons receive (thalamic input and perhaps thalamic-driven inhibition). Intracortical recurrent input primarily determines the gain (the absolute response strength) but not the selectivity of responses (although a role of cortical recurrence in selectivity may arise in adaptation or perceptual learning [e.g. 47] or blockade of cortical areas lateral to a site [48]).

This was first established in cat V1, in which cortex, and thus intracortical input, was silenced either by cooling [49] or by electrical shock (which evoked massive inhibition) [50], leaving only thalamic input. These manipulations did not change the tuning for stimulus orientation of the membrane potential in recorded neurons, but modified the gain of response. The estimated ratio of voltage response for thalamic input alone to that with cortex intact averaged about 1/3 to 1/2, with ratios broadly distributed between 0 and 1 [50,51]. Similar results were obtained for measurements of excitatory conductance in mouse V1 [52^{••},53^{••}] and A1 [54[•]], using optogenetic methods to silence cortex by activating parvalbumin-expressing (PV) inhibitory neurons. In rat S1 representing whiskers, the tuning of L4 neurons for direction of whisker movement corresponds closely to the directional tuning of the thalamic inputs they receive [55,56]. In cat V1, many other tuning properties can also be explained from the tuning of feedforward input [57,58], as suggested by earlier models [59–62].

The role of feedforward inhibitory input in determining tuning remains unclear. Assuming that orientation selectivity in a L4 V1 neuron is determined by the spatial arrangement of thalamic afferents [52^{••},63,64], rather than being inherited from a set of orientation-tuned afferents with similar preferred orientations (the latter has been argued as possible in mouse, e.g. [65–67]), the mean input to a L4 cell in response to a drifting luminance grating is not tuned for grating orientation. Rather, the modulation of the input about that mean is orientation tuned, with strong peaks and troughs of input at the preferred orientation but little modulation at orthogonal orientations [61,68,69]. This untuned mean and tuned modulation was observed for thalamic excitatory conductance in mice [52^{••}], arguing against inheritance of orientation tuning from thalamic afferents, and has not otherwise been examined for thalamic input (other studies measured only the modulation, not the mean, of response [49] or used non-periodic stimuli [50,53^{••}]). The untuned mean thalamic input grows with stimulus contrast, yet the orientation orthogonal to the preferred (the 'null' orientation) typically does not evoke spiking responses [70]. Why does the increasing thalamic input

evoked by increasing null-orientation contrast not drive spiking? An untuned component of feedforward inhibition was proposed as one answer [61,68,69,71]. In a series of elegant studies, Ferster and colleagues demonstrated that this can instead be explained by a decrease in voltage variability with increasing stimulus contrast: as contrast increases, the null-oriented stimulus evokes increasing depolarization, but the decreasing variability keeps unchanged the probability of a spike-evoking suprathreshold voltage fluctuation [51,72,73]. However, inhibition could play a role in reducing the size of the depolarization. The same authors have also argued that inhibition is not needed to explain other cat V1 tuning properties [57]. Other studies in mice have suggested roles of inhibition in sharpening selectivity [74–80]. In L4 of rodent S1 and A1, feedforward inhibition cuts off excitation and so sets a temporal window of a few milliseconds for the arrival of effective thalamic excitatory inputs [81–84].

These results largely describe selectivity in L4. In species with columnar organization of selectivity (similar selectivity across the layers at a given cortical point, further discussed below), selectivity in other layers may be inherited in a fairly straightforward way from L4, so accounting for the origin of selectivity in L4 may suffice. However, in rodents, which lack columnar organization, intracortical recurrent input could play a greater role in deriving selectivity outside L4 (e.g. see [85*,86*,87*] for different versions of one proposal).

Computation of gain

How is gain modulated in the local cortical circuit? In recent years studies of specific cell types in mice have suggested a number of specific circuits that lower or raise gain through inhibition or disinhibition respectively [26*,27*,28*,29*,30*,31*,32–34]. However, these circuits alone do not explain nonlinearities of modulation. For example, responses in visual cortical areas typically grow with increasing stimulus size up until a critical size (the ‘summation field’ size), with further size increases suppressing responses (‘surround suppression’); but the same surround stimulus may facilitate response to a weak center stimulus yet suppress response to a stronger stimulus [88*,89–92] and the summation field size shrinks with increasing stimulus contrast [44,93–100]. Similarly, responses to two stimuli in the receptive field may add linearly or perhaps supralinearly when they are weak but sublinearly when they are stronger ([101]; T. Ohshiro, D.E. Angelaki and G.C. DeAngelis, abstract in *Soc Neurosci Abstr* 2013, Program No. 360.19). Recently a very general mechanism of gain modulation [102**,103**] has been proposed that can explain these nonlinearities and may be instantiated through a variety of specific circuits, including those recently characterized.

The mechanism is based on the input/output (I/O) functions of cortical neurons, which specify firing rate as a

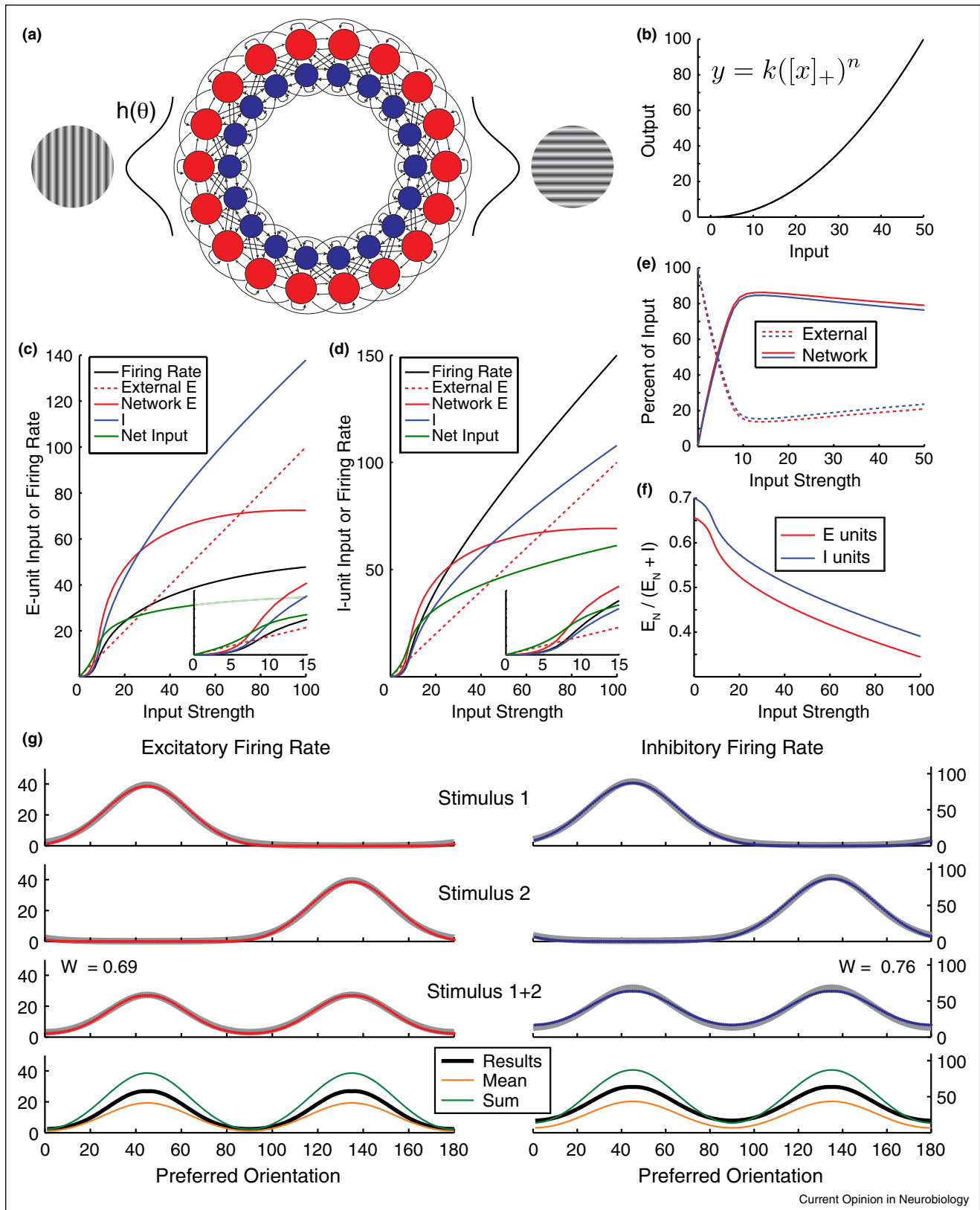
function of mean input, being supralinear (meaning that the slope of the I/O function is monotonically increasing, Figure 1b), as judged by a supralinear relationship between mean membrane voltage and firing rate in 30 ms bins in cat V1 [104]. A supralinear I/O function arises if the mean input neurons receive is subthreshold, so that they fire on the fluctuations in their input rather than on integration of the mean input [105,106]. Importantly, over the full dynamic range of V1 visual responses, the I/O function stays supralinear, not saturating even for the strongest visual stimuli [104].

The supralinear I/O function causes effective synaptic strengths — the change in output firing rate induced by a change in input rate — to increase with increasing postsynaptic activity. The effective strength is the product of the synaptic strength and the gain of the postsynaptic cell — the change in its rate induced by a given change in synaptic input — which is precisely the slope of the I/O function.

This change in gain leads to a change in operating regime of the circuit, as follows. For weak activation, the weak effective synaptic strengths cause the input to neurons to be dominated by external drive (Figure 1c,d, insets) (synapses both from external and network neurons are weak, but network drive involves two or more weak synapses — external drive moves neuronal rates which alters network drive — and so is weak relative to external drive). Thus, neurons are only weakly coupled. Since the I/O function is supralinear, and neurons are roughly responding directly to their external input, responses to multiple stimuli add supralinearly: the response to two simultaneously presented stimuli exceeds the sum of the responses to the individual stimuli.

With increasing activation strength, effective synaptic strengths grow and network drive comes to dominate over external (Figure 1c–e). In particular, connections between excitatory neurons become strong enough that the excitatory subnetwork becomes unstable by itself — subject to runaway activity — but feedback inhibition, if sufficiently strong, keeps the network stable. To stabilize against the increasing effective excitatory strength, however, the network input must ‘conspire’ to largely cancel the external input received by cells, leaving a residual net input (Figure 1c,d, green) that grows much more slowly than the growth of the external input (Figure 1c,d, dashed red) [102**]. Thus, when two stimuli are simultaneously presented, much of the external input will be cancelled and the response will be less than the sum of the responses to the two stimuli presented individually: responses will sum sublinearly (Figure 1g). This is a key property commonly referred to as ‘normalization’ — the response can be described as if it were divided (‘normalized’) by some function of the summed strengths of all stimuli presented, in a form that makes summation sublinear — that has been

Figure 1



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postulated to be a fundamental ‘canonical’ computation underlying many aspects of cortical processing [37].

This simple mechanism, instantiated in simple circuit models, can explain the nonlinear changes in surround suppression and normalization mentioned earlier (it is possible that spontaneous activity is already sufficiently strong to eliminate the supralinear regime, so that weak activation leads to linear rather than supralinear summation). With increasing input drive, the cancellation becomes stronger (Figure 1c,d) and the network input becomes more dominated by inhibition (Figure 1f). This increasingly strong stabilization of the network can potentially explain (D.B. Rubin and K.D. Miller, abstract in *Soc Neurosci Abstr* 2011, Program No. 428.10) the suppression of variability observed in multiple cortical areas with introduction of a stimulus or motor plan [107] and the suppression of correlated variability associated with attention [108,109]. The network can also potentially account for multiple aspects of attentional modulation of neuronal responses (K.D. Miller and D.B. Rubin, abstract in *Soc Neurosci Abstr* 2011, Program No. 428.09), in line with multiple studies showing that attentional increase of input to a normalizing circuit can explain this modulation [45,110–112]. The cancellation of external input is essentially the same mechanism as in the previously proposed ‘balanced network’ [113], but in a very different parameter regime in which inputs need not be large relative to threshold, balance is loose rather than tight, and nonlinear behavior like that seen in cortex arises (whereas only linear behavior is possible in the balanced network) [discussed in 102**].

The most direct tests of this circuit mechanism would be to measure the corresponding currents, for example, regarding each layer as an independent network, but there is currently no way to disentangle a layer’s external vs. intrinsic inputs except in L4, where external inputs come from thalamus. Recurrent input has been shown to be increasingly inhibition-dominated, as predicted, in optogenetic activation of upper layers of rodent S1 [114], but the predicted network effect must be disentangled from effects due to facilitation of excitatory synapses to somatostatin-expressing (SOM) inhibitory neurons [115,116].

A key set of predictions involve the network being an inhibition-stabilized-network (ISN): stabilized by inhibition despite the excitatory subnetwork being unstable. An ISN shows a ‘paradoxical’ property: increased (decreased) drive to inhibitory cells causes both excitatory (E) and, paradoxically, inhibitory (I) cells to lower (raise) their firing rates in the new steady state [117]. This is one example of ‘balanced amplification’ [118], in which, when strong feedback inhibition balances strong excitation, small biases of input towards I (E) cells causes strong decreases (increases) in both E and I firing rates. Given the multiple subtypes of I neurons and complicated circuitry between them [e.g. 15,35], it is possible that different subtypes of I neurons will have opposite responses, but the robust prediction [discussed in 103**] is that, in an ISN, the net inhibition received by E cells will go down (up) when their firing is suppressed (facilitated) by increases (decreases) in net external drive to inhibitory neurons. The inhibition received by neurons indeed is paradoxically decreased when they are surround suppressed in cats [119] (but see [120], discussed in [103**]). In mice, with increasing stimulus size that suppresses E cell firing, the firing rates of PV I neurons decrease while those of SOM I neurons either increase [121] or decrease [122]; the latter suggests an ISN, the former is equivocal. In mouse A1, locomotion activates layer 1 inhibitory neurons, causing E cells in layers 2/3 to be suppressed while receiving decreased inhibition, suggesting an ISN [123]. Tests of the transition to an ISN with increasing stimulus strength are possible by manipulating both sensory stimulus strength and optogenetic stimulus spatial frequency [discussed in 103**, Fig. 8].

Challenges for understanding canonical cortical computations

The unknowns and challenges greatly outweigh our current understandings. Obviously we need to better understand computations within the local circuit, including those specific for cell type and/or layer; the hierarchical computations across cortical areas, including the role in the latter of feedback [e.g. 19,124–128] as well as feedforward connections; and the role of structural variations between cortical areas and across species [e.g. 1,2,3*,4–9]. Additional challenges include:

(Figure 1 Legend) Modulation of gain in a simple model circuit. Reproduced from [103**]. **(a)**. 180 excitatory (E, red) and inhibitory (I, blue) units, with coordinates θ on a ring thought of as preferred orientations. Lines schematize connections. A stimulus grating evokes input $ch(\theta)$ equally to E and I units, with $h(\theta)$ a unit-height Gaussian centered at the stimulus orientation with standard deviation $\sigma_{FF} = 30^\circ$. We consider gratings at 45° , 135° or both simultaneously. **(b)**. The power-law input/output function: $k = 0.04$, $n = 2.0$. y represents a cell’s firing rate, x its net input, $[x]_+$ is x with negative values set to zero. **(c–f)** use a single-grating stimulus. **(c,d)**. For E (c) and I (d) units at stimulus center: with increasing external input strength c (x -axis; dashed lines), network input (E, red; I, blue) transitions from weak to dominating (insets) and substantially cancels external input, so net input (green) grows slowly. Firing rates (black) are proportional to net input squared. **(e,f)**. Summing input received by all E (red) or I (blue) units: with increasing c , input to network (sum of absolute values of E and I input) is increasingly network-driven (**E**; dashed, external input; solid, network input) and network input is increasingly inhibitory (**f**): $E_N/(E_N + I)$ where I , E_N are inhibitory and network excitatory input respectively). **(g)**. Sublinear response summation for multiple stimuli. Top two rows: responses of E (left, red) and I (right, blue) units across network to 45° (top) and 135° (2nd row) stimulus, $c = 50$. 3rd row: responses to both stimuli presented simultaneously. 4th row: responses from 3rd row (black) vs. mean (orange) and linear sum (green) of responses to the two individual stimuli. Gray: rows 1–2, best fit of Gaussians to response; row 3, best fit of weight W (indicated) times sum of fits from rows 1–2.

Understanding canonical computations of motor/frontal cortex. An important possible difference between cortical areas is that motor and frontal areas, anterior to the central sulcus, appear to generate their own activity without an obvious external driving stimulus, whereas sensory areas, posterior to the central sulcus, appear to remain at a relatively low ‘spontaneous’ level of activity in the absence of driving sensory stimuli. This distinction is far from clear: we do not well understand the inputs anterior cortical areas receive and their role in the activity generated there; and sensory areas can generate task-related activity in the absence of a sensory stimulus, but this may require top-down input from more anterior areas (discussed for monkey area LIP in [129]).

If this difference exists, a reasonable hypothesis is that it arises from the increasingly strong connectivity between excitatory neurons seen with posterior-to-anterior movement from primary sensory areas to ‘higher’ sensory areas to motor areas and frontal cortex [130]. In theoretical models, such increasing excitatory connectivity can generate longer timescales for neural activity changes [131] until reaching a critical strength that allows independent activity generation (discussed e.g. in [118,132]; this also depends on inhibitory connectivity which, if properly arranged, can cancel these effects of strong excitation, e.g. [118]). Indeed, increasingly slow changes in neuronal activity are seen from posterior to anterior cortex [133,134*], although the contributions of increasingly slow variations of external inputs vs. intrinsic circuit properties remain undetermined. Whereas our accounts of sensory canonical computations focused on the structure of a specific response to a specific stimulus, connectivity that allows independent activity generation allows more complicated behaviors such as multiple possible response states depending on history or chaotic wandering of firing rates [e.g. 135–139], requiring expanded ideas of canonical computations.

Columnar vs. non-columnar species. Most mammalian species show ‘columnar organization’ of selectivity, in which multiple aspects of neuronal selectivity (e.g. orientation preference in primary visual cortex (V1)) are invariant among nearby cells and across the layers at a given cortical position and vary periodically with horizontal movement across cortex over distances on order of 1 mm [e.g. 16,17]. In rodents and lagomorphs, however, only the topographic position in the sensory periphery, which is common to all thalamic afferents to a given cortical position, is invariant locally or across the layers, while properties such as orientation selectivity that must be derived by selection of specific subsets of thalamic afferents [e.g. 64] appear uncorrelated from cell to cell [e.g. 140]. While this has been best characterized in V1, long-range intrinsic horizontal connectivity in layers 2/3 is patchy or stripe-like, with periods on order 1 mm, across multiple areas of primate and cat cortex from sensory to

prefrontal [141], suggesting linking of periodically spaced cells with related response features, whereas such long-range connections are diffuse without obvious periodicity in areas studied thus far in rodents [140–142]. The reasons for these differences remain unknown; theoretically, lack of columnar organization is likely to arise from local interactions during development that are heterogeneous or suppressive rather than facilitatory [reviewed in 143*]. Other notable differences exist between rodent and primate cortex (e.g. [3*,6,9] and discussion of [95]), including upper layers being thinner and containing fewer neurons, relative to deep layers, in rodents [3*,6]. It remains to be determined whether rodents differ slightly or profoundly from columnar species in their cortical organization and computations.

Conclusions

The rapid growth of cortical surface area in mammalian evolution suggests the development of a ‘canonical’ computational unit that, deployed in great numbers both within cortical areas and hierarchically across cortical areas, along with some specializations across species and areas, allowed the evolution of mammalian intelligence. We have outlined here some current ideas as to some elements of this computational unit. We can only hope that in coming decades our understandings will grow more definite and precise.

Conflict of interest

Nothing declared.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Barbas H: **General cortical and special prefrontal connections: principles from structure to function.** *Annu Rev Neurosci* 2015, **38**:269-289.
 2. Bernard A, Lubbers LS, Tanis KQ, Luo R, Podtelezchnikov AA, Finney EM, McWhorter MM, Serikawa K, Lemon T, Morgan R *et al.*: **Transcriptional architecture of the primate neocortex.** *Neuron* 2012, **73**:1083-1099.
 3. Charvet CJ, Cahalane DJ, Finlay BL: **Systematic, cross-cortex variation in neuron numbers in rodents and primates.** *Cereb Cortex* 2015, **25**:147-160.
- Demonstrates an increase in the number of neurons per cortical surface area with movement anterior-to-posterior across cortex. This increase is primarily due to the number of neurons in layers 2–4, and is much larger in primates than in rodents.
4. DeFelipe J, Alonso-Nanclares L, Arellano JI: **Microstructure of the neocortex: comparative aspects.** *J Neurocytol* 2002, **31**:299-316.
 5. Herculano-Houzel S, Collins CE, Wong P, Kaas JH, Lent R: **The basic nonuniformity of the cerebral cortex.** *Proc Natl Acad Sci U S A* 2008, **105**:12593-12598.

6. Hutsler JJ, Lee DG, Porter KK: **Comparative analysis of cortical layering and supragranular layer enlargement in rodent carnivore and primate species.** *Brain Res* 2005, **1052**:71-81.
7. Krubitzer L: **In search of a unifying theory of complex brain evolution.** *Ann N Y Acad Sci* 2009, **1156**:44-67.
8. Srinivasan S, Carlo CN, Stevens CF: **Predicting visual acuity from the structure of visual cortex.** *Proc Natl Acad Sci U S A* 2015, **112**:7815-7820.
9. Ventura-Antunes L, Mota B, Herculano-Houzel S: **Different scaling of white matter volume, cortical connectivity, and gyrification across rodent and primate brains.** *Front Neuroanat* 2013, **7**:3.
10. Carlo CN, Stevens CF: **Structural uniformity of neocortex, revisited.** *Proc Natl Acad Sci U S A* 2013, **110**:1488-1493.
11. Harris KD, Shepherd GM: **The neocortical circuit: themes and variations.** *Nat Neurosci* 2015, **18**:170-181.
Good review of the different genetically and molecularly defined subclasses of excitatory and inhibitory neurons and their intra-area and inter-area patterns of connectivity.
12. Jones EG: **Microcolumns in the cerebral cortex.** *Proc Natl Acad Sci U S A* 2000, **97**:5019-5021.
13. Molnar Z, Kaas JH, de Carlos JA, Hevner RF, Lein E, Nemeč P: **Evolution and development of the mammalian cerebral cortex.** *Brain Behav Evol* 2014, **83**:126-139.
14. Rakic P: **Confusing cortical columns.** *Proc Natl Acad Sci U S A* 2008, **105**:12099-12100.
15. Jiang X, Shen S, Cadwell CR, Berens P, Sinz F, Ecker AS, Patel S, Tolias AS: **Principles of connectivity among morphologically defined cell types in adult neocortex.** *Science* 2015, **350**:aac9462.
16. Hubel DH, Wiesel TN: **Ferrier lecture: functional architecture of macaque monkey visual cortex.** *Proc R Soc Lond B* 1977, **198**:1-59.
17. Mountcastle VB: *Perceptual Neuroscience: The Cerebral Cortex.* Cambridge, MA: Harvard University Press; 1998.
18. Hofman MA: **On the evolution and geometry of the brain in mammals.** *Prog Neurobiol* 1989, **32**:137-158.
19. Felleman DJ, Van Essen DC: **Distributed hierarchical processing in the primate cerebral cortex.** *Cereb Cortex* 1991, **1**:1-47.
20. Callaway E: **Local circuits in primary visual cortex of the macaque monkey.** *Ann Rev Neurosci* 1998, **21**:47-74.
21. Constantinople CM, Bruno RM: **Deep cortical layers are activated directly by thalamus.** *Science* 2013, **340**:1591-1594.
Shows that in mouse S1, response of layer 5 neurons to a whisker movement is driven by direct thalamic inputs, independently of layers 2-4. This challenges the common picture to date that feedforward input largely drives layer 4.
22. Pluta S, Naka A, Veit J, Telian G, Yao L, Hakim R, Taylor D, Adesnik H: **A direct translaminar inhibitory circuit tunes cortical output.** *Nat Neurosci* 2015, **18**:1631-1640.
23. Sherman SM, Guillery RW: **Distinct functions for direct and transthalamic corticocortical connections.** *J Neurophysiol* 2011, **106**:1068-1077.
24. Kim EJ, Juavinett AL, Kyubwa EM, Jacobs MW, Callaway EM: **Three types of cortical layer 5 neurons that differ in brain-wide connectivity and function.** *Neuron* 2015, **88**:1253-1267.
25. Velez-Fort M, Rousseau CV, Niedworok CJ, Wickersham IR, Rancz EA, Brown AP, Strom M, Margrie TW: **The stimulus selectivity and connectivity of layer six principal cells reveals cortical microcircuits underlying visual processing.** *Neuron* 2014, **83**:1431-1443.
26. Fu Y, Tucciarone JM, Espinosa JS, Sheng N, Darcy DP, Nicoll RA, Huang ZJ, Stryker MP: **A cortical circuit for gain control by behavioral state.** *Cell* 2014, **156**:1139-1152.
This study along with Refs [27*,28*,29*,30*,31*] identifies a circuit by which vasoactive-intestinal-peptide (VIP)-expressing inhibitory neurons can be activated by outside influences — motor cortex or cholinergic input from brainstem in response to motor activity, attentional input from cingulate cortex — and in turn inhibit SOM inhibitory neurons, thus disinhibiting PV inhibitory neurons and (in most cases) excitatory neurons and potentially opening a window for adult plasticity. If the result is that excitatory neurons increases their firing rate while receiving increased inhibition, this would suggest a network operating in the ISN regime. In response to locomotion, which activates VIP neurons (this study), Ref [29*] reported that both SOM and PV interneurons increased their firing in V1, indicating an ISN, while this study reported a decrease in SOM and increase in PV firing in V1, which is equivocal.
27. Pi HJ, Hangya B, Kvitsiani D, Sanders JI, Huang ZJ, Kepecs A: **Cortical interneurons that specialize in disinhibitory control.** *Nature* 2013, **503**:521-524.
See annotation to Ref [26*].
28. Lee S, Kruglikov I, Huang ZJ, Fishell G, Rudy B: **A disinhibitory circuit mediates motor integration in the somatosensory cortex.** *Nat Neurosci* 2013, **16**:1662-1670.
See annotation to Ref [26*].
29. Polack PO, Friedman J, Golshani P: **Cellular mechanisms of brain state-dependent gain modulation in visual cortex.** *Nat Neurosci* 2013, **16**:1331-1339.
See annotation to Ref [26*].
30. Fu Y, Kaneko M, Tang Y, Alvarez-Buylla A, Stryker MP: **A cortical disinhibitory circuit for enhancing adult plasticity.** *Elife* 2015, **4**:e05558.
See annotation to Ref [26*].
31. Zhang S, Xu M, Kamigaki T, Hoang Do JP, Chang WC, Jenvey S, Miyamichi K, Luo L, Dan Y: **Selective attention. Long-range and local circuits for top-down modulation of visual cortex processing.** *Science* 2014, **345**:660-665.
See annotation to Ref [26*].
32. Jiang X, Wang G, Lee AJ, Stornetta RL, Zhu JJ: **The organization of two new cortical interneuronal circuits.** *Nat Neurosci* 2013, **16**:210-218.
33. Bortone DS, Olsen SR, Scanziani M: **Translaminar inhibitory cells recruited by layer 6 corticothalamic neurons suppress visual cortex.** *Neuron* 2014, **82**:474-485.
34. Hangya B, Pi HJ, Kvitsiani D, Ranade SP, Kepecs A: **From circuit motifs to computations: mapping the behavioral repertoire of cortical interneurons.** *Curr Opin Neurobiol* 2014, **26**:117-124.
35. Pfeffer CK, Xue M, He M, Huang ZJ, Scanziani M: **Inhibition of inhibition in visual cortex: the logic of connections between molecularly distinct interneurons.** *Nat Neurosci* 2013, **16**:1068-1076.
36. Douglas RJ, Martin KAC, Whitteridge D: **A canonical microcircuit for neocortex.** *Neural Comput* 1989, **1**:480-488.
37. Carandini M, Heeger DJ: **Normalization as a canonical neural computation.** *Nat Rev Neurosci* 2012, **13**:51-62.
38. Roe AW, Pallas SL, Hahn J-O, Sur M: **A map of visual space induced in primary auditory cortex.** *Science* 1990, **250**:818-820.
39. LeCun Y, Bengio Y, Hinton G: **Deep learning.** *Nature* 2015, **521**:436-444.
A review of deep learning, the cortically inspired learning algorithms that have revolutionized artificial intelligence in the last few years.
40. Yamins DL, Hong H, Cadieu CF, Solomon EA, Seibert D, DiCarlo JJ: **Performance-optimized hierarchical models predict neural responses in higher visual cortex.** *Proc Natl Acad Sci U S A* 2014, **111**:8619-8624.
Demonstrates that the response properties across the hierarchical layers of a deep learning network trained to identify objects in visual images correspond surprisingly closely to the response properties across hierarchically arranged areas in the ventral visual stream, suggesting similar representational strategies in the two.
41. DiCarlo JJ, Zoccolan D, Rust NC: **How does the brain solve visual object recognition?** *Neuron* 2012, **73**:415-434.
42. Badre D: **Cognitive control, hierarchy, and the rostro-caudal organization of the frontal lobes.** *Trends Cogn Sci (Regul Ed)* 2008, **12**:193-200.

43. Badre D, D'Esposito M: **Is the rostro-caudal axis of the frontal lobe hierarchical?** *Nat Rev Neurosci* 2009, **10**:659-669.
44. Cavanaugh JR, Bair W, Movshon JA: **Nature and interaction of signals from the receptive field center and surround in macaque V1 neurons.** *J Neurophysiol* 2002, **88**:2530-2546.
45. Reynolds JH, Heeger DJ: **The normalization model of attention.** *Neuron* 2009, **61**:168-185.
46. Martinez-Trujillo JC, Treue S: **Feature-based attention increases the selectivity of population responses in primate visual cortex.** *Curr Biol* 2004, **14**:744-751.
47. Teich AF, Qian N: **V1 orientation plasticity is explained by broadly tuned feedforward inputs and intracortical sharpening.** *Vis Neurosci* 2010, **27**:57-73.
48. Crook JM, Kisvardy ZF, Eysel UT: **Evidence for a contribution of lateral inhibition to orientation tuning and direction selectivity in cat visual cortex: reversible inactivation of functionally characterized sites combined with neuroanatomical tracing techniques.** *Eur J Neurosci* 1998, **10**:2056-2075.
49. Ferster D, Chung S, Wheat H: **Orientation selectivity of thalamic input to simple cells of cat visual cortex.** *Nature* 1996, **380**:249-252.
50. Chung S, Ferster D: **Strength and orientation tuning of the thalamic input to simple cells revealed by electrically evoked cortical suppression.** *Neuron* 1998, **20**:1177-1189.
51. Finn IM, Priebe NJ, Ferster D: **The emergence of contrast-invariant orientation tuning in simple cells of cat visual cortex.** *Neuron* 2007, **54**:137-152.
52. Lien AD, Scanziani M: **Tuned thalamic excitation is amplified by visual cortical circuits.** *Nat Neurosci* 2013, **16**:1315-1323.
This study along with Ref [53**] demonstrates that thalamic excitation and intracortical excitation onto a L4 cell in primary visual cortex have similar orientation tuning, in agreement with previous studies of voltage tuning by the Ferster lab [49,50].
53. Li YT, Ibrahim LA, Liu BH, Zhang LI, Tao HW: **Linear transformation of thalamocortical input by intracortical excitation.** *Nat Neurosci* 2013, **16**:1324-1330.
See annotation to Ref [52**].
54. Li LY, Li YT, Zhou M, Tao HW, Zhang LI: **Intracortical multiplication of thalamocortical signals in mouse auditory cortex.** *Nat Neurosci* 2013, **16**:1179-1181.
Demonstrates a match of the tuning of thalamic and intracortical excitation to L4 neurons in primary auditory cortex.
55. Bruno R, Simons D: **Feedforward mechanisms of excitatory and inhibitory cortical receptive fields.** *J Neurosci* 2002, **22**:10966-10975.
56. Bruno R, Sakmann B: **Cortex is driven by weak but synchronously active thalamocortical synapses.** *Science* 2006, **312**:1622-1627.
57. Priebe NJ, Ferster D: **Mechanisms of neuronal computation in mammalian visual cortex.** *Neuron* 2012, **75**:194-208.
58. Li B, Thompson JK, Duong T, Peterson MR, Freeman RD: **Origins of cross-orientation suppression in the visual cortex.** *J Neurophysiol* 2006, **96**:1755-1764.
59. Lauritzen TZ, Krukowski AE, Miller KD: **Local correlation-based circuitry can account for responses to multi-grating stimuli in a model of cat V1.** *J Neurophysiol* 2001, **86**:1803-1815.
60. Kayser AS, Priebe NJ, Miller KD: **Contrast-dependent nonlinearities arise locally in a model of contrast-invariant orientation tuning.** *J Neurophysiol* 2001, **85**:2130-2149.
61. Troyer TW, Krukowski AE, Priebe NJ, Miller KD: **Contrast-invariant orientation tuning in cat visual cortex: feedforward tuning and correlation-based intracortical connectivity.** *J Neurosci* 1998, **18**:5908-5927.
62. Wimbauer S, Wenisch O, van Hemmen JL, Miller KD: **Development of spatiotemporal receptive fields of simple cells: II. Simulation and analysis.** *Biol Cybern* 1997, **77**:463-477.
63. Hubel DH, Wiesel TN: **Receptive fields, binocular interaction and functional architecture in the cat's visual cortex.** *J Physiol* 1962, **160**:106-154.
64. Reid RC, Alonso JM: **Specificity of monosynaptic connections from thalamus to visual cortex.** *Nature* 1995, **378**:281-284.
65. Piscopo DM, El-Danaf RN, Huberman AD, Niell CM: **Diverse visual features encoded in mouse lateral geniculate nucleus.** *J Neurosci* 2013, **33**:4642-4656.
66. Scholl B, Tan AY, Corey J, Priebe NJ: **Emergence of orientation selectivity in the mammalian visual pathway.** *J Neurosci* 2013, **33**:10616-10624.
67. Sun W, Tan Z, Mensh BD, Ji N: **Thalamus provides layer 4 of primary visual cortex with orientation- and direction-tuned inputs.** *Nat Neurosci* 2015 <http://dx.doi.org/10.1038/nn.4196>.
68. Ferster D, Miller KD: **Neural mechanisms of orientation selectivity in the visual cortex.** *Annu Rev Neurosci* 2000, **23**:441-471.
69. Troyer TW, Krukowski AE, Miller KD: **LGN input to simple cells and contrast-invariant orientation tuning: an analysis.** *J Neurophysiol* 2002, **87**:2741-2752.
70. Alitto HJ, Usrey WM: **Influence of contrast on orientation and temporal frequency tuning in ferret primate visual cortex.** *J Neurophysiol* 2004, **91**:2797-2808.
71. Palmer SE, Miller KD: **Effects of inhibitory gain and conductance fluctuations in a simple model for contrast-invariant orientation tuning in cat v1.** *J Neurophysiol* 2007, **98**:63-78.
72. Anderson JS, Lampl I, Gillespie D, Ferster D: **The contribution of noise to contrast invariance of orientation tuning in cat visual cortex.** *Science* 2000, **290**:1968-1972.
73. Sadagopan S, Ferster D: **Feedforward origins of response variability underlying contrast invariant orientation tuning in cat visual cortex.** *Neuron* 2012, **74**:911-923.
74. Li YT, Liu BH, Chou XL, Zhang LI, Tao HW: **Strengthening of direction selectivity by broadly tuned and spatiotemporally slightly offset inhibition in mouse visual cortex.** *Cereb Cortex* 2015, **25**:2466-2477.
75. Li LY, Ji XY, Liang F, Li YT, Xiao Z, Tao HW, Zhang LI: **A feedforward inhibitory circuit mediates lateral refinement of sensory representation in upper layer 2/3 of mouse primary auditory cortex.** *J Neurosci* 2014, **34**:13670-13683.
76. Li YT, Ma WP, Pan CJ, Zhang LI, Tao HW: **Broadening of cortical inhibition mediates developmental sharpening of orientation selectivity.** *J Neurosci* 2012, **32**:3981-3991.
77. Liu BH, Li YT, Ma WP, Pan CJ, Zhang LI, Tao HW: **Broad inhibition sharpens orientation selectivity by expanding input dynamic range in mouse simple cells.** *Neuron* 2011, **71**:542-554.
78. Liu BH, Li P, Sun YJ, Li YT, Zhang LI, Tao HW: **Intervening inhibition underlies simple-cell receptive field structure in visual cortex.** *Nat Neurosci* 2010, **13**:89-96.
79. Wu GK, Arbuckle R, Liu BH, Tao HW, Zhang LI: **Lateral sharpening of cortical frequency tuning by approximately balanced inhibition.** *Neuron* 2008, **58**:132-143.
80. Wu GK, Li P, Tao HW, Zhang LI: **Nonmonotonic synaptic excitation and imbalanced inhibition underlying cortical intensity tuning.** *Neuron* 2006, **52**:705-715.
81. Cruikshank SJ, Lewis TJ, Connors BW: **Synaptic basis for intense thalamocortical activation of feedforward inhibitory cells in neocortex.** *Nat Neurosci* 2007, **10**:462-468.
82. Swadlow HA: **Thalamocortical control of feed-forward inhibition in awake somatosensory 'barrel' cortex.** *Philos Trans R Soc Lond B Biol Sci* 2002, **357**:1717-1727.
83. Wehr M, Zador AM: **Balanced inhibition underlies tuning and sharpens spike timing in auditory cortex.** *Nature* 2003, **426**:442-446.

84. Wilent W, Contreras D: **Dynamics of excitation and inhibition underlying stimulus selectivity in rat somatosensory cortex.** *Nat Neurosci* 2005, **8**:1364-1370.
85. Pehlevan C, Sompolinsky H: **Selectivity and sparseness in randomly connected balanced networks.** *PLOS ONE* 2014, **9**:e89992.
- This study along with Refs [86*,87*] argues that the balanced network [113] will induce sparse firing to any input, with different neurons firing for different input patterns, and so will induce selective responses even for broadly tuned inputs. This mechanism produces neurons selective for a given pattern of inputs, and so can only explain selectivity for a property that always gives a similar input pattern. This mechanism could not explain orientation selectivity in L4 (e.g. the thalamic input pattern will be very different in response to a short or long bar or a low or high-frequency grating of a given orientation); but given invariant orientation selectivity in L4, it could explain how such selectivity is reconstituted in other layers in rodents from diverse L4 inputs.
86. Hansel D, van Vreeswijk C: **The mechanism of orientation selectivity in primary visual cortex without a functional map.** *J Neurosci* 2012, **32**:4049-4064.
- See annotation to Ref [85*].
87. Sadeh S, Rotter S: **Orientation selectivity in inhibition-dominated networks of spiking neurons: effect of single neuron properties and network dynamics.** *PLoS Comput Biol* 2015, **11**:e1004045.
- See annotation to Ref [85*].
88. Sato TK, Hausser M, Carandini M: **Distal connectivity causes summation and division across mouse visual cortex.** *Nat Neurosci* 2014, **17**:30-32.
- Demonstrates that input from surrounding primary visual cortex can increase responses to low-contrast visual stimuli but suppress responses to high-contrast visual stimuli. Other studies [89–92] had demonstrated that surrounding visual stimuli could have this effect on responses to a center visual stimulus, but had not directly shown that this effect could be caused by lateral connections from the surrounding regions of the same cortical area.
89. Sengpiel F, Blakemore C, Sen A: **Characteristics of surround inhibition in cat area 17.** *Exp Brain Res* 1997, **116**:216-228.
90. Polat U, Mizobe K, Pettet MW, Kasamatsu T, Norcia AM: **Collinear stimuli regulate visual responses depending on cell's contrast threshold.** *Nature* 1998, **391**:580-584.
91. Ichida JM, Schwabe L, Bressloff PC, Angelucci A: **Response facilitation from the "suppressive" receptive field surround of macaque V1 neurons.** *J Neurophysiol* 2007, **98**:2168-2181.
92. Schwabe L, Ichida JM, Shushruth S, Mangapathy P, Angelucci A: **Contrast-dependence of surround suppression in Macaque V1: experimental testing of a recurrent network model.** *Neuroimage* 2010, **52**:777-792.
93. Sceniak M, Ringach DL, Hawken M, Shapley R: **Contrast's effect on spatial summation by macaque v1 neurons.** *Nat Neurosci* 1999, **2**:733-739.
94. Anderson JS, Lampl I, Gillespie DC, Ferster D: **Membrane potential and conductance changes underlying length tuning of cells in cat primary visual cortex.** *J Neurosci* 2001, **21**:2104-2112.
95. Nienborg H, Hasenstaub A, Nauhaus I, Taniguchi H, Huang ZJ, Callaway EM: **Contrast dependence and differential contributions from somatostatin- and parvalbumin-expressing neurons to spatial integration in mouse V1.** *J Neurosci* 2013, **33**:11145-11154.
96. Vaicellunaite A, Eriskien S, Franzen F, Katzner S, Busse L: **Spatial integration in mouse primary visual cortex.** *J Neurophysiol* 2013, **110**:964-972.
97. Ayaz A, Saleem AB, Scholvinck ML, Carandini M: **Locomotion controls spatial integration in mouse visual cortex.** *Curr Biol* 2013, **23**:890-894.
98. Shushruth S, Ichida JM, Levitt JB, Angelucci A: **Comparison of spatial summation properties of neurons in macaque V1 and V2.** *J Neurophysiol* 2009, **102**:2069-2083.
99. Tsui JM, Pack CC: **Contrast sensitivity of MT receptive field centers and surrounds.** *J Neurophysiol* 2011, **106**:1888-1900.
100. Hunter JN, Born RT: **Stimulus-dependent modulation of suppressive influences in MT.** *J Neurosci* 2011, **31**:678-686.
101. Heuer HW, Britten KH: **Contrast dependence of response normalization in area MT of the rhesus macaque.** *J Neurophysiol* 2002, **88**:3398-3408.
102. Ahmadian Y, Rubin DB, Miller KD: **Analysis of the stabilized supralinear network.** *Neural Comput* 2013, **25**:1994-2037.
- This study along with Ref [103] introduced the stabilized supralinear network, the mechanism for cortical gain modulation described in the main text, showing its properties mathematically [102] and showing how it explains a large set of cortical nonlinear behaviors [103*].
103. Rubin DB, Van Hooser SD, Miller KD: **The stabilized supralinear network: a unifying circuit motif underlying multi-input integration in sensory cortex.** *Neuron* 2015, **85**:402-417.
- See annotation to Ref [102*].
104. Priebe NJ, Ferster D: **Inhibition, spike threshold and stimulus selectivity in primary visual cortex.** *Neuron* 2008, **57**:482-497.
105. Miller KD, Troyer TW: **Neural noise can explain expansive, power-law nonlinearities in neural response functions.** *J Neurophysiol* 2002, **87**:653-659.
106. Hansel D, van Vreeswijk C: **How noise contributes to contrast invariance of orientation tuning in cat visual cortex.** *J Neurosci* 2002, **22**:5118-5128.
107. Churchland MM, Yu BM, Cunningham JP, Sugrue LP, Cohen MR, Corrado GS, Newsome WT, Clark AM, Hosseini P, Scott BB, Bradley DC, Smith MA, Kohn A, Movshon JA, Armstrong KM, Moore T, Chang SW, Snyder LH, Lisberger SG, Priebe NJ, Finn IM, Ferster D, Ryu SI, Santhanam G, Sahani M, Shenoy KV: **Stimulus onset quenches neural variability: a widespread cortical phenomenon.** *Nat Neurosci* 2010, **13**:369-378.
108. Mitchell JF, Sundberg KA, Reynolds JH: **Spatial attention decorrelates intrinsic activity fluctuations in macaque area V4.** *Neuron* 2009, **63**:879-888.
109. Cohen MR, Maunsell JH: **Attention improves performance primarily by reducing interneuronal correlations.** *Nat Neurosci* 2009, **12**:1594-1600.
110. Ghose GM: **Attentional modulation of visual responses by flexible input gain.** *J Neurophysiol* 2009, **101**:2089-2106.
111. Lee J, Maunsell JH: **A normalization model of attentional modulation of single unit responses.** *PLoS ONE* 2009, **4**:e4651.
112. Boynton GM: **A framework for describing the effects of attention on visual responses.** *Vis Res* 2009, **49**:1129-1143.
113. van Vreeswijk C, Sompolinsky H: **Chaotic balanced state in a model of cortical circuits.** *Neural Comput* 1998, **10**:1321-1371.
114. Shao YR, Isett BR, Miyashita T, Chung J, Pourzia O, Gasperini RJ, Feldman DE: **Plasticity of recurrent I2/3 inhibition and gamma oscillations by whisker experience.** *Neuron* 2013, **80**:210-222.
115. Silberberg G, Markram H: **Disynaptic inhibition between neocortical pyramidal cells mediated by Martinotti cells.** *Neuron* 2007, **53**:735-746.
116. Kapfer C, Glickfeld L, Atallah B, Scanziani M: **Supralinear increase of recurrent inhibition during sparse activity in the somatosensory cortex.** *Nat Neurosci* 2007, **10**:743-753.
117. Tsodyks MV, Skaggs WE, Sejnowski BL, McNaughton TJ: **Paradoxical effects of external modulation of inhibitory interneurons.** *J Neurosci* 1997, **17**:4382-4388.
118. Murphy BK, Miller KD: **Balanced amplification: a new mechanism of selective amplification of neural activity patterns.** *Neuron* 2009, **61**:635-648.
119. Ozeki H, Finn IM, Schaffer ES, Miller KD, Ferster D: **Inhibitory stabilization of the cortical network underlies visual surround suppression.** *Neuron* 2009, **62**:578-592.
120. Haider B, Krause MR, Duque A, Yu Y, Touryan J, Mazer JA, McCormick DA: **Synaptic and network mechanisms of sparse and reliable visual cortical activity during nonclassical receptive field stimulation.** *Neuron* 2010, **65**:107-121.

121. Adesnik H, Bruns W, Taniguchi H, Huang ZJ, Scanziani M: **A neural circuit for spatial summation in visual cortex.** *Nature* 2012, **490**:226-231.
122. Pecka M, Han Y, Sader E, Mrcsic-Flogel TD: **Experience-dependent specialization of receptive field surround for selective coding of natural scenes.** *Neuron* 2014, **84**:457-469.
123. Zhou M, Liang F, Xiong XR, Li L, Li H, Xiao Z, Tao HW, Zhang LI: **Scaling down of balanced excitation and inhibition by active behavioral states in auditory cortex.** *Nat Neurosci* 2014, **17**:841-850.
124. Bullier J, Hupe JM, James AC, Girard P: **The role of feedback connections in shaping the responses of visual cortical neurons.** *Prog Brain Res* 2001, **134**:193-204.
125. Nassi JJ, Lomber SG, Born RT: **Corticocortical feedback contributes to surround suppression in V1 of the alert primate.** *J Neurosci* 2013, **33**:8504-8517.
126. Nassi JJ, Gomez-Laberge C, Kreiman G, Born RT: **Corticocortical feedback increases the spatial extent of normalization.** *Front Syst Neurosci* 2014, **8**:105.
127. Alitto HJ, Usrey WM: **Dissecting the dynamics of corticothalamic feedback.** *Neuron* 2015, **86**:605-607.
128. Crandall SR, Cruikshank SJ, Connors BW: **A corticothalamic switch: controlling the thalamus with dynamic synapses.** *Neuron* 2015, **86**:768-782.
129. Ganguli S, Biseley JW, Roitman JD, Shadlen MN, Goldberg ME, Miller KD: **One-dimensional dynamics of attention and decision-making in LIP.** *Neuron* 2008, **58**:15-25.
130. Elston GN: **Cortex, cognition and the cell: new insights into the pyramidal neuron and prefrontal function.** *Cereb Cortex* 2003, **13**:1124-1138.
131. Chaudhuri R, Knoblauch K, Gariel MA, Kennedy H, Wang XJ: **A large-scale circuit mechanism for hierarchical dynamical processing in the primate cortex.** *Neuron* 2015, **88**:419-431.
132. Dayan P, Abbott LF: *Theoretical Neuroscience.* Cambridge, MA: MIT Press; 2001.
133. Honey CJ, Theisen T, Donner TH, Silbert LJ, Carlson CE, Devinsky O, Doyle WK, Rubin N, Heeger DJ, Hasson U: **Slow cortical dynamics and the accumulation of information over long timescales.** *Neuron* 2012, **76**:423-434.
134. Murray JD, Bernacchia A, Freedman DJ, Romo R, Wallis JD, Cai X, Padoa-Schioppa C, Pasternak T, Seo H, Lee D, Wang XJ: **A hierarchy of intrinsic timescales across primate cortex.** *Nat Neurosci* 2014, **17**:1661-1663.
- Shows increasing slowness of cortical activity with movement posterior-to-anterior across cortical areas.
135. Amit D: *Modelling Brain Function: The World of Attractor Neural Networks.* Cambridge: Cambridge University Press; 1989.
136. Sussillo D, Abbott LF: **Generating coherent patterns of activity from chaotic neural networks.** *Neuron* 2009, **63**:544-557.
137. Laje R, Buonomano DV: **Robust timing and motor patterns by taming chaos in recurrent neural networks.** *Nat Neurosci* 2013, **16**:925-933.
138. Barak O, Sussillo D, Romo R, Tsodyks M, Abbott LF: **From fixed points to chaos: three models of delayed discrimination.** *Prog Neurobiol* 2013, **103**:214-222.
139. Sussillo D: **Neural circuits as computational dynamical systems.** *Curr Opin Neurobiol* 2014, **25**:156-163.
140. Van Hooser SD: **Similarity and diversity in visual cortex: is there a unifying theory of cortical computation?** *Neuroscientist* 2007, **13**:639-656.
141. Lund JS, Yoshioka T, Levitt JB: **Comparison of intrinsic connectivity in different areas of macaque monkey cerebral cortex.** *Cereb Cortex* 1993, **3**:148-162.
142. Van Hooser SD, Heimel JA, Chung S, Nelson SB: **Lack of patchy horizontal connectivity in primary visual cortex of a mammal without orientation maps.** *J Neurosci* 2006, **26**:7680-7692.
143. Kaschube M: **Neural maps versus salt-and-pepper organization in visual cortex.** *Curr Opin Neurobiol* 2014, **24**:95-102.
- An interesting review of the structural and possible functional differences between cortices that show columnar organization of selectivity and those that instead show 'salt-and-pepper' organization, and of possible differences in interactions during development that would lead to development of these alternative structures.