

Enzyme Kinetics

In this exercise we will look at the catalytic behavior of enzymes. You will use Excel to answer the questions in the exercise section. At the end of this session, you must hand in answers to all the questions, along with print outs of any plots you created.

Background

Enzymes are the catalysts of biological systems and are extremely efficient and specific as catalysts. In fact, typically, an enzyme accelerates the rate of a reaction by factors of at least a million compared to the rate of the same reaction in the absence of the enzyme. Most biological reactions do not occur at perceptible rates in the absence of enzymes. One of the simplest biological reactions catalyzed by an enzyme is the hydration of CO₂. The catalyst in this reaction is carbonic anhydrase. This reaction is part of the respiration cycle which expels CO₂ from the body. Carbonic anhydrase is a highly efficient enzyme – each enzyme molecule can catalyze the hydration of 10⁵ CO₂ molecules per second.

Enzymes are highly specific. Typically a particular enzyme catalyzes only a single chemical reaction or a set of closely related chemical reactions. As is true of any catalyst, enzymes do not alter the equilibrium point of the reaction. This means that the enzyme accelerates the forward and reverse reaction by precisely the same factor. For example, consider the interconversion of A and B.



Suppose that in the absence of the enzyme the forward rate constant (k_f) is 10^{-4} s^{-1} and the reverse rate constant (k_r) is 10^{-6} s^{-1} . The equilibrium constant (K_{eq}) is given by the ratio of the two rate constants.

$$K_{\text{eq}} = \frac{[B]}{[A]} = \frac{k_f}{k_r} = \frac{10^{-4}}{10^{-6}} = 100 \quad (2)$$

The equilibrium concentration of B is 100 times that of A whether or not an enzyme is present. However, in the absence of an enzyme the reaction could take more than an hour to approach this equilibrium, whereas in the presence of an enzyme, equilibrium could be attained within a

second. The enzyme lowers the height of the energy barrier to the reaction thereby increasing the rate of the reaction, but since the rate of both the forward and reverse reaction are affected by the same amount, the equilibrium constant is not affected by the presence of the enzyme. the same amount (see Figure 1)

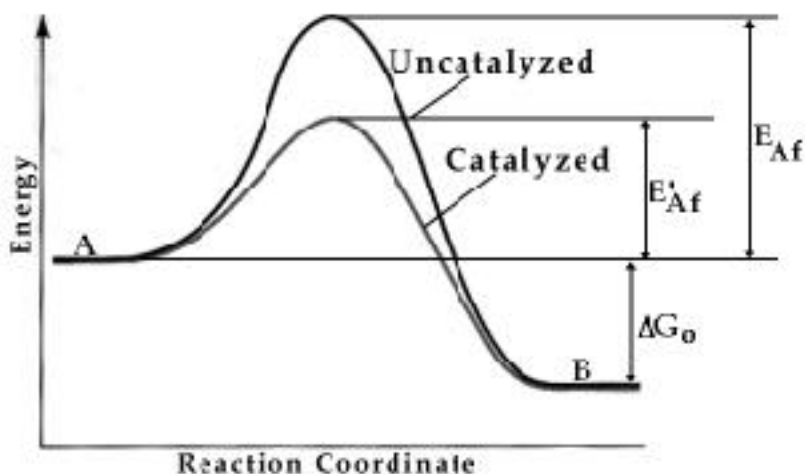


Figure 1

where, E_{Af} is the activation energy for the forward reaction ($A \rightarrow B$) in the absence of a catalyst and E'_{Af} is the activation energy for the forward reaction ($A \rightarrow B$) in the presence of a catalyst, and ΔG_o is the change in free energy for the reaction.

The equilibrium constant is related to ΔG_o as follows:

$$K_{eq} = e^{-\Delta G_o / RT}$$

Since ΔG_o is the same for the catalyzed and uncatalyzed reaction, K_{eq} is the same for both reactions.

One reason for the efficiency and specificity of an enzyme is the way the enzyme interacts with the reactant molecule, more commonly known as the substrate, in enzyme catalyzed reactions. The enzyme and substrate interact to form an enzyme-substrate complex. The interactions between the substrate and active site are weak, noncovalent interactions (i.e. the substrate does not covalently bind to the active site but weakly interacts with it through interactions like hydrogen-bonding, van der Waals interactions, etc). The orientation in which the two interact

is highly favorable for facilitating conversion of the substrate to product. In the enzyme-substrate complex, the substrate molecule binds to a very specific region of the enzyme molecule called the active site. These active sites are highly selective for a specific substrate molecule with which the enzyme binds. This is why enzymes are such highly specific catalysts, catalyzing a single reaction, or a set of closely related reactions. There are two proposed models to explain the specificity of the interaction between the substrate molecule and the active site of an enzyme.

(a) The “lock and key” model – in this model the substrate has a shape matching the enzyme’s active site (see figure 2)

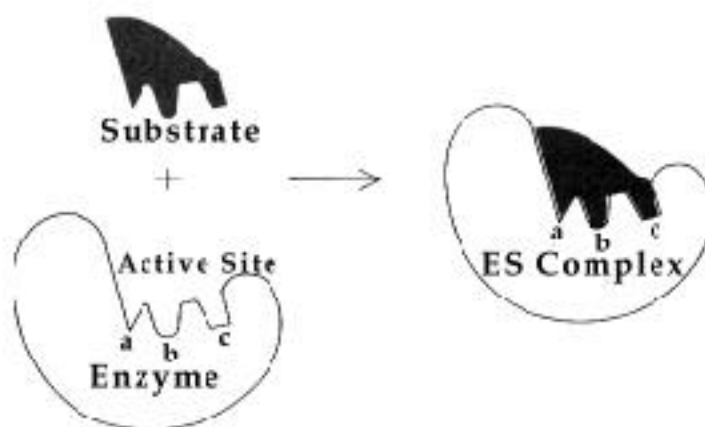


Figure 2

(b) The “induced fit model” – the active site has a shape complementary to that of the substrate *after* the substrate is bound (see figure 3)

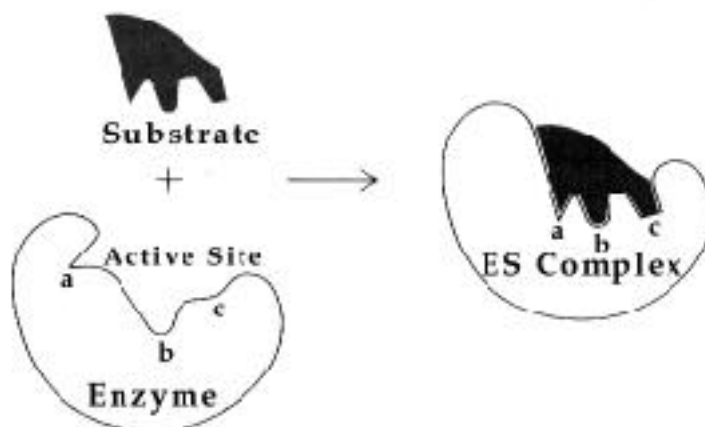


Figure 3

Enzyme kinetics

The mechanism of enzyme catalyzed reactions is often studied by making kinetic measurements on enzyme-substrate reaction systems. These studies include measuring rates of the enzyme-catalyzed reactions at different substrate and enzyme concentrations. Here we will look at a simple model for the catalytic behavior of an enzyme and the kinetic model that arises from this model.

For many enzymes, if we were to plot the rate of catalysis, V (also known as the reaction velocity), vs. the substrate concentration, $[S]$ (at a fixed enzyme concentration) we would see a plot as shown in figure 4.

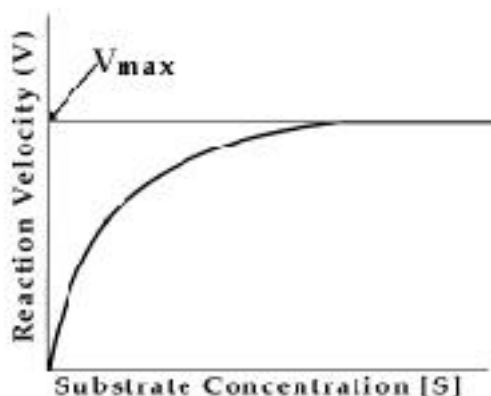


Figure 4

Looking at this plot, we see that V varies linearly with $[S]$ for small $[S]$. As $[S]$ increases, V “plateaus” indicating that V becomes independent of $[S]$ at large values of $[S]$. The simplest model which accounts for this behavior is:



where E is the enzyme, S the substrate, ES the enzyme-substrate complex, P the product of the enzyme-catalyzed reaction, k_1 the rate constant of the forward reaction of $E+S$, k_{-1} the rate of the reverse reaction where the enzyme-substrate complex, ES , falls apart to $E+S$ and k_2 the rate constant of the forward reaction of ES forming $E+P$. In this model, it is assumed that none of

the product reacts with the enzyme to form the enzyme-substrate complex, ES (this is true during the initial stages of the reaction when [P] is low, but towards the end of the reaction when [P] is high this may no longer be true).

We need to derive an expression that relates the reaction velocity, V , to the concentrations of the substrate and enzyme and the rates of the individual steps. From equation (3) the reaction velocity, V can be expressed as:

$$V = k_2 [ES] \quad (4)$$

Since ES is an intermediate and hence its concentration unknown, we have to express [ES] in terms of known values. The rates at which [ES] is formed and falls apart are:

$$\text{Rate of formation of [ES]} = k_1 [E] [S] \quad (5)$$

$$\text{Rate at which [ES] falls apart} = (k_{-1} + k_2) [ES] \quad (6)$$

We can use the *steady-state* approximation to express V in terms of known quantities. Under the steady-state approximation, the concentration of the intermediate [ES] stays a constant, while the concentrations of reactants and product change. The steady state occurs when eqn 5 = eqn 6 i.e.

$$k_1 [E] [S] = (k_{-1} + k_2) [ES] \quad (7)$$

Rearranging,

$$[ES] = \frac{[E][S]k_1}{k_{-1} + k_2} \quad (8)$$

Define, K_M , the Michaelis constant, as

$$K_M = \frac{k_{-1} + k_2}{k_1} \quad (9)$$

Substituting (9) into (8)

$$[ES] = \frac{[E][S]}{K_M} \quad (10)$$

Since in most situations the enzyme concentration is very small ($[E] \ll [S]$), the concentration of the uncombined S is almost equal to the total concentration of S. The concentration of

uncombined E is equal to the total enzyme concentration $[E_o]$ minus the concentration of the complex $[ES]$

$$[E] = [E_o] - [ES] \quad (11)$$

Substituting (11) into (10)

$$[ES] = \frac{([E_o] - [ES])[S]}{K_M} \quad (12)$$

Solving (12) for $[ES]$,

$$[ES] = [E_o] \frac{[S]/K_M}{1 + S/K_M} \quad (13)$$

or,

$$[ES] = [E_o] \frac{[S]}{[S] + K_M} \quad (14)$$

Substituting (14) into (4)

$$V = k_2 [E_o] \frac{[S]}{[S] + K_M} \quad (15)$$

The maximum reaction velocity, V_{max} , is reached when all enzyme sites are saturated with the substrate. This will happen when $[S] \gg K_M$, so that $[S]/([S]+K_M)$ approaches 1. In this limit, we can express V_{max} (from (15)) as:

$$V_{max} = k_2 [E_o] \quad (16)$$

Substituting (16) into (15),

$$V = V_{max} \frac{[S]}{[S] + K_M} \quad (17)$$

If we were to plot V vs S the resulting plot will have a shape as shown in figure 4. Hence, equation (17) describes the kinetic behavior of an enzyme as modeled by the kinetic scheme in equation (3). Looking at equation (17) at very low $[S]$, when $[S] \ll K_M$, $V \sim [S]V_{max}/K_M$, that is, the rate is proportional to $[S]$ (describes the linear region of the plot in figure 4). At high $[S]$,

when $[S] \gg K_M$, $V = V_{\max}$ and hence independent of $[S]$ (the “plateau” region of the plot in figure 4).

Equation (17) can be re-arranged as:

$$\frac{1}{V} = \frac{1}{V_{\max}} + \frac{K_M}{V_{\max}} \frac{1}{[S]} \quad (18)$$

If we were to plot $1/V$ vs. $1/[S]$ we would obtain a straight line with a y-intercept = $1/V_{\max}$ and a slope = K_M/V_{\max} (see figure 5). This plot is called a Lineweaver-Burke plot.

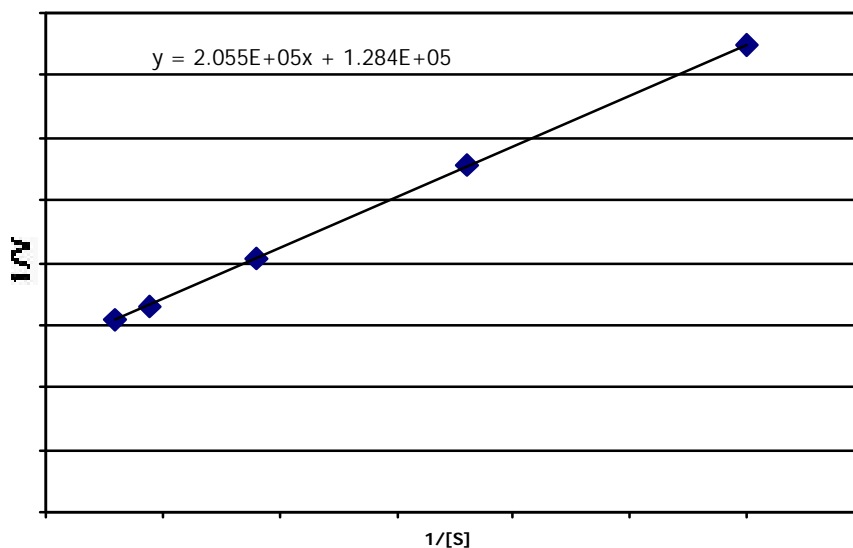


Figure 5

Significance of K_M

From equation 18, when $[S] = K_M$, then $V = V_{\max}/2$. Hence K_M is equal to the substrate concentration at which the reaction rate is half its maximum value. In other words, if an enzyme has a small value of K_M , it achieves its maximum catalytic efficiency at low substrate concentrations. Hence, *the smaller the value of K_M , the more efficient is the catalyst*. The value of K_M for an enzyme depends on the particular substrate. It also depends on the pH of the solution and the temperature at which the reaction is carried out. For most enzymes K_M lies between 10^{-1} and 10^{-7} M.

Determining K_M and V_{max} experimentally

To characterize an enzyme-catalyzed reaction K_M and V_{max} need to be determined. The way this is done experimentally is to measure the rate of catalysis (reaction velocity) for different substrate concentrations. In other words, determine V at different values of $[S]$. Then plotting $1/V$ vs. $1/[S]$ we should obtain a straight line described by equation (18). From the y-intercept and the slope, the values of K_M and V_{max} can be determined. For example, use EXCEL to plot the data shown below. Fit the data to a straight line, and from the equation of the straight line determine the values of K_M and V_{max} .

[S] (mM)	V (mM/sec)	1/[S] (mM ⁻¹)	1/V (sec/mM)
8.33	3.62E-06	0.12	2.76E+05
5.55	3.39E-06	0.18	2.95E+05
2.77	2.75E-06	0.36	3.64E+05
1.38	1.99E-06	0.72	5.02E+05
0.83	1.49E-06	1.2	6.73E+05

You should obtain a plot as shown in figure 6 below. From the fit to the data show that $K_M = 1.6$ for this data and V_{max} is 4.32×10^{-6} mM/sec

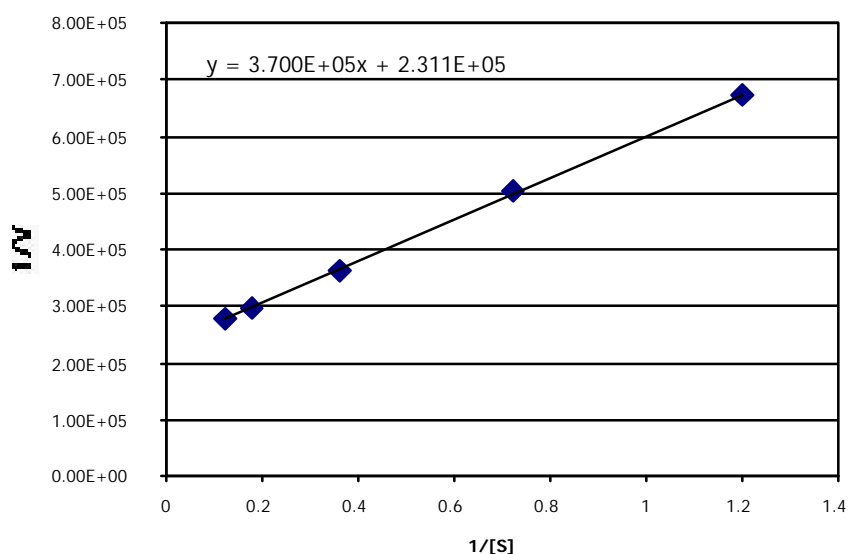


Figure 6

EXERCISES

Use Excel to answer the questions given below.

1) In a typical experiment to characterize an enzyme, K_M and V_{max} need to be measured. To do this, first a series of experiments are run, where for a given initial substrate concentration, the rate at which the product is formed is monitored. The rate of product formation is monitored by measuring the product concentration as a function of time. A plot of concentration of product vs. time will be linear at short time (as long as most of the substrate molecules have not reacted) and the slope of the linear fit to the data is the rate of product formation for a given initial substrate concentration. This is the rate of the reaction (V). The experiment is repeated for different initial substrate concentrations (keeping the enzyme concentration fixed) and the rate of product formation at each of these substrate concentrations determined. This series of experiments results in a set of data: reaction rates (V) as a function of substrate concentration $[S]$. Using this data a plot of $1/V$ vs. $1/[S]$ is constructed – this plot should be linear as dictated by equation (18), and from the linear fit K_M and V_{max} are determined.

You will use Excel to analyze data from an experiment of the type just described to determine the K_M and V_{max} that characterizes the enzyme used. Open the file “**exercise1**” in Excel. *Save this file to your floppy disk and work from the saved file.* This file contains the data from five runs (RUN 1 to RUN 5) monitoring the product concentration as a function of time for different initial substrate concentrations (all carried out with the same enzyme concentration). Plot each data set as product concentration vs. time. Fit the data to a straight line and determine the slope. The slope is the rate of the reaction, V , for a particular value of $[S]$.

Note: In the “**Add Trendline**” dialog box (accessed from the Chart menu), first select “**Linear**” (for a linear fit to the data) and then click on **Options** and select the “**Display equation on chart**” option. This should show the equation of the straight line fit to the data. Double click on the equation. This will bring up another dialog box. Click on the “**Number**” option and choose **Scientific** and increase the number of decimal places to **3**.

You should end up with five values of V for each of the five values of $[S]$. Plot $1/V$ vs. $1/[S]$ and determine K_M and V_{max} from the straight line fit to the data and equation 18.

Submit a printout of the plot of $1/V$ vs. $1/[S]$ and report the values of K_M and V_{max} .

2) Open file “**exercise2**” in Excel. *Save this file to your floppy disk.*

This file contains a series of data from three experiments. In each experiment, the reaction rates (V) were monitored as a function of substrate concentration $[S]$, but in each of the three experiments a DIFFERENT ENZYME was used. The concentration of the enzymes was kept fixed, as well as the pH of the solution and the temperature of the reaction. In addition, the V 's for each experiment were determined for the same substrate concentration.

From the data given, plot Lineweaver-Burke plots for each experiment, and using the values of K_M for each enzyme, determine which enzyme (X, Y or Z) is catalytically most efficient.

Submit printouts of the plots you created along with your calculations for K_M .