Lecture 17. Viruses that Infect Lymphocytes: EBV and HIV
Long term Host - Pathogen Relationships

Host immune defense consistently fail to clear a few viral infections

Examine mechanisms pathogens use to evade immune defense

Contrasting host-pathogen relationships of two prototype infections

Epstein-Barr Virus (EBV)
- Large DNA virus, e.g., herpesviruses
- Coevolved with host species over millions of years
- Genetically stable
- Persist in host in latent pattern of viral gene expression in response to T cell surveillance

Human immunodeficiency virus (HIV-1)
- RNA retrovirus
- Recently introduced into humans
- Error-prone viral replication mechanisms resulting in “swarms” of distinct strains “quasispecies”
- Overwhelm host by escaping from immune surveillance

Host-pathogen relationships
Some mechanisms of avoiding immune surveillance

1. Avoid recognition by cytotoxic T cells
   - Evolution of viral strains that avoid presentation by MHC by mutating class I molecule peptide anchor amino acids or amino acids recognized by T cells in immunodominant peptides
   - Blocking of antigen processing and presentation

2. Modification of the immune response
   - e.g., release of anti-inflammatory cytokines, IL-10

Syndromes resulting from EBV infection

Age of host influences character of primary infection

- Primary infection with EBV in childhood usually subclinical
- 25-70% of newly infected adolescents and adults develop infectious mononucleosis:
  - Fever
  - Lymphadenopathy
  - Pharyngitis
  - Transient heterophil antibodies
  - Activated CD8 cytotoxic anti EBV T cells (“atypical lymphocytosis”)

EBV is a B cell lymphotropic herpesvirus
Stages of EBV infection

Binding
- EBV surface glycoprotein to CD21 (CR2)
  - CD21 expressed on B cells as BCR co-receptor complex with CD19
  - CD21 also expressed on some epithelial cells, accounting for tropism

Triggers
- T-independent polyclonal B cell activation Ig synthesis and B cell proliferation

Results in T-independent release of heterophil and other antibodies (Rheumatoid factor, cold agglutinins, ANA)

EBV enters the cell by receptor mediated endocytosis

Syndromes resulting from EBV infection

EBV-driven neoplasms
- Nasopharyngeal carcinoma, Gastric cancer subset
- B cell lymphomas:
  - Burkitt’s Lymphoma
  - Immunoblastic lymphoma in immunosuppressed host
  - Subset of Hodgkin’s disease
Initially EBV replicates as a productive lytic infection

- IgM antibodies to Viral Capsid Antigens (VCA) and Early Antigens (EA) are found at clinical presentation, indicating lytic replication and persists for 1-2 months.
- IgG anti VCA appears at time of clinical presentation and persists lifelong “standard EBV titre”
- Antibodies to EA Peak at 3-4 weeks; marker of more severe disease
- Lytically infected cells are largely eliminated by EBV-specific cytotoxic cells, NK cells, interferon-mediated mechanisms and ADCC

By 3-6 weeks EBV enters latent stage in majority of remaining infected B cells

- Antibodies to latent EB Nuclear Antigens (EBNA’s) appear 3-6 weeks after initial infection; last lifelong
- Virus evades cytotoxic response in latent form
- CD8 T cells play a crucial role in enforcing the maintenance of latency and thwart proliferation of EBV infected B cells by killing the B cell

EBV is maintained in its latent infective cycle as a multicopy circular 172Kd ds plasmid minichromosome with replication linked to B cell proliferation

9 Latent proteins:
- Six nuclear antigens: EBNA1, 2, 3A, 3B, 3C, and EBNA leader protein (EBNA-LP)
- Three latent membrane proteins: LMP1, LMP2A, and LMP2B

- EBNA1 binds to the EBV ori, initiating replication and also acting as a transcriptional enhancer

EBNA1 contains a gly-al a repeat region that inhibits the ATP motor of the proteasome, impeding further insertion of EBNA1 into the proteasome, thus halting its degradation, a strategy for avoiding surveillance

B cell lymphoblastoid cell line (BLCL)

BLCL can be derived from nearly everyone, in vitro
Express all latency genes, a pattern designated latency III
BLCL phenotype high expression of:
- B cell activation markers CD23, CD30, CD39, and CD70
- Cellular adhesion molecules LFA1 (CD11a/18), LFA3 (CD58), and ICAM1 (CD54)

Because of the adhesion molecules these BLCLs grow in large clumps in tissue culture

Immunoblastic lymphomas

Develop in transplant recipients or other patients receiving T cell immunosuppressive therapies

- Resembles the BLCL phenotype
- Express all latency genes
- Start as multi-/polyclonal proliferations
- Withdrawal of immunosuppression results in regression
- Secondary transformation events may occur: monoclonal lymphoma

Burkitt’s lymphomas

Chronic immune system drive, e.g. by malaria implicated as cofactor, but no overt immune deficiency

- Exhibit a different gene expression pattern “latency I”
- Only abundant EBNA1 transcription is found
- Lymphoma cells display a distinct phenotype: CD10+ (CALLA) and CD77+ (BLA), but lack expression of activation and adhesion molecules
- In culture Burkitt tumor B cell lines grow as dispersed single cells
The T cell immune response to viruses often uses a very small number of different CD8 T cell clones directed to one or a few “immunodominant” peptides encoded by the viral genome, that are often presented by just one allelic type of an individual’s HLA molecules.

The achilles’ heel of the immune system

EBV infection results in nasopharyngeal carcinomas in Papua New Guinea and southern China

HLA-A11 distribution: African 1.5%, Caucasian 6.9%, Asian 16.3%

HLA-A11-positive Caucasians nearly always respond to two immunodominant HLA-A*1101 epitopes of the nuclear antigen EBNA3B (EBNA4):

\[
\text{IV TDFSVIK} \quad 416 \text{ to } 424
\]

AV FDRKSDAK \quad 399 \text{ to } 408

These sequence motifs were often mutated in EBV strains in lowland Papua New Guinea and southern China, areas where more than 50% of individuals carry the HLA-A*1101 allele.

All mutated strains have a point A -> C mutation, which produces a Lys -> Thr (K -> T) change in residue 424 of EBNA4 at position 9 of the CTL epitope (Frequency dependent selection).

Loss of recognition of immunodominant epitope and ability to recognize EBV is a mechanism of escaping the CTL response implicated in neoplastic transformation.

Organization of HIV-1 Provirus

Size \(9 \text{ kb}\)

Contains 9 genes encoding 15 proteins

Early events of HIV-1 infection

Binding of envelope gp120 prompts p41 to project 3 fusion domains that harpoon the membrane, resulting in fusion.

Integration leads to either latent or transcriptionally active infection.

Host Response to HIV-1 infection

First Phase: CD8 T cell response of immune system controls initial destruction of memory/effector CD4 T cells, but does not eliminate infectious virus primarily located in monocytes and memory CD4 T cells.

Antibodies to HIV-1 are formed but these neither clear the infection nor are protective.

- Acute illness- “flu-like”
- Clinical asymptomatic phase- 2-12 or more years

Second Phase: HIV-1 escapes the CD8 T cell response and mutations in the viral envelope now favor infection and destruction of naïve CD4 T cells.

Acquired immune deficiency (AIDS) appears upon depletion of critical CD4 T cell subsets.

Immune response to HIV-1 and effects of HIV infection

CD4 T cells #/μl

CLINICAL

- Flu-like illness
- Asymptomatic phase
- Chronic lymphadenopathy
- “Set Point”
- Symptomatic phase
- AIDS
- Mucous membrane infections, etc.
HIV must adapt and evolve in an environment determined by attributes of the host’s immune system:
- MHC alleles
- TCR repertoire
- Polymorphism of viral entry receptors
- Chemokine and cytokine milieu (e.g., parasitic infections)
- Other genes regulating immune response
- Prior immune history
- Age

Outcome of infection depends on biology of host, especially whether immune response targets critical HIV structures and HIV-1 mutational capacity, etc.

HIV-1 genomically highly diverse

Phylogenetic relationships

Cellular Specificity, “Tropism” of HIV strains

Based on envelope structure

- The viral envelope contains sequences that interact with a membrane viral receptor complex composed of CD4 and one of several chemokine receptors
- The sequence of a given viral envelope is specific for one of the chemokine receptor types
- The main two chemokine receptors are CCR5 and CXCR4 that are distributed on different cell lineages
- Strains that bind to CCR5 are termed “R5” tropic and those that bind CXCR4 are termed “R4” tropic

Chemokine Receptors:

**CCR5**
- Ligands: RANTES, MIP-1α, MIP-1β are inflammatory cytokines made by activated CD8 and CD4 T cells in the immune response to HIV and compete with R5 HIV binding to membrane receptor complex, blocking progress of the infection
- Distribution: CCR5 found on monocytes, DC and effector, memory or activated T cells, not naïve CD4 T cells
- Biology: CCR5 responsible for migration of memory and effector T cells, monocytes and dendritic cells to sites of inflammation
- Several CCR5 polymorphisms: e.g., a32 mutant allele render CCR5 unexpressed and incapable of binding HIV R5 strains.
  - a32 Homozygote frequency 1%, heterozygote ~10% in N.European Caucasoids, but X4 strains are still infective

**CXCR4**
- Ligand: Stromal derived growth factor 1 (SDF-1) produced by stromal cells. Competes with HIV binding, but not produced in inflammation or by T cells
- Receptor: expressed on monocytes, naïve T-cells, B-cells, etc. X4 virus preferentially infects naïve/activated T cells
- Biology: SDF-1/CXCR4 responsible for migration/homing of naïve T cells to lymph node

Chemokine Receptors: Coreceptors for HIV entry

**CXCR4**

- **R5** is almost always the sexually transmissible form of the virus
- Primary isolates from newly infected individuals are usually R5
- R5 strains mainly replicate in monocytes. Activated and memory T cells are infected, but at lower efficiency
- Much of the viral load in earlier phase of HIV infection is in the monocytes and macrophages and the number of CD4 T cells though decreased, remains stable
**Acute HIV-1 Infection "Flu-Like"**

**Clinical**
- Headache, retro-orbital pain, myalgias, pharyngitis, fever, Adenopathy and malaise may last for several months
- Transient thrombocytopenia and CD4 T-cell lymphopenia

**Viral**
- Rapid appearance of marked viremia with an R5 strain infecting monocytes and memory CD4 T cells
- This results in acute CD4 T-cell lymphopenia
- Integration in memory CD4 T cells provides a long-lived reservoir where HIV can remain latent
- Structurally the initial virus strain has no, or very limited diversity

**Acute Infection**

**Development of anti HIV Immune Response**
- With onset of a CD8 T-cell immune response viremia falls from ~5x10^6 /ml to <10^4 /ml
- The CD4 T-cell count rises from ~400 to >800/μl
- Degree of viral suppression and return of CD4 T cell levels (set point) varies and correlates with the length of the asymptomatic period
- HIV species begin to diversify, viral variants appear reflecting successful attempts to escape the suppression of the CD8 T cell response

**CD8 T-cell Response to HIV-1**
- Establishes asymptomatic phase of infection
- Specific CD8 CTL lysis of HIV- infected target cells (macrophages and CD4 T cells) via perforin pathway and/or apoptosis via upregulation of fas ligand
- Strong inhibition of viral infectivity by release of chemokines (MIP-1α/β, RANTES) that bind to CCR5 and compete with coreceptor dependent entry of R5 HIV-1
- Release of IFN-γ and secondarily TNF-α, decrease LTR-driven transcription

**Excessive anti HIV CD8 T cell response may result in diffuse infiltrative lymphocytosis syndrome (DILS) simulating Sjogren’s syndrome**

**H & E**

**HLA-DR stain**

**Salivary gland biopsy**

**CT scan**

**CD8 T cells >2000/μl**

**Nuclide scan**
DLS is usually associated with long term non progression and a favorable outlook. However, it is also associated with a type of B cell lymphoma that occurs early in the course of HIV infection, reflecting chronic B cell stimulation.

Anti-HIV antibodies usually appear in several weeks, they play a minor role.

Variants emerge too quickly for effective in vivo antibody neutralization. Other mechanisms.

**Immune Responses in asymptomatic phase**

- Depends on a relatively few CD8 T cell clones
- Maintenance of a few CD8 T-cell expanded memory/effector CTL clones, each comprising 1-5% of CD8 T cell repertoire
- Clones each recognize different immunodominant HIV peptides, great individual variation in number and particular peptide recognized
- More clones = generally good outlook for long asymptomatic period (>12yrs), fewer clones = rapid progression of HIV infection (<2yrs)

**Long term non progressors**

- A subset of infected individuals that remain asymptomatic for >12 years
- Particular HLA types, e.g. HLA-B27, B57, etc.
- Low levels of plasma virions, CD4 counts >500/ul
- High CD8 T-cell counts, may be > 3,000/ul
- High chemokine release (RANTES, MIP)
- CTL response is against critical conserved region of HIV gag, env, pol that cannot readily be mutated without loss of viral function-This appears to be the key factor!

The environment formed by peptide binding properties of MHC molecules influences evolution of the HIV infection.

**Role of MHC in Recognition of HIV peptides**

**Rapid HIV progression in HLA-B35 individuals**

<table>
<thead>
<tr>
<th>Peptide able to bind each allelic molecule</th>
<th>HLA-B*27052</th>
<th>HLA-B*3501</th>
<th>HLA-B*0702</th>
</tr>
</thead>
<tbody>
<tr>
<td>IGRQVQERK</td>
<td>KHRVQOREK</td>
<td>DPHPQEVVL</td>
<td></td>
</tr>
<tr>
<td>IRPVSTOLL</td>
<td>ARILMRY</td>
<td>FPGVKTPL</td>
<td></td>
</tr>
<tr>
<td>TNPNSNKRE</td>
<td>ERDRSRIR</td>
<td>RPVSTQ LL</td>
<td></td>
</tr>
<tr>
<td>IRQROGKVR</td>
<td>LRSCLLFST</td>
<td>SFLSFQTHL</td>
<td></td>
</tr>
<tr>
<td>SRAKMSNLTL</td>
<td>TRIVELGR</td>
<td>FPRARQGL</td>
<td></td>
</tr>
<tr>
<td>LREQPGQDNR</td>
<td>CRARRHIPR</td>
<td></td>
<td></td>
</tr>
<tr>
<td>FRPGGDMR</td>
<td>IRQGERL</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WSRELYRK</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

# of peptides | 15 | 0 | 6
Viral Response near end of asymptomatic period

- Rate of cellular infection and potential mutations increases
- Definitive viral escape occurs when virus is no longer presented by MHC to available CD8 T cell clones
- Continual generation of env mutations
- Selection against R5 variants by CD8 T-cell CCR5 chemokines that blocks infection is finally bypassed
- Change in cellular tropism by env mutations leads to X4 phenotype (CXC4, T-tropic)
- Enhanced T-tropism of X4 leads to more significant impairment of CD4 T-cell compartment
  
  Loss of the “epitope war”

Reasons for CD4 T cell loss in HIV-1 Infection

During asymptomatic phase and transition to AIDS
Accelerated loss in number of CD4 T cells

- Activation of large numbers of mature and naive CD4 T cells by cytokines, etc. during antiviral response (Bystander activation, homeostatic regulation) leads to loss of repertoire by physiologic apoptosis
- Thymic derangement results in failure to generate new naive CD4 T cells to repopulate repertoire
- CD8 T cell killing of infected CD4 T cells
- ADCC by NK cells, etc. to infected CD4 T cells

Another reason for CD4 T cell loss

CD4 T cell activation initiates HIV replication

T cell activation causes, among other effects, a marked increase in cyclin T1, NFAT and NFκB

AIDS is the consequence of progressive CD4 loss

Appearance of different infections as severity of immune deficiency increases

- Candida (Thrush)
- Salmonella - microbial persistence
- Mycobacterium tuberculosis reactivation, Cryptosporidium
- Activation of latent herpes zoster
- EBV reactivation and development of polyclonal immunoblastic lymphomas, Kaposi’s sarcoma (HHV-8)
- Pneumocystis carinii
- Progressive cytomegalovirus infections, M. avium complex

HIV virus vaccines have failed, Why?

- Immunization with rENV produce anti HIV antibodies
- But antibodies induced by immunization fail to protect as shown in multiple trials
- A live attenuated virus has not yet proved achievable
- But recombinant viral vectors vaccines with portions of the HIV genome have been developed and produce CD8 immunity
HIV virus vaccines have failed, Why?

• Heterogeneity of HIV strains: need many immunodominant peptides directed to critical regions of viral genome for different MHC types because no cross protection (Think Zinkernagel-Doherty experiment)

However, the most telling reason is that we lack critical information about what is occurring during HIV infection

Vaccination produces CD8 T cell immunity

But:

Does not confer protection
May cause the infection to progress more rapidly

Two examples:

vCP205 a recombinant live virus canarypox vector vaccine expressing gp41, Gag and Protease HIV genes induces CD8 T cell immunity

Case Report of a failure of a recombinant live vaccine
Betts et al. PNAS 2005, 102:4512
Case # 202-T07, an HLA-B*2705 HIV-negative male homosexual
vCP205 canarypox vector expressing gp41, Gag and Protease vaccination course given over 5 months
Immune response documented to two CD8 epitopes and one CD4 epitope including response to the HLA-B*2705-restricted Gag peptide KRWIIlGLNK in central and peripheral memory/effector CD8 T cells
CD28+CCR7+CD45RO+ and CD28-CCR7-CD45RO-

Approximately 18 months later 202-T07 had unprotected anal intercourse with an undisclosed HIV+ partner

Shortly thereafter, he developed flu-like symptoms and was then found to be positive for HIV antibodies, with a plasma viral load of 234,695 HIV-1 virions/ml

The acute infection induced a recall response to the B*2705-restricted clone, expanding it from 0.05% of CD8 T cells to 9.8% of CD8 T cells, and this remained the dominant clonotype during acute infection

By 32 months after diagnosis the predominant virion-encoded Gag peptide sequence mutated from KRWIIlGLNK to KGWIIlGLNK, thus thwarting binding and presentation of the peptide by HLA-B*2705

Viral escape this early is extremely unusual, the average time to development of this escape mutation in unvaccinated individuals is >9 years

Moreover, the average survival until AIDS in an HLA-B*2705 individual is >14 years
His CD4 T cell count continues to decline, presently 400 cells/μl at 32 months post infection, and viral titre remains high, despite optimal anti-retroviral therapy.

The authors raise the strong possibility that a vaccine developed according to the best notions of current immunological knowledge not only did not protect against HIV infection but accelerated development of the escape mutation in the vaccinated individual, thus hastening progression of the viral infection.

Merck AIDS Failure Hurts Global HIV Vaccine Research (Updated)

By John Laumer

Merck vaccine candidate (V520) for CD8 immunity

adenovirus type 5 vector containing gag, pol and nef

- The STEP study enrolled 3,000 HIV-negative volunteers from diverse backgrounds between 18 and 45 years of age at high risk of HIV infection
- The vaccine did not prevent infection
  - 19 developed HIV /672 vaccinated
  - 11 developed HIV /691 placebo control
- And did not reduce the amount of virus in the blood of those who became infected
  - 40,000 copies/ml in vaccine group
  - 37,000 copies/ml in placebo group

Merck & Co.'s AIDS vaccine has now joined a long list of other failed trials that have disappointed researchers worldwide. And the results, which are expected to be announced next week, will make vaccine development even more difficult for other companies. The vaccine, which collapsed after Merck announced in September that it would be testing in a large clinical trial, 性格 HIV and several other viruses. The finding only causes the U.S. to cancel a study of a similar vaccine and raise concerns about vaccine projects at GlaxoSmithKline (GSK). The Merck vaccine failed to induce a strong T-cell response in a small group of volunteers, and researchers are re-examining their vaccine strategies, said Brian Hsiu, who has studied HIV since the epidemic began.

"It's a giant step backwards," he said, adding that the vaccine could not prevent infection and would not reduce the viral load. "It's a setback for everyone else."
Basis of outcome with HLA type

**HLA-B35  RAPID PROGRESSION**

xPxxxxxy peptides recognized, if any, are in non critical parts of HIV genome permitting mutations in MHC anchor residues. Peptides weak stimulators Rapid viral replication and evolution not restrained

**HLA-B27  SLOW PROGRESSION**

xRxxxxX[KRYL] peptides recognized are often in critical parts of HIV genome and mutations not permitted in MHC anchor or TCR recognition residues Viral replication and evolution greatly slowed

An example of HIV-1 escape from a CD8 T cell clone HLA-B27 hemophiliac, infected ~1983 by blood products

CTL clone to gag p24 263-272 controlled HIV-1 replication for >10 years

<table>
<thead>
<tr>
<th>Virions/ml</th>
<th>CD4/μl</th>
<th>Gag p24</th>
<th>Tropism</th>
<th>CTL</th>
</tr>
</thead>
<tbody>
<tr>
<td>1984</td>
<td>1,800</td>
<td>K</td>
<td>R5</td>
<td>+++</td>
</tr>
<tr>
<td>1993</td>
<td>780</td>
<td>M</td>
<td>X4</td>
<td>+++</td>
</tr>
<tr>
<td>1995</td>
<td>21,400</td>
<td>M</td>
<td>X4</td>
<td>0</td>
</tr>
<tr>
<td>1996</td>
<td>530,970</td>
<td>K</td>
<td>X4</td>
<td>0</td>
</tr>
</tbody>
</table>

Kelleher, JEM 2001