Review Articles

Mechanisms of Disease

FRANKLIN H. EPSTEIN, M.D., Editor

Eosinophilia

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MARKED accumulation of eosinophils occurs in several important disorders, such as allergic diseases, parasitic infections, and cancer.1 The level of eosinophils in the body is normally tightly regulated. In normal subjects, eosinophils account for only a small minority of peripheral-blood leukocytes, and their presence in tissues is primarily limited to the gastrointestinal mucosa.² In certain disease states, however, eosinophils can selectively accumulate in the peripheral blood or any tissue in the body. Any perturbation that results in eosinophilia, defined here as an abnormal accumulation of eosinophils in blood or tissue, can have profound clinical effects. Eosinophilia may be harmful, because of the proinflammatory effects of eosinophils,3 or it may be helpful, because of the antiparasitic effects of these cells.4 This article focuses on recent advances in our understanding of the accumulation of eosinophils, as well as treatment approaches and the development of new therapeutic agents.

CLINICAL ASPECTS OF EOSINOPHILIA

Eosinophils normally account for only 1 to 3 percent of peripheral-blood leukocytes, and the upper limit of the normal range is 350 cells per cubic millimeter of blood. Eosinophilia occurs in a variety of disorders (Table 1)⁵ and is arbitrarily classified as mild (351 to 1500 cells per cubic millimeter), moderate (>1500 to 5000 cells per cubic millimeter), or severe (>5000 cells per cubic millimeter). The most common cause of eosinophilia worldwide is helminthic infections, and the most common cause in industrialized nations is atopic disease.

The differential diagnosis of eosinophilia requires

a review of the patient's history, which may reveal wheezing, rhinitis, or eczema (indicating atopic causes); travel to areas where helminthic infections (e.g., schistosomiasis) are endemic; the presence of a pet dog (indicating possible infection with Toxocara canis); symptoms of cancer; or drug ingestion (indicating a possible hypersensitivity reaction). Eosinophilia caused by drugs is usually benign but can sometimes be accompanied by tissue damage, as in hypersensitivity pneumonitis. In most cases, the eosinophilia resolves once the drug is withdrawn, but in some cases, such as the eosinophilia-myalgia syndrome due to the ingestion of contaminated tryptophan, the disease can persist despite withdrawal of the drug.6 Abnormal morphologic features of eosinophils, an increase in immature cells in the bone marrow or blood, or a karyotypic abnormality indicates the presence of eosinophilic leukemia. An accumulation of eosinophils that is limited to specific organs is characteristic of particular diseases, such as eosinophilic cellulitis (Well's syndrome), eosinophilic pneumonias (e.g., Löffler's syndrome), and eosinophilic fasciitis (Shulman's syndrome). The association of eosinophilia with vasculitis, neuropathy, and a history of asthma indicates the presence of the Churg-Strauss syndrome. In the absence of an identifiable cause of moderate-to-severe eosinophilia and in the presence of end-organ involvement, the diagnosis of the idiopathic hypereosinophilic syndrome should be considered. This disorder occurs predominantly in men and is usually a progressive, fatal disease in the absence of effective medical management.7

Diagnostic studies that should be performed in patients with moderate-to-severe eosinophilia and should be considered in patients with persistent mild eosinophilia include morphologic examination of a blood smear, urinalysis, and serial stool examinations for ova and parasites.^{5,8,9} Parasitic infections that cause eosinophilia are usually limited to helminthic parasites, with the exception of two enteric protozoans, Isospora belli and Dientamoeba fragilis. Strongyloides stercoralis infection is important to diagnose, because it can cause disseminated fatal disease in immunosuppressed patients; its detection often requires serologic testing. Other infections to rule out are those with filarial parasites, trichinosis, and T. canis infection. Bone marrow and chromosomal analysis (to detect hematologic cancer) and a tissue biopsy may be indicated.

Moderate-to-severe eosinophilia may persist in the absence of an identifiable cause or end-organ involvement. Patients with persistent, apparently be-

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TABLE 1. DISEASES ASSOCIATED WITH EOSINOPHILIA.

Type of D isease	Eosinophilia		Examples of Causes
	PERIPHERAL BLOOD	TISSUE	
Infectious	Present	Present or absent	Infections with especially invasive helminths
Respiratory	Present or absent	Present	Eosinophilic pneumonitis, asthma
Gastrointestinal	Present or absent	Present	Inflammatory bowel dis- ease, eosinophilic gastro- enteritis, allergic colitis
Allergic	Present or absent	Present	Allergic rhinoconjunctivitis, asthma, eczema
Systemic	Present	Present	Idiopathic hypereosinophil- ic syndrome, vasculitis
Iatrogenic	Present	Present or absent	Drug reaction, cytokine in- fusions (e.g. granulo- cyte-macrophage colo- ny-stimulating factor)
Malignant	Present or absent	Present or absent	Lymphoma, colonic carci- noma

nign eosinophilia usually do not need therapy, and spontaneous resolution generally occurs within several years. However, such patients should have periodic clinical and echocardiographic examinations to detect eosinophil-mediated cardiac damage, which can occur insidiously at any time and which may not be correlated with the severity of eosinophilia.8 Although no therapy is indicated, it is often helpful to determine whether eosinophilia will resolve with a short trial of prednisone (1 mg per kilogram of body weight per day for three to five days). If so, this may be a suitable option in the event of future deterioration. Glucocorticoid-resistant cases may respond to drugs that are reserved for therapy rather than trials. The therapeutic approach to patients who have eosinophilia with an identifiable cause is discussed below.

PHYSIOLOGIC FEATURES OF EOSINOPHILS

Eosinophilia occurs as a result of four processes (Fig. 1): differentiation of progenitor cells and proliferation of eosinophils in bone marrow; interactions between eosinophils and endothelial cells that involve rolling, adhesion, and migration of eosinophils; chemoattraction directing eosinophils to a specific location; and activation and destruction of eosinophils.

Proliferation

Eosinophils are produced in bone marrow from pluripotential stem cells. The latter cells differentiate first into hybrid precursors with properties of basophils and eosinophils and then into a separate eosinophil lineage.¹⁰ Three cytokines — interleukin-3, interleukin-5, and granulocyte-macrophage colonystimulating factor (GM-CSF) — are particularly important in regulating the development of eosinophils. These cytokines are encoded by closely linked genes on chromosome 5q31 and bind to receptors that have a common beta chain and different alpha chains.

Of the three cytokines, interleukin-5 (also known as eosinophil-differentiation factor) is the most specific for the eosinophil lineage and is responsible for selective differentiation of eosinophils.11 Interleukin-5 also stimulates the release of eosinophils from bone marrow into the peripheral circulation.¹² The critical role of interleukin-5 in the production of eosinophils is best demonstrated by genetic manipulation in mice. Overproduction of interleukin-5 in transgenic mice results in profound eosinophilia,11 and deletion of the interleukin-5 gene causes a marked reduction of eosinophils in the blood and lungs after an allergen challenge.13 The overproduction of one or more of the three cytokines occurs in humans with eosinophilia. Diseases involving eosinophilia without increases in other blood-cell lineages are usually accompanied by an overproduction of interleukin-5.11 The mechanisms of cytokine overproduction may involve a response of T-helper lymphocytes of the Th2 type in patients with allergic conditions (see below) or parasitic diseases,^{14,15} the malignant expansion of T-cell clones that produce interleukin-5 in some patients with lymphoma,¹⁶ or the activation of gene transcription due to a chromosomal translocation in some patients with leukemia.^{17,18}

Adhesion and Migration

The migration of eosinophils from the circulation into tissues involves a stepwise interaction between eosinophils and endothelial cells.19 The steps are mediated by adhesion molecules on endothelial cells and counter-ligands on eosinophils and are followed by the passage of eosinophils between endothelial cells (Fig. 1). Although the different types of leukocytes migrate into tissues in similar ways, their migration is mediated by different molecules. Eosinophils initially adhere to the endothelium by means of three selectins (adhesion molecules on endothelial cells) and their corresponding ligands. The rolling of circulating eosinophils on the endothelium is mediated primarily by P-selectin, whereas neutrophil rolling is mediated primarily by E-selectin.^{20,21} After cellular activation (e.g., by exposure to chemoattractants such as platelet-activating factor or eotaxin), eosinophils adhere firmly to the endothelium through adhesion molecules of the integrin family. These include the CD18 family (β_2 integrins) and very-late-antigen-4 (VLA-4) molecules (β_1 integrins). The β_2 integrins interact with intercellular adhesion molecule 1 (ICAM-1) on endothelial cells, whereas the β_1 integrins interact with vascular-cell ad-

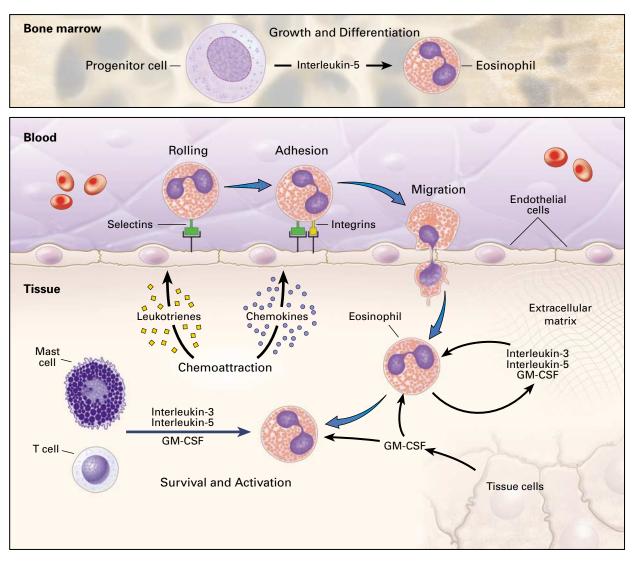


Figure 1. Processes Involved in Eosinophilia.

Eosinophils develop in the bone marrow in response to the stimulation of progenitor cells by interleukin-5. Mature eosinophils in the peripheral blood adhere to endothelial cells through the interaction of selectins and integrins (CD18 and very late antigen 4) with endothelial receptors for these molecules. On exposure to chemoattractant mediators, eosinophils undergo diapedesis between endothelial cells and migrate into the tissues. The accumulation of eosinophils is regulated by the generation of survival and activation factors (interleukin-3, interleukin-5, and granulocyte-macrophage colony-stimulating factor [GM-CSF]) by T cells and probably mast cells. In response to extracellular-matrix components, eosinophils themselves can also generate the cytokines that prolong their survival.

hesion molecule 1 (VCAM-1). Although the CD18– ICAM-1 pathway is used by all leukocytes, the VLA-4–VCAM-1 pathway is used by eosinophils and mononuclear cells but not by neutrophils. ICAM-1 is induced by a variety of proinflammatory mediators such as interleukin-1 and tumor necrosis factor α , whereas VCAM-1 is induced primarily by interleukin-4. Resting eosinophils normally express β_1 and β_2 integrins, but the level of expression of these adhesion molecules and their affinity for their appropriate endothelial receptors is increased by chemoattractants.²²

Chemoattraction

The migration of eosinophils into tissues is initiated by local chemoattractant molecules. These molecules are likely to be responsible for both physiologic homing, in which eosinophils are directed into the lamina propria of the gastrointestinal tract, and the recruitment of eosinophils into inflamed tissues. Numerous chemotactic substances act on eosinophils, including derivatives of arachidonic acid such as leukotriene B₄, other lipid mediators such as platelet-activating factor,²³ bacterial products, interleukins (e.g., interleukin-16), and various chemokines.²⁴ Although all these substances mediate the recruitment of eosinophils, most are not selective for eosinophils. However, two newly described chemokines, eotaxin-1 and eotaxin-2, are relatively specific for eosinophils.^{25,26}

Chemokines are a family of low-molecular-weight (8 kd to 10 kd) chemotactic cytokines that regulate leukocyte movement. Most chemokines interact with eosinophils by binding to a chemokine receptor (CCR-3) that is relatively restricted to eosinophils. In mice with no gene for eotaxin-1, eosinophils do not migrate to the intestinal tract; in addition, after allergen challenge, they are reduced in the lungs.²⁷ The chemoattractive effect of eotaxin is augmented by interleukin-5.^{12,28} It is remarkable that two distinct steps in the accumulation of tissue eosinophils (proliferation and chemoattraction) are regulated by molecules that are relatively specific for eosinophils and that act synergistically in promoting their accumulation.

Survival and Destruction in Tissue

Eosinophils, unlike neutrophils, can survive in tissues for extended periods (perhaps weeks), depending on the cytokines in the microenvironment.²⁹ Only eosinophils and basophils have receptors for interleukin-3, interleukin-5, and GM-CSF present on both the precursor cells in bone marrow and the circulating cells. The lifespan of tissue eosinophils is not known, but interleukin-3, interleukin-5, and GM-CSF inhibit eosinophil apoptosis for at least 12 to 14 days in vitro and in explants of allergic sinus tissue.³⁰ In contrast, eosinophils survive for less than 48 hours in the absence of these cytokines.²⁹ Tissue eosinophils can also regulate their own survival through an autocrine pathway (Fig. 1).³¹⁻³³

PATHOGENESIS OF EOSINOPHILIA IN CLINICAL DISORDERS

Moderate-to-severe eosinophilia occurs as a pathophysiologic response to infection with helminthic parasites. Eosinophilia induced by parasitic infection is dependent on interleukin-5 produced by Th2 lymphocytes (discussed below). Eosinophils participate in the immune response against helminthic parasites by discharging their cytotoxic granular contents onto the parasites, which kills them.^{4,34} However, depletion of eosinophils in mice with antibodies to interleukin-5 does not always increase their susceptibility to helminthic infections.³⁵ Since tissue eosinophilia is a hallmark of atopic diseases and eosinophils are a major effector cell in these disorders, allergic diseases serve as a prototype for understanding the pathogenesis and consequences of eosinophilia.

Genetic Aspects of Atopy

Twenty to 30 percent of people inherit a predisposition to atopy and the associated production of IgE antibodies against common environmental antigens. Several genes are likely to be responsible,³⁶ and mapping studies have identified candidate genes that include the gene for the high-affinity IgE receptor³⁷ and a locus near the genes for interleukin-4 and interleukin-5 on chromosome 5q31.^{36,38} The occurrence of eosinophilia has recently been genetically mapped to a locus near the class I genes of the major histocompatibility complex on chromosome 6.³⁹

Late-Phase Response

After exposure to allergen, many patients with allergies have a progressive clinical response that begins in three to four hours, reaches a peak at about eight hours, and subsides in several days. This process, known as the late-phase response, is accompanied by an influx of inflammatory cells containing many eosinophils (Fig. 2). The inflammatory component of the response is believed to be primarily responsible for the chronic inflammation in patients with repeated exposure to an allergen (e.g., housedust mites). Eosinophils under the control of T cells are the essential effector cells in the late-phase response.

Regulation of Eosinophils by Th2 Lymphocytes and Mast Cells

Mast cells participate in the initial events after exposure to allergen, but their importance in orchestrating eosinophilia is uncertain (Fig. 2).40 After IgEtriggered activation, mast cells may promote inflammation of the airways with eosinophils by producing proinflammatory mediators (e.g., interleukin-1 and tumor necrosis factor α) and eosinophil-directed cytokines (e.g., interleukin-4 and interleukin-5). These substances, in turn, induce chemokines that attract eosinophils. However, mast cells do not appear to be required in some animal models of allergic disease. In allergen-sensitized mice with a deficiency of mast cells and allergen-sensitized mice with a targeted deletion of the gene for IgE, recruitment of eosinophils into the lungs is not impaired after allergen challenge.41-43 In contrast, helper T lymphocytes are essential for the late-phase response, because they produce three cytokines that promote allergic responses: interleukin-4 and interleukin-13, both of which regulate IgE and VCAM-1 production, and interleukin-5. The helper cells that orchestrate this type of response are Th2 cells (Fig. 2). In contrast, Th1 cells produce interferon- γ and tumor necrosis factor β .^{14,15} Genetic factors and the conditions of antigen exposure determine the relative contribu-

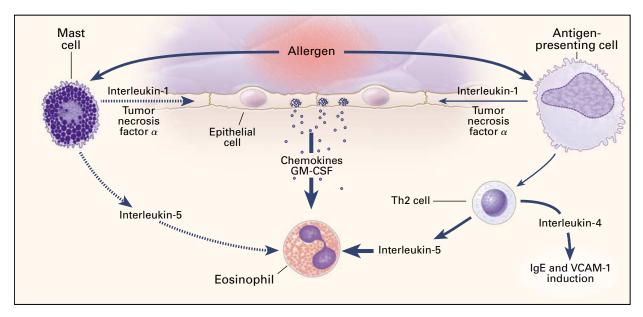


Figure 2. Events Leading to Eosinophilia during Late-Phase Responses.

After allergen exposure in sensitized subjects, two non-mutually exclusive pathways are thought to lead to the accumulation of eosinophils. In one pathway, allergen exposure results in the cross-linking of IgE receptors on mast cells and basophils and the immediate release of inflammatory mediators (histamine, prostaglandin, and leukotrienes). Mast cells then generate proinflammatory cytokines (e.g., interleukin-1 and tumor necrosis factor α) that induce respiratory epithelial cells to produce eosinophil-directed cytokines (e.g., granulocyte-macrophage colony-stimulating factor [GM-CSF]) and chemokines. In the other pathway, allergen is initially recognized by antigen-presenting cells such as dendritic cells and subsequently presented to type 2 helper T lymphocytes (Th2 cells). In contrast to mast cells, which do not appear to be required for the accumulation of eosinophils (indicated by the hatched arrows), Th2 cells are necessary for their accumulation (indicated by the solid arrows). These cells regulate allergic reactions by generating the eosinophil hematopoietin (interleukin-5) as well as interleukin-4, which induces IgE and vascular-cell adhesion molecule 1 (VCAM-1).

tions of mast cells and T cells in the regulation of eosinophils.⁴⁰ Antigen-presenting cells not only activate Th2 cells but also secrete proinflammatory mediators that induce resident cells (e.g., epithelial cells) to produce the chemokines that attract eosinophils.

Proinflammatory and Cytotoxic Effects

Once eosinophils arrive at an inflammatory focus, they may undergo apoptosis with rapid clearance by macrophages, but if they are stimulated by interleukin-3, interleukin-5, or GM-CSF, they survive for prolonged periods and have increased responsiveness to other activating agents. Eosinophils activated in this way express numerous receptors for cytokines, immunoglobulins, and complement.

Eosinophils produce unique toxic inflammatory mediators, which are stored in granules and synthesized after cellular activation. The granules contain a crystalloid core composed of major basic protein and a matrix composed of eosinophil cationic protein, eosinophil-derived neurotoxin, and eosinophil peroxidase (Fig. 3). These cationic proteins share certain proinflammatory properties but differ in other ways. For example, at concentrations similar to those in fluids from patients with eosinophilia, major basic protein, eosinophil peroxidase, and eosinophil cationic protein have cytotoxic effects on respiratory epithelium. In addition, eosinophil cationic protein and eosinophil-derived neurotoxin are ribonucleases.44,45 Eosinophil cationic protein can cause voltage-insensitive, ion-nonselective toxic pores in the membranes of target cells, and these pores may facilitate the entry of other toxic molecules.⁴⁶ Major basic protein directly increases smooth-muscle reactivity by causing the dysfunction of vagal muscarinic M2 receptors.⁴⁷ It also triggers the degranulation of mast cells and basophils. In addition, eosinophils amplify the inflammatory cascade by producing their own chemoattractants (e.g., RANTES [regulated upon activation normal T-cell expressed and secreted], eotaxin, and platelet-activating factor), which accelerate the recruitment of eosinophils into the inflammatory focus.

Further damage is caused by hydrogen peroxide and halide acids, which are generated by eosinophil peroxidase, and by superoxide, which is generated by the respiratory-burst-oxidase pathway in eosinophils. Eosinophils also generate large amounts of the cysteinyl leukotriene, leukotriene C_4 , which is me-

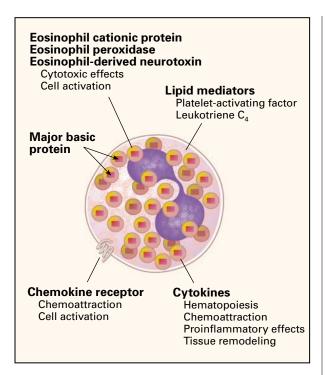


Figure 3. An Eosinophil and Its Contents.

The granules of eosinophils contain a crystalloid core composed of major basic protein and a matrix composed of eosinophil cationic protein, eosinophil peroxidase, and eosinophil-derived neurotoxin. Eosinophils also produce a variety of cytokines, some of which are stored in the granules, and lipid mediators that are generated after cellular activation. Eosinophils express one predominant chemokine receptor that interacts with multiple chemokines.

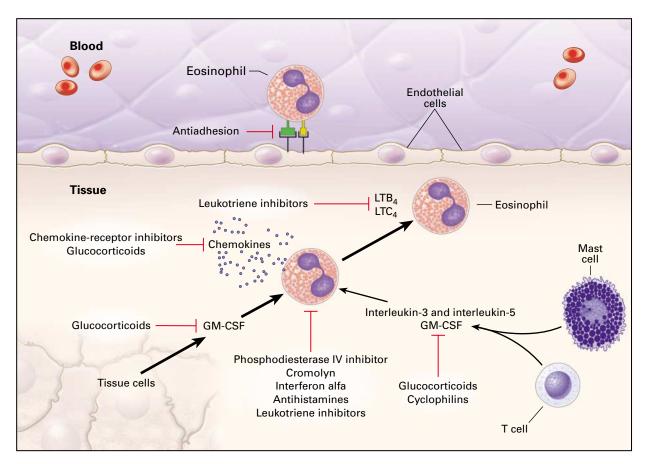
tabolized to leukotriene D_4 and leukotriene E_4 . These three lipid mediators are the slow-reacting substances of anaphylaxis that increase vascular permeability and mucus secretion and are potent stimulators of smooth-muscle contraction.⁴⁸ Lastly, activated eosinophils produce a wide range of inflammatory cytokines that have the potential to modulate multiple aspects of the immune response (Fig. 3).^{31,49}

DRUGS THAT INTERFERE WITH EOSINOPHILIA OR EOSINOPHIL PRODUCTS

Patients with eosinophilia of any magnitude who have end-organ involvement should be treated with the goal of reducing eosinophil counts or blocking the effect of eosinophil products. Numerous drugs inhibit the production of eosinophils or the production or action of their products (Fig. 4). These agents include glucocorticoids, myelosuppressive drugs, and interferon alfa (Table 2). Glucocorticoids, the most effective agents for reducing eosinophilia, suppress the transcription of a number of genes for inflammatory mediators, including the genes for interleukin-3, interleukin-4, interleukin-5, GM-CSF, and various chemokines. In addition, glucocorticoids inhibit the cytokine-dependent survival of eosinophils.⁵⁰ In most patients, treatment with systemic or topical (inhaled or intranasal) glucocorticoids causes a rapid reduction in eosinophils, but a few patients have a resistance to glucocorticoids, with persistent eosinophilia despite high doses.⁵¹ The mechanism of resistance to glucocorticoids is unclear, but a reduced level of glucocorticoid receptors and alterations in activator protein 1, a transcription factor, are at least partly responsible in some patients.^{52,53} Patients with glucocorticoid resistance sometimes require other therapy, such as myelosuppressive drugs (hydroxyurea or vincristine) or interferon alfa.^{51,54} Interferon alfa appears to be especially promising, because it inhibits the degranulation and effector function of eosinophils.55 Cyclophilins (e.g., cyclosporine) have also been used, because they block the transcription of numerous eosinophil-active cytokines (e.g., interleukin-5 and GM-CSF) (Fig. 4).56

Drugs that interfere with eosinophil chemotactic signals include recently approved leukotriene antagonists and inhibitors. The 5-lipoxygenase inhibitors (e.g., zileuton) block the rate-limiting step in leukotriene synthesis and inhibit the generation of the eosinophil chemoattractant leukotriene B4 and the sulfidopeptide leukotrienes, leukotrienes \dot{C}_4 , D_4 , and E_4 (Fig. 4).^{57,58} These drugs therefore decrease airway infiltration by eosinophils during the late-phase response.⁵⁹ Drugs (e.g., zafirlukast) that block the receptor for leukotriene D₄, which is also a receptor for leukotriene C_4 and leukotriene E_4 , prevent the muscle contraction and increased vascular permeability mediated by eosinophil-derived leukotrienes. These drugs have been found to decrease exerciseinduced bronchoconstriction and improve base-line lung obstruction in patients with asthma.⁶⁰ Some of the third-generation antihistamines (e.g., cetirizine) inhibit the vacuolization61 and accumulation62 of eosinophils after an allergen challenge and directly inhibit eosinophils in vitro.61,63 Cromolyn and nedocromil inhibit the effector function of eosinophils, such as antibody-dependent cellular cytotoxicity.63 Phosphodiesterase inhibitors raise intracellular cyclic AMP concentrations in eosinophils, and this in turn inhibits intracellular signaling, leading to decreased activation of eosinophils.52

The identification of molecules that specifically regulate the function or production of eosinophils offers new therapeutic strategies. Antibodies against interleukin-5 are especially promising, because they have been effective in animals with allergic airway disease.^{64,65} A humanized form of an antihuman interleukin-5 monoclonal antibody has been developed. Another approach involves blocking the interaction of interleukin-5 or eotaxin with its receptor. In preliminary in vitro experiments, a monoclonal





Treatment of eosinophilia involves inhibiting the interaction between eosinophils and endothelial cells through interference with the adhesion molecules used by these cells. Such approaches include the use of neutralizing antibodies against adhesion molecules such as intercellular adhesion molecule 1 or very late antigen 4. The chemoattraction process can be targeted at various steps, including interference with the synthesis or activity of leukotriene B_4 (LTB₄) and leukotriene C_4 (LTC₄), chemokine-receptor inhibitors, or G-protein inhibitors that block receptors for chemoattractant molecules. The proliferation, survival, and activation of eosinophil hematopoietins such as granulocyte-macrophage colony-stimulating ing factor (GM-CSF), interleukin-5 with glucocorticoids or cyclophilins (e.g., cyclosporine). Eosinophils are inhibited by phosphodiesterase inhibitors, cromolyn, interferon alfa, antihistamines, and leukotriene inhibitors.

antibody against eosinophil chemokine receptor 3 (CCR-3) inhibited all ligands for this receptor (Fig. 4).⁶⁶ The production of interleukin-5 may also be inhibited by modulating the immune response to allergens so that a Th2-lymphocyte response does not predominate. This has been accomplished in animals by administering interleukin-12 during allergenic sensitization.⁶⁷ Interleukin-12 inhibited the production of interleukin-4 and interleukin-5 and reduced pulmonary eosinophilia after exposure to allergen. Another molecular target interrupts the adhesion of eosinophils to the endothelium through the interaction of CD18 with ICAM-1⁶⁸ or VLA-4 with VCAM-1 (Fig. 4).^{69,70} In addition, phosphodiesterase inhibitors that are specific for leukocyte isoen-

zyme type IV are being developed.⁷¹ Lastly, lidocaine and sulfonylurea-receptor blockers have been shown to inhibit interleukin-5 activity on eosinophils, and lidocaine appeared to be promising in a preliminary clinical trial in patients with asthma.⁷²

SUMMARY

Eosinophilia occurs in a large number of diseases, and in some of them, eosinophils are the principal effector cells. The production of eosinophils involves the proliferation and differentiation of hematopoietic progenitor cells, and the accumulation of eosinophils involves interactions between eosinophils and endothelial cells, chemotaxis and cellular activation, and a balance between the survival and apoptosis of

Drugs	Mechanism of Action	
In use		
Primary		
Glucocorticoids	Inhibit transcription of eosinophil-directed cytokines	
Interferon alfa	Inhibits degranulation and effector function of eosinophils	
Myelosuppressive drugs	Suppress proliferation of eosinophils	
Secondary	- *	
Antihistamines	Inhibit degranulation and accumulation of eosinophils	
Cromolyn	Inhibits effector function of eosinophils	
Cyclosporine	Inhibits transcription of eosinophil-directed cytokines	
Leukotriene inhibitors and antagonists	Prevent synthesis of leukotrienes or block leukotriene function	
Phosphodiesterase inhibitors	Elevate intracellular cyclic AMP in eosino- phils, inhibiting intracellular signaling	
In development*		
Agents that block the CD18–ICAM-1 pathway	Inhibit adhesion of eosinophils	
Agents that block selectins	Inhibit rolling of eosinophils	
Agents that block the VLA-4–VCAM-1 pathway	Inhibit adhesion of eosinophils	
Chemokine inhibitors and antagonists	Interfere with chemotaxis and activation of eosinophils	
Interleukin-5 inhibitors and antagonists	Inhibit growth, survival, priming, and acti- vation of eosinophils	
Interleukin-12	Shifts Th2 immunity to Th1 immunity	
Lidocaine and sulfonylurea-receptor inhibitors	Inhibit eosinophil survival	
Phosphodiesterase IV inhibitors	Inhibit leukocyte-specific isoenzyme	

 TABLE 2. PHARMACOLOGIC APPROACHES TO BLOCKING EOSINOPHILIA OR THE ACTION OF EOSINOPHILS.

*ICAM-1 denotes intercellular adhesion molecule 1, VLA-4 very late antigen 4, and VCAM-1 vascular-cell adhesion molecule 1.

eosinophils. An understanding of these processes gives the clinician an insight into the pathogenesis of disorders involving eosinophils and an appreciation of the increasing number of drugs available to treat these disorders. The identification of molecules specifically involved in eosinophilia (e.g., interleukin-5 and eotaxin) offers hope for the development of new drugs that specifically target eosinophil pathways.

I am indebted to Drs. R. Wilmott, F. Finkelman, R. Hirsch, A. Assa'ad, and D. Glass for critical review of the manuscript; to Drs. E. Gelfand, P. Weller, K.F. Austen, P. Leder, and J. Rothenberg for helpful discussions; and to Dr. D.G. Nathan for the encouragement to write this article.

REFERENCES

1. Weller PF. The immunobiology of eosinophils. N Engl J Med 1991;324: 1110-8.

2. Beeken W, Northwood I, Beliveau C, Gump D. Phagocytes in cell suspensions of human colon mucosa. Gut 1987;28:976-80.

3. Gleich GJ, Adolphson CR. The eosinophilic leukocyte: structure and function. Adv Immunol 1986;39:177-253.

4. Capron M. Eosinophils and parasites. Ann Parasitol Hum Comp 1991; 66:Suppl 1:41-5.

5. Spry CJF. Eosinophils: a guide to the scientific and medical literature. Oxford, England: Oxford University Press, 1988.

6. Sternberg EM. Pathogenesis of L-tryptophan eosinophilia myalgia syndrome. Adv Exp Med Biol 1996;398:325-30.

7. Fauci AS, Harley JB, Roberts WC, Ferrans VJ, Gralnick HR, Bjornson BH. The idiopathic hypereosinophilic syndrome: clinical, pathophysiologic, and therapeutic considerations. Ann Intern Med 1982;97:78-92. **8.** Parrillo JE, Fauci AS, Wolff SM. Therapy of the hypereosinophilic syndrome. Ann Intern Med 1978;89:167-72.

9. Weller PF. The idiopathic hypereosinophilic syndrome. Blood 1994;83: 2759-79.

10. Boyce JA, Friend D, Matsumoto R, Austen KF, Owen WF. Differentiation in vitro of hybrid eosinophil/basophil granulocytes: autocrine function of an eosinophil developmental intermediate. J Exp Med 1995;182: 49-57.

11. Sanderson CJ. Interleukin-5, eosinophils, and disease. Blood 1992;79: 3101-9.

12. Collins PD, Marleau S, Griffiths-Johnson DA, Jose PJ, Williams TJ. Cooperation between interleukin-5 and the chemokine eotaxin to induce eosinophil accumulation in vivo. J Exp Med 1995;182:1169-74.

13. Foster PS, Hogan SP, Ramsay AJ, Matthaei KI, Young IG. Interleukin 5 deficiency abolishes eosinophilia, airways hyperreactivity, and lung damage in a mouse asthma model. J Exp Med 1996;183:195-201.

14. Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. Annu Rev Immunol 1989;7:145-73.

15. Robinson DS, Hamid Q, Ying S, et al. Predominant T_{H2} -like bronchoalveolar T-lymphocyte population in atopic asthma. N Engl J Med 1992; 326:298-304.

 Cogan E, Schandené L, Crusiaux A, Cochaux P, Velu T, Goldman M. Brief report: clonal proliferation of type 2 helper T cells in a man with the hypereosinophilic syndrome. N Engl J Med 1994;330:535-8.
 Grimaldi JC, Meeker TC. The t(5;14) chromosomal translocation in a

17. Grimaldi JC, Meeker TC. The t(5;14) chromosomal translocation in a case of acute lymphocytic leukemia joins the interleukin-3 gene to the immunoglobulin heavy chain gene. Blood 1989;73:2081-5.

18. Liu P, Tarle SA, Hajra A, et al. Fusion between transcription factor CBF beta/PEBP2 beta and a myosin heavy chain in acute myeloid leukemia. Science 1993;261:1041-4.

19. Resnick MB, Weller PF. Mechanisms of eosinophil recruitment. Am J Respir Cell Mol Biol 1993;8:349-55.

20. Symon FA, Walsh GM, Watson SR, Wardlaw AJ. Eosinophil adhesion to nasal polyp endothelium is P-selectin-dependent. J Exp Med 1994;180: 371-6.

21. Wein M, Sterbinsky SA, Bickel CA, Schleimer RP, Bochner BS. Com-

parison of human eosinophil and neutrophil ligands for P-selectin: ligands for P-selectin differ from those for E-selectin. Am J Respir Cell Mol Biol 1995;12:315-9.

22. Weber C, Katayama J, Springer TA. Differential regulation of $\beta 1$ and $\beta 2$ integrin avidity by chemoattractants in eosinophils. Proc Natl Acad Sci U S A 1996;93:10939-44.

23. Wardlaw AJ, Moqbel R, Cromwell O, Kay AB. Platelet-activating factor: a potent chemotactic and chemokinetic factor for human eosinophils. J Clin Invest 1986;78:1701-6.

24. Kita H, Gleich GJ. Chemokines active on eosinophils — potential roles in allergic inflammation. J Exp Med 1996;183:2421-6.

25. Jose PJ, Griffiths-Johnson DA, Collins PD, et al. Eotaxin: a potent eosinophil chemoattractant cytokine detected in a guinea pig model of allergic airways inflammation. J Exp Med 1994;179:881-7.

26. Forssman U, Uguccioni M, Loetscher P, et al. Eotaxin-2, a novel CC chemokine that is selective for the chemokine receptor CCR3, and acts like eotaxin on human eosinophil and basophil leukocytes. J Exp Med 1997; 185:2171-6.

27. Rothenberg ME, MacLean JA, Pearlman E, Luster AD, Leder P. Targeted disruption of the chemokine eotaxin partially reduces antigen-induced tissue eosinophilia. J Exp Med 1997;185:785-90.

28. Rothenberg ME, Ownbey R, Mehlhop PD, et al. Eotaxin triggers eosinophil-selective chemotaxis and calcium flux via a distinct receptor and induces pulmonary eosinophilia in the presence of interleukin 5 in mice. Mol Med 1996;2:334-48.

29. Rothenberg ME, Owen WF Jr, Silberstein DS, Soberman RJ, Austen KF, Stevens RL. Eosinophils cocultured with endothelial cells have in-

creased survival and functional properties. Science 1987;237:645-7. **30.** Simon HU, Yousefi S, Schranz C, Schapowal A, Bachert C, Blaser K.

Direct demonstration of delayed eosinophil apoptosis as a mechanism causing tissue eosinophilia. J Immunol 1997;158:3902-8.

31. Kita H. The cosinophil: a cytokine-producing cell? J Allergy Clin Immunol 1996;97:889-92.

32. Sullivan S, Broide DH. Compartmentalization of eosinophil granulocyte-macrophage colony-stimulating factor expression in patients with asthma. J Allergy Clin Immunol 1996;97:966-76.

33. Kay AB, Ying S, Durham SR. Phenotype of cells positive for interleukin-4 and interleukin-5 mRNA in allergic tissue reactions. Int Arch Allergy Immunol 1995;107:208-10.

34. Davidson RA. Immunology of parasitic infections. Med Clin North Am 1985;69:751-8.

35. Sher A, Coffman RL, Hieny S, Cheever AW. Ablation of cosinophil and IgE responses with anti-IL-5 or anti-IL-4 antibodies fails to affect immunity against Schistosoma mansoni in the mouse. J Immunol 1990;145: 3911-6.

36. Marsh DG, Neely JD, Breazeale DR, et al. Linkage analysis of IL4 and other chromosome 5q31.1 markers and total serum immunoglobulin E concentrations. Science 1994;264:1152-6.

37. van Herwerden L, Harrap SB, Wong ZY, et al. Linkage of high-affinity IgE receptor gene with bronchial hyperreactivity, even in absence of atopy. Lancet 1995;346:1262-5.

38. Rosenwasser LJ, Klemm DJ, Dresback JK, et al. Promoter polymorphisms in the chromosome 5 gene cluster in asthma and atopy. Clin Exp Allergy 1995;25:Suppl 2:74-8.

39. Daniels SE, Bhattacharrya S, James A, et al. A genome-wide search for quantitative trait loci underlying asthma. Nature 1996;383:247-50.

40. Galli SJ. Complexity and redundancy in the pathogenesis of asthma: reassessing the roles of mast cells and T cells. J Exp Med 1997;186:343-7.41. Mehlhop PD, van de Rijn M, Goldberg AB, et al. Allergen-induced

bronchial hyperreactivity and eosinophilic inflammation occur in the absence of IgE in a mouse model of asthma. Proc Natl Acad Sci U S A 1997; 94:1344-9.

42. Hamelmann E, Vella AT, Oshiba A, Kappler JW, Marrack P, Gelfand EW. Allergic airway sensitization induces T cell activation but not airway hyperresponsiveness in B cell-deficient mice. Proc Natl Acad Sci U S A 1997;94:1350-5.

43. Brusselle GG, Kips JC, Tavernier JH, et al. Attenuation of allergic airway inflammation in IL-4 deficient mice. Clin Exp Allergy 1994;24:73-80.
44. Slifman NR, Loegering DA, McKean DJ, Gleich GJ. Ribonuclease activity associated with human eosinophil-derived neurotoxin and eosinophil cationic protein. J Immunol 1986;137:2913-7.

45. Gleich GJ, Loegering DA, Bell MP, Checkel JL, Ackerman SJ, Mc-Kean DJ. Biochemical and functional similarities between human eosinophil-derived neurotoxin and eosinophil cationic protein: homology and ribonuclease. Proc Natl Acad Sci U S A 1986;83:3146-50.

46. Young JD, Peterson CG, Venge P, Cohn ZA. Mechanism of membrane damage mediated by human cosinophil cationic protein. Nature 1986;321:613-6.

47. Jacoby DB, Gleich GJ, Fryer AD. Human cosinophil major basic protein is an endogenous allosteric antagonist at the inhibitory muscarinic M2 receptor. J Clin Invest 1993;91:1314-8.

48. Lewis RA, Austen KF, Soberman RJ. Leukotrienes and other products of the 5-lipoxygenase pathway: biochemistry and relation to pathobiology in human diseases. N Engl J Med 1990;323:645-55.

49. Moqbel R, Levi-Schaffer F, Kay AB. Cytokine generation by eosinophils. J Allergy Clin Immunol 1994;94:1183-8

50. Schleimer RP, Bochner BS. The effects of glucocorticoids on human eosinophils. J Allergy Clin Immunol 1994;94:1202-13.

51. Barnes PJ, Adcock IM. Steroid resistance in asthma. QJM 1995;88: 455-68.

52. Barnes PJ. Molecular mechanisms of antiasthma therapy. Ann Med 1995;27:531-5.

53. Adcock IM, Lane SJ, Brown CR, Lee TH, Barnes PJ. Abnormal glucocorticoid receptor-activator protein 1 interaction in steroid-resistant asthma. J Exp Med 1995;182:1951-8.

54. Coutant G, Blétry O, Prin L, et al. Traitement des syndromes hyperéosinophiliques á expression myéloproliférative par l'association hydroxyurée-interféron alpha: apropos de 7 observations. Ann Med Interne (Paris) 1993;144:243-50.

55. Aldebert D, Lamkhioued B, Desaint C, et al. Eosinophils express a functional receptor for interferon alpha: inhibitory role of interferon alpha on the release of mediators. Blood 1996;87:2354-60.

56. Morley J. Cyclosporin A in asthma therapy: a pharmacological rationale. J Autoimmun 1992;5:Suppl A:265-9.

57. Kane GC, Tollino M, Pollice M, et al. Insights into IgE-mediated lung inflammation derived from a study employing a 5-lipoxygenase inhibitor. Prostaglandins 1995;50:1-18.

58. Kane GC, Pollice M, Kim CJ, et al. A controlled trial of the effect of the 5-lipoxygenase inhibitor, zileuton, on lung inflammation produced by segmental antigen challenge in human beings. J Allergy Clin Immunol 1996;97:646-54.

59. Henderson WR Jr, Lewis DB, Albert RK, et al. The importance of leukotrienes in airway inflammation in a mouse model of asthma. J Exp Med 1996;184:1483-94.

60. Gaddy JN, Margolskee DJ, Bush RK, Williams VC, Busse WW. Bronchodilation with a potent and selective leukotriene D4 (LTD4) receptor antagonist (MK-571) in patients with asthma. Am Rev Respir Dis 1992; 146:358-63.

61. Snyman JR, Sommers DK, Gregorowski MD, Boraine H. Effect of cetirizine, ketotifen and chlorpheniramine on the dynamics of the cutaneous hypersensitivity reaction: a comparative study. Eur J Clin Pharmacol 1992; 42:359-62.

62. Redier H, Chanez P, De Vos C, et al. Inhibitory effect of cetirizine on the bronchial eosinophil recruitment induced by allergen inhalation challenge in allergic patients with asthma. J Allergy Clin Immunol 1992;90: 215-24.

 Rand TH, Lopez AF, Gamble JR, Vadas MA. Nedocromil sodium and cromolyn (sodium cromoglycate) selectively inhibit antibody-dependent granulocyte-mediated cytotoxicity. Int Arch Allergy Appl Immunol 1988; 87:151-8.

64. Mauser PJ, Pitman AM, Fernandez X, et al. Effects of an antibody to interleukin-5 in a monkey model of asthma. Am J Respir Crit Care Med 1995;152:467-72.

65. Égan RW, Athwahl D, Chou CC, et al. Inhibition of pulmonary eosinophilia and hyperreactivity by antibodies to interleukin-5. Int Arch Allergy Immunol 1995;107:321-2.

66. Heath H, Qin SX, Rao P, et al. Chemokine receptor usage by human eosinophils: the importance of CCR3 demonstrated using an antagonistic monoclonal antibody. J Clin Invest 1997;99:178-84.

67. Kips JC, Brusselle GG, Joos GF, et al. Importance of interleukin-4 and interleukin-12 in allergen-induced airway changes in mice. Int Arch Allergy Immunol 1995;107:115-8.

68. Wegner CD, Gundel RH, Reilly P, Haynes N, Letts LG, Rothlein R. Intercellular adhesion molecule-1 (ICAM-1) in the pathogenesis of asthma. Science 1990;247:456-9.

69. Weg VB, Williams TJ, Lobb RR, Nourshargh S. A monoclonal antibody recognizing very late activation antigen-4 inhibits eosinophil accumulation in vivo. J Exp Med 1993;177:561-6.

70. Kuijpers TW, Mul EP, Blom M, et al. Freezing adhesion molecules in a state of high-avidity binding blocks cosinophil migration. J Exp Med 1993;178:279-84.

71. Dent G, Giembycz MA. Phosphodiesterase inhibitors: Lily the Pink's medicinal compound for asthma? Thorax 1996;51:647-9.

72. Hunt LW, Swedlund HA, Gleich GJ. Effect of nebulized lidocaine

on severe glucocorticoid-dependent asthma. Mayo Clin Proc 1996;71:361-8.