BRIEF COMMUNICATION

Multicenter Validation of the CamGFR Model for Estimated Glomerular Filtration Rate

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Abstract

Important oncological management decisions rely on kidney function assessed by serum creatinine–based estimated glomerular filtration rate (eGFR). However, no large-scale multicenter comparisons of methods to determine eGFR in patients with cancer are available. To compare the performance of formulas for eGFR based on routine clinical parameters and serum creatinine not calibrated with isotope dilution mass spectrometry, we studied 3620 patients with cancer and 166 without cancer who had their glomerular filtration rate (GFR) measured with an exogenous nuclear tracer at one of seven clinical centers. The mean measured GFR was 86 mL/min. Accuracy of all models was center dependent, reflecting intercenter variability of isotope dilution mass spectrometry–creatinine measurements. CamGFR was the most accurate model for eGFR (root-mean-squared error 17.3 mL/min) followed by the Chronic Kidney Disease Epidemiology Collaboration model (root-mean-squared error 18.2 mL/min).

Knowledge of kidney function measured as the glomerular filtration rate (GFR) informs clinical practice (1). GFR can be accurately measured (mGFR) using exogenous nuclear tracer clearance, but in practice, is frequently estimated (eGFR) using models based on routine clinical and biochemical data, specifically serum creatinine concentration. Creatinine is commonly measured using Jaffe or enzymatic methods, which in turn are calibrated using an isotope dilution mass spectrometry (IDMS) standard or a non–IDMS standard (2).

Recently, we derived a new model for GFR (CamGFR) using data from patients with cancer treated at the Cambridge University Hospitals NHS Foundation Trust, United Kingdom (3). CamGFR modeled GFR on a square root scale using non-IDMS–creatinine and biometric patient data and estimated GFR more accurately compared with other published models. This gain increased accuracy in GFR-based carboplatin chemotherapy dose calculations (3). Here we validate these findings for non-IDMS–creatinine-based estimation of GFR using multicenter data from patients with and without cancer.

Data were from the University Hospitals NHS Foundation Trusts in Cambridge, Southampton (4), and Manchester; Barts Health NHS Trust, London; a combined Welsh dataset (5,6); Western General Hospital, Edinburgh; and the Peter MacCallum Cancer Centre, Melbourne (7). Data on age, sex, height, weight,
serum creatinine concentration, histopathologically confirmed cancer diagnosis, ethnicity, and mGFR were obtained. Either chromium-51–labeled ethylenediamine tetraacetic acid (51Cr-EDTA) or 99mTc-diethylenetriaminepentaaetic acid (99mTc-DTPA) clearance was used to measure GFR (8,9). Serum creatinine was determined by enzymatic or Jaffe methods within 30 days of the mGFR date (Supplementary Table S1, available online). Adult patients with creatinine levels between 0.20 mg/dL and 4.5 mg/dL were included. From patients with multiple mGFR values, we included only the first value by date. Body surface area (BSA) was calculated using the DuBois and DuBois equation (10). The study was conducted at each institution according to its relevant regulatory and ethical requirements.

We compared mGFR with eGFR provided by six published models (CamGFR [3]; Martin [11]; Wright [12]; Mayo [13]; Modification of Diet in Renal Disease version 186 [14]; and Chronic Kidney Disease Epidemiology Collaboration [CKD-EPI [15]], along with two models for creatinine clearance (Cockcroft-Gault [16] and Jelliffe [17]).

To assess model performance, statistics were determined for bias (residual median), precision (residual interquartile range [IQR]), and accuracy (root-mean-squared error [RMSE]) and clinical robustness, by calculating the proportion of patients with an absolute percentage error greater than 20% (1-P20) for eGFR; 95% confidence interval (CI) = 0.16 to 0.17 mL/min for the CamGFR model was 17.3 mL/min (confidence interval [CI] = 16.7 to 17.9 mL/min) and 18.2 mL/min (CI = 17.6 to 18.7 mL/min) for the CKD-EPI model (P = .03) and the 1-P20 results for CamGFR were 0.295 (CI = 0.280 to 0.309) and 0.318 (CI = 0.303 to 0.333) for CKD-EPI, respectively (P = .03). In subgroup analyses, CamGFR was the most accurate model for most patient subgroups divided by tumor type, age, BSA, serum creatinine, or sex (Supplementary Figures S7–S10, available online).

Finally, CamGFR had the lowest RMSE both for male and female patients and in six out of seven centers. Model performance was not consistent between centers (Supplementary Table S5, available online, Figure 1), probably reflecting differences in non-IDMS–creatinine values (Supplementary Figure S3, available online).

We did not adjust the CamGFR model to include race as a potential variable for two reasons: the small number of black patients (n = 22) and the absence of a statistically significant difference in BSA, mGFR, or serum creatinine when we compared 10 random data draws matched for age and sex between non-black and black patients (Supplementary Figure S5 and Supplementary Table S4, available online). Other studies have documented systematic differences for the relationship between eGFR and creatinine for black patients (14,15), and our study is probably underpowered to detect this. The use of non-IDMS–creatinine data in this study represents a further limitation (19). Differences between non-IDMS and IDMS creatinine exist (2), and future work should expand the CamGFR model to

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**Table 1. Characteristics of study patients: summary of categorical variables split by center**

<table>
<thead>
<tr>
<th>Center</th>
<th>Total</th>
<th>Solid cancer</th>
<th>Hematological cancer</th>
<th>Noncancer</th>
<th>Female</th>
<th>Race, black</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cambridge</td>
<td>404</td>
<td>227</td>
<td>114</td>
<td>63</td>
<td>198</td>
<td>6</td>
</tr>
<tr>
<td>Edinburgh</td>
<td>597</td>
<td>472</td>
<td>22</td>
<td>103</td>
<td>245</td>
<td>0</td>
</tr>
<tr>
<td>London-Barts</td>
<td>108</td>
<td>108</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Manchester</td>
<td>1777</td>
<td>1777</td>
<td>0</td>
<td>0</td>
<td>1066</td>
<td>16</td>
</tr>
<tr>
<td>Melbourne</td>
<td>308</td>
<td>308</td>
<td>0</td>
<td>0</td>
<td>111</td>
<td>0</td>
</tr>
<tr>
<td>Southampton</td>
<td>436</td>
<td>436</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Wales</td>
<td>156</td>
<td>156</td>
<td>0</td>
<td>0</td>
<td>89</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>3786</td>
<td>3484</td>
<td>136</td>
<td>166</td>
<td>1709</td>
<td>22</td>
</tr>
</tbody>
</table>

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**Table 2. Characteristics of study patients: summary of continuous variables for all patients**

<table>
<thead>
<tr>
<th>Variable</th>
<th>Mean</th>
<th>SD</th>
<th>Minimum</th>
<th>Q1</th>
<th>Median</th>
<th>Q3</th>
<th>Maximum</th>
</tr>
</thead>
<tbody>
<tr>
<td>GFR, mL/min</td>
<td>86</td>
<td>32</td>
<td>9</td>
<td>61</td>
<td>85</td>
<td>109</td>
<td>209</td>
</tr>
<tr>
<td>Creatinine, mg/dL</td>
<td>0.99</td>
<td>0.28</td>
<td>0.43</td>
<td>0.83</td>
<td>0.95</td>
<td>1.11</td>
<td>4.45</td>
</tr>
<tr>
<td>Age, years</td>
<td>57</td>
<td>16</td>
<td>18</td>
<td>45</td>
<td>60</td>
<td>70</td>
<td>91</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>76</td>
<td>19</td>
<td>33</td>
<td>63</td>
<td>74</td>
<td>87</td>
<td>200</td>
</tr>
<tr>
<td>Height, cm</td>
<td>169</td>
<td>11</td>
<td>137</td>
<td>160</td>
<td>169</td>
<td>177</td>
<td>204</td>
</tr>
<tr>
<td>BSA, m²</td>
<td>1.85</td>
<td>0.25</td>
<td>1.17</td>
<td>1.68</td>
<td>1.85</td>
<td>2.02</td>
<td>3.17</td>
</tr>
</tbody>
</table>

*GFR was measured using either 99mTc-DTPA (Edinburgh and Melbourne) or 51Cr-EDTA (all others). 51Cr-EDTA = chromium-51-labeled ethylenediamine tetraacetic acid; GFR = glomerular filtration rate; BSA = body surface area (calculated using DuBois and DuBois); Q1 = 25th percentile; Q3 = 75th percentile; 99mTc-DTPA = 99mTc-diethylenetriaminepentaaetic acid.*
IDMS–creatinine data use. Of note, the CKD-EPI model was developed for use with IDMS–creatinine measurements specifically, but it still outperformed other models that have been developed with non-IDMS data.

The data were mostly from chemotherapy treatment–naive patients with cancer, and the longitudinal effect of treatment on eGFR requires further study. Probably attributable to the near-normal renal function of the majority of the patients in our study, we find that the underlying diagnosis of the patients does not affect the suitability of the models. CamGFR, developed on data from patients with cancer, performs best in noncancer patients, and CKD-EPI, developed on data from patients without cancer, performs well for data from patients with cancer.

This work is based on data from seven centers, and it confirms that of the available models, the CamGFR model estimates GFR most accurately, but the CKD-EPI model performs nearly as well overall and across the spectrum of relevant subgroups. The greatest gain in accuracy by these newer models over the older models, such as Cockcroft-Gault and Wright, was observed in younger patients and patients with lower creatinine values, probably reflecting the differences in model-development populations. However, even considering the different patient populations in different centers, it is likely that errors in estimating GFR can be reduced by standardizing the methods used to measure serum creatinine at different laboratories and using appropriate models. Given the linear relationship between GFR and carboplatin dose via the Calvert equation (20), improved estimates of GFR using CamGFR will translate into carboplatin prescriptions that are more accurate.

**Figure 1.** Performance analysis of commonly used and well-performing models. Results for the five best-performing models (CamGFR, CKD-EPI, Wright, MDRD-186, and Cockcroft-Gault) for the 3776 patients from the non-IDMS–creatinine validation dataset are displayed. Performance analysis of the other models is included in Supplementary Table S5 (available online). A pooled analysis of data from all centers and the individual center analyses are shown (first row). The residual (measured GFR–estimated GFR) median, which is a measure of a model’s bias, is displayed (second row). The residual interquartile range (IQR), which is a measure of a model’s precision, is displayed (third row). The RMSE, which is a measure of a model’s accuracy, is displayed. Accuracy is a combination metric of bias and precision (fourth row). The proportion of patients who have an absolute percentage error more than 20% (1-P20), which reflects clinical robustness by illustrating the proportion of patients with a clinically relevant error, is displayed. The best results are closest to zero for the residual median and the smallest value for IQR, RMSE, and 1-P20. All error bars are 95% confidence intervals calculated using bootstrap resampling with 2000 repetitions and a normal distribution approximation. CKD-EPI = Chronic Kidney Disease Epidemiology Collaboration; IDMS = isotope dilution mass spectrometry; MDRD-186 = Modification of Diet in Renal Disease version 186; RMSE = root-mean-squared error.
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Notes

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CamGFR is available online at https://sites.google.com/site/janowitzwilliamsgrf/.

References