Systemic sclerosis: a prototypic multisystem fibrotic disorder

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A unique feature of systemic sclerosis (SSc) that distinguishes it from other fibrotic disorders is that autoimmunity and vasculopathy characteristically precede fibrosis. Moreover, fibrosis in SSc is not restricted to a single organ, but rather affects many organs and accounts for much of the morbidity and mortality associated with this disease. Although immunomodulatory drugs have been used extensively in the treatment of SSc, no therapy to date has been able to reverse or slow the progression of tissue fibrosis or substantially modify the natural progression of the disease. In this Review, we highlight recent studies that shed light on the cellular and molecular mechanisms underlying the fibrotic process in SSc and that identify cellular processes and intra- and extracellular proteins as potential novel targets for therapy in this prototypic multisystemic fibrotic disease.

Clinical overview

Systemic sclerosis (SSc) is a connective tissue disease of unknown etiology that is highly heterogeneous in its multisystemic clinical manifestations and follows a variable and unpredictable course. The hallmarks of SSc are autoimmunity and inflammation, widespread vasculopathy (blood vessel damage) affecting multiple vascular beds, and progressive interstitial and perivascular fibrosis (1). This constellation of seemingly disparate yet interlinked features differentiates SSc from other connective tissue diseases and organ-specific fibrosing disorders. Patients with SSc are commonly classified into two distinct subsets on the basis of the pattern of skin involvement. Diffuse cutaneous SSc, the focus of this Review, is dominated by rapidly progressive fibrosis of the skin, lungs, and other internal organs (2). By contrast, limited cutaneous SSc is dominated by vascular manifestations, and skin and organ fibrosis is generally limited and slow to progress. Although clinical outcomes have improved considerably, presumably due to better management of the complications, SSc is still considered incurable, and the diffuse cutaneous form carries the highest risk of fatality of the connective tissue diseases, with 55% survival at 10 years (3).

SSc has a worldwide distribution and is more frequent in women than men. Based on incidence and survival rates, an estimated 75,000–100,000 individuals in the United States have SSc (3). The genetics of SSc are complex, and although the disease is inherited, it is not inherited in a Mendelian fashion. Twins show a low disease concordance rate (<5%) that is similar between monozygotic and dizygotic twin pairs (4). SSc occurs substantially more frequently in families (1.6%) than in the general population (0.026%), and a positive family history represents the strongest risk factor for SSc yet identified (5). The etiology of SSc is unknown. Viruses, including human CMV, have been implicated as potential causative agents (6), along with drugs and environmental and occupational exposures to organic solvents, vinyl chloride, and silica (7). Antibodies specific for human CMV can frequently be detected in patients with SSc (8, 9). Some of these antibodies induce endothelial cell apoptosis and fibroblast activation in cell culture assays, suggesting that they have a direct pathogenic role in tissue damage (10). Furthermore, infection with human CMV is associated with enhanced production of connective tissue growth factor (CTGF), which can drive fibroblast activation and is implicated in pathological fibrosis (11).

Animal models

The pathogenesis of SSc involves a distinctive triad of small-vessel vasculopathy, inflammation and autoimmunity, and interstitial and vascular fibrosis in the skin, lungs, and multiple other organs (1). Various animal models have been investigated as spontaneous or inducible models for SSc. Although none of them reproduce all three pathogenetic components of the disease, some models do recapitulate selected phenotypic features (Table 1). The tight skin (Tsk1/+) mouse is characterized by diffuse thickening and tethering of the skin (12). Although mice homozygous for the Tsk1 mutation die in utero at 8–10 days of gestation, heterozygous (Tsk1/+) mice are viable, and develop tight skin that is firmly bound to the underlying subcutaneous tissue. In contrast to human SSc, which is characterized by thickening and sclerosis of the dermis, Tsk1/+ mice manifest hyperplasia of the subcutaneous tissue but the dermis is unaffected (13). Furthermore, Tsk1/++ mice develop emphysema-like changes in the lungs rather than fibrosis, and vasculopathy does not occur. The Tsk1 mutation is a tandem duplication in the gene encoding fibrillin-1 (14), a microfibrillar connective tissue protein; mutation of the gene encoding fibrillin-1 is also implicated in Marfan syndrome, an inherited connective tissue disorder affecting the skin, ligaments, major arteries, and heart valves, but not associated with fibrosis (14). Although the mechanisms linking the Tsk1 mutation to the fibrotic skin phenotype are currently not known, it is noteworthy that the mutation in the gene encoding fibrillin-1 in Marfan syndrome is associated with increased signaling by TGF-β, and that the mouse Marfan phenotype can be reversed by inhibiting TGF-β signaling (15). Because fibrillin-1 mediates the bioavailability of TGF-β by direct binding, or by interactions with latent TGF-β binding proteins (16), it has been suggested that the Tsk1 mouse phenotype represents tissue fibrosis due to deregulated TGF-β activation and enhanced profibrotic signaling by this cytokine.
Fibrosis can be induced in mice by subcutaneous injection of bleomycin (17). In this mouse model, the sequence of histopathological changes in the skin closely resembles that seen in SSc — early mononuclear cell accumulation and upregulated TGF-β and chemokine expression followed by dermal fibrosis with accumulation of α-SMA–expressing myofibroblasts (18, 19). The mice also manifest evidence of pulmonary and renal fibrosis. In light of its reproducibility, relative strain independence, and ease of induction, the subcutaneous bleomycin model is used with increasing frequency to investigate the roles of specific gene products in SSc-like disease (19, 20). Transplantation of MHC-mismatched BM or spleen cells into sublethally irradiated recipient mice results in skin and lung fibrosis (19, 20). Studies indicate that the failure in vasculogenesis is due to a reduction in the number of angiogenic progenitors in the BM of SSc patients compared with healthy controls (21, 22). Recently, various mouse strains with genetic modifications resulting in SSc-like phenotypes have been created. For example, mice lacking the genes encoding relaxin (23), caveolin (24), THY-1 (25), or FLI-1 (26) and transgenic mice expressing a mutant TGF-β receptor type II (TGFβRII) in fibroblasts (27) spontaneously manifest certain fibrotic or vascular features of SSc or show enhanced sensitivity to fibrosis induced by bleomycin. These transgenic and knockout mice provide robust novel experimental tools for advancing SSc research.

### Pathogenesis: an integrated view of vascular damage and autoimmunity culminating in organ fibrosis

The pathogenesis of SSc involves interplay between obliterator vascular pathology in multiple vascular beds, inflammation and autoimmunity, and progressive fibrosis (Figure 1). Vascular injury and activation are the earliest and possibly primary events in the pathogenesis of SSc (28). Histopathological evidence of vascular damage is present before fibrosis, and clinical manifestations such as Raynaud phenomenon, an episodic and reversible cold-induced vasospasm of the fingers and toes, precede other disease manifestations. Additional manifestations of SSc-associated vasculopathy include cutaneous telangiectasia, nail fold capillary alterations, and pulmonary arterial hypertension, gastric antral vascular ectasia, and scleroderma renal crisis with malignant hypertension (29). In late-stage SSc, there is a striking paucity of small blood vessels in lesional skin and other organs. Endothelial cell injury might be triggered by granzymes, endothelial cell–specific autoantibodies, vasculotropic viruses, inflammatory cytokines, or reactive oxygen radicals generated during ischemia/reperfusion (29).

Injury causes endothelial cell activation and dysfunction, altered capillary permeability, increased expression of VCAM-1 and endothelial leukocyte adhesion molecule 1, altered secretion of vasoactive mediators, and activation of platelets and fibrinolytic pathways (30). Activated endothelial cells release endothelin-1 (ET-1), a potent vasconstrictor that also promotes leukocyte adhesion to the endothelium as well as vascular smooth muscle cell proliferation and fibroblast activation. Levels of ET-1 are elevated in the blood and in bronchoalveolar lavage fluids from patients with SSc (31, 32). Microvascular injury and damage lead to vascular remodeling, with hypertrophy of the intimal and medial layers and adventitial fibrosis resulting in progressive luminal narrowing and obliteration (33). Combined with endothelial cell apoptosis, the process culminates in a striking absence of blood vessels visible on angiograms of SSc patients with late-stage disease. Loss of microvasculature is associated with tissue hypoxia, which normally induces strong expression of VEGF and its receptors (34, 35). An apparent paradox is that, in the face of tissue hypoxia and ongoing angiogenic drive, SSc is associated with a lack of angiogenesis and progressive disappearance of blood vessels. Recent studies indicate that the failure in vasculogenesis is due to a reduc-

### Table 1

<table>
<thead>
<tr>
<th>Model</th>
<th>Vasculopathy</th>
<th>Reproduces key features of SSc</th>
<th>Further details</th>
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<tbody>
<tr>
<td>Naturally occurring, spontaneous</td>
<td></td>
<td></td>
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<tr>
<td>Tsk1/+</td>
<td>–</td>
<td>–</td>
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<td>Tsk2</td>
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<td>Induced</td>
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<tr>
<td>Bleomycin</td>
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<tr>
<td>GVHD I</td>
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<tr>
<td>GVHD II</td>
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<td>Transgenic</td>
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<tr>
<td>DN TGFβRII</td>
<td>–</td>
<td>–</td>
<td>–</td>
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<tr>
<td>Conditional TGFβRI</td>
<td>+</td>
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DN, dominant-negative; GVHD, graft-versus-host disease; +, present; –, absent.

The Journal of Clinical Investigation

http://www.jci.org

Volume 117

Number 3

March 2007

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tion in BM-derived CD34+ circulating endothelial progenitor cells as well as their impaired differentiation into mature endothelial cells (36, 37). Whether the reduction in circulating endothelial progenitor cells in SSc is due to “exhaustion” of the BM, destruction of progenitor cells in the peripheral circulation, or some other mechanism(s) remains unresolved.

The innate and the adaptive immune systems both have a role in the pathogenesis of SSc (1). In patients with early-stage disease, leukocyte activation is evident in lesional tissues as well as in the peripheral blood and has a direct role in tissue injury. Perivascular infiltrates are predominantly CD3+ and CD4+ mononuclear cells that express the activation markers CD45, HLA-DR, and the IL-2 receptor and secrete fibrogenic cytokines and chemokines (38). Lesional T cells have restricted receptor specificities indicative of oligoclonal T cell expansion (39); however, it is not known whether these clones are activated nonspecifically (by cytokines or chemokines) or specifically (by unknown antigens). An emerging hypothesis for the pathogenesis of fibrotic disorders implicates an altered balance between Th1 and Th2 cytokines (40). T cells polarized toward a Th2 cell phenotype secrete abundant IL-4, IL-5, and IL-13 and demonstrate a paucity of the hallmark Th1 cytokine IFN-γ (40). Animal studies provide support for the role of a polarized immune response in the pathogenesis of fibrosis. For example, Th2-polarized cells can induce fibrosis when passively transferred in vivo (41), and mice lacking the Th1-specific transcription factor T-bet show exaggerated skin fibrosis in response to bleomycin (42). Patients with SSc display a relative shift in the Th1-Th2 cytokine balance toward Th2 predominance. As assessed by DNA microarray analysis, peripheral blood leukocytes from patients with SSc show elevated GATA3, which drives Th2 polarization (43), and CD4+ T cell clones from SSc skin biopsies show a Th2 cytokine profile (44). Furthermore, alveolar CD8+ cells show elevated production of Th2 cytokines, and the Th2 predominance predicts accelerated decline in lung function (45). In addition, proteomic analysis confirms the predominant Th2 cytokine profile in SSc bronchoalveolar lavage fluids (46).

Highly specific autoantibodies can be detected in the serum in virtually all patients with SSc. These autoantibodies also show strong association with individual disease phenotypes, and their levels fluctuate with disease activity (47). Mechanisms invoked to explain autoantibody generation in patients with SSc include molecular mimicry, chronic B cell hyperreactivity due to intrinsic B cell abnormalities, and increased expression or altered subcellular localization of potential autoantigenic peptides (48). Although SSc-associated autoantibodies have clinical utility as diagnostic markers, their contributions to disease manifestations remain uncertain. Recent studies highlight the biological activities and potential pathogenic roles of autoantibodies in patients with SSc, suggesting that antibodies specific for fibroblasts, endothelial cells, and PDGF receptors might directly cause fibroblast or endothelial cell activation and contribute to tissue damage. For example, endothelial cell–specific antibodies in SSc induce adhe-
C
B

CD19 spontaneously develop high titers of topoisomerase I–specific antibodies characteristically associated with SSc, although microarray analysis in SSc skin (52). Furthermore, circulating solute, an activated B cell signature has been demonstrated by DNA

Figure 2
Skin inflammation and fibrosis in SSc. (A) In early diffuse cutaneous SSc, moderate fibrosis in the upper dermis and at the dermal-epidermal junction is accompanied by evidence of keratinocyte hypertrophy with a flattening of the epidermis, leading to loss of reticular structure and decreased length of rete peggs (fingerlike structures that project up from the dermis and down from the epidermis, increasing the area of contact between the layers of the skin). In addition, inflammatory infiltrates are found in the dermis and near the dermal-epidermal junction, predominantly around small blood vessels. (B) Early-stage diffuse disease showing profuse dermal inflammation characterized by perivascular mononuclear cellular infiltrate composed of monocytes and activated lymphocytes, with perivascular fibrosis and loss of pericytes and vessel integrity. (C) In established fibrosis, collagen accumulation leads to dermal thickening and the deposition of dense and closely packed collagen fibers throughout the dermis, with the loss of the microvasculature and dermal structures and the dermis-subcutaneous adipose tissue interface. All images are stained with H&E and photographed using a Zeiss AxioScope confocal microscope under light field. Original magnification, ×100 (A and C); ×200 (B).

Overview: molecular and cellular determinants of the ECM
Fibrosis in SSc
Overview: molecular and cellular determinants of the ECM. Fibrosis is the most characteristic pathological hallmark of SSc, and it is especially prominent in the diffuse cutaneous form of the disease (Figure 2; ref. 57). Progressive replacement of tissue architecture by collagen-rich ECM results in functional impairment of affected organs. The fibrotic process is most prominent in the skin, lungs, gastrointestinal tract, heart, tendons and ligaments, and endocrine glands; widespread perivascular fibrosis also occurs. Fibrotic damage to these affected organs accounts for much of the morbidity and mortality associated with SSc. The ECM consists of a cellular compartment of resident and infiltrating cells and a connective tissue compartment composed of collagens, proteoglycans, fibrillins, and adhesion molecules (58). The ECM also functions as a reservoir for TGF-β, CTGF, and other growth factors and matricellular proteins that, together with the connective tissue compartment, control mesenchymal cell differentiation, function, and survival. Excessive connective tissue accumulation is due to overproduction by fibroblasts and related mesenchymal cells activated by soluble factors in an autocrine and/or paracrine manner, or by cell-cell or cell-ECM interactions. Impaired ECM degradation and turnover and expansion of the pool of mesenchymal cells in lesional tissues further contribute to ECM accumulation (58).

The most abundant ECM components are collagens, a family of proteins that consists of over two dozen structural proteins with critical roles in organ development, growth, and differentiation. The genes encoding the various collagen proteins harbor conserved regulatory elements that are specifically recognized by DNA-binding transcription factors. The complement of generalized and tissue-specific transcription factors implicated in regulating the expression of the genes encoding collagen proteins includes SP1, SMAD3, ETS1, early growth response 1, and CCAAT-binding factor, which mediate stimulation, and SP3, CCAAT/enhancer binding protein, Y box-binding protein 1, c-KROX, and FLI-1, which suppress transcription (59). These transcription factors not only interact with one another but also with cofactors and chromatin-modifying enzymes such as p300/CREB-binding protein (p300/CBP), p300/CBP–associated factor, and histone deacetylases that are recruited to target genes by DNA-bound transcription factors. The relative levels, intracellular activities, and interactions among transcription factors and cofactors are controlled by extracellular cues, and alterations in these contribute to persistent fibroblast activation in SSc (60).

Cellular determinants of fibrosis: fibroblasts, myofibroblasts, pericytes, and transdifferentiation. Fibroblasts are key effectors of the fibrotic process. Soluble mediators generated in the local cellular microenvironment by platelets, endothelial cells, epithelial cells, and...
inflammatory cells provide cues that induce fibroblasts to secrete collagens and other ECM macromolecules to adhere to, contract, organize, and remodel connective tissue; to secrete and activate growth factors and cytokines; and to undergo transdifferentiation into contractile myofibroblasts (61). Together, these biosynthetic, contractile, and adhesive functions enable fibroblasts to mediate effective wound healing. Under normal circumstances, the fibroblast repair program is self limited, but pathological fibrotic responses are characterized by sustained and amplified fibroblast activation. Inappropriate fibroblast activation is the fundamental pathogenetic alteration underlying fibrosis in SSc. Recent DNA microarray studies have revealed that fibroblasts from different anatomic locations differ markedly in their gene expression patterns and show site-specific variations in their transcriptional profiles that seem to be related to their location within the body (62). Moreover, these variations in gene expression are retained and might be considered the “positional memory” of these fibroblasts (62). It will be important to investigate how alterations in fibroblast positional memory might contribute to inappropriate fibroblast activation and fibrosis.

The tissue pool of biosynthetically activated mesenchymal cells contributing to ECM accumulation and remodeling is expanded in fibrosis by proliferation and migration of resident fibroblasts, by in situ transdifferentiation of other cell types, and by the influx of mesenchymal progenitor cells from the circulation (Figure 1; ref. 1). Myofibroblasts are specialized fibroblasts that show features of smooth muscle cell differentiation, having prominent cytoplasmic stress fibers with α-SMA. Myofibroblasts synthesize collagens and other ECM components and are a major source of TGF-β during the fibrotic response (61). In normal wound healing, myofibroblasts can be detected transiently in the granulation tissue; their removal by apoptosis is a crucial step in wound resolution (63). By contrast, myofibroblasts persist in pathological fibrogenesis, resulting in excessively contracted ECM characteristic of chronic scar (63). Myofibroblast transdifferentiation from normal fibroblasts can be induced by TGF-β and requires coexpression of the α-SMA variant ED-A fibronectin. The presence of α-SMA+ myofibroblasts expressing ED-A fibronectin and THY-1 is strongly associated with SSc but is absent from normal skin (64, 65). Pericytes are smooth muscle–like structural cells that are found normally in the walls of small blood vessels and have key roles in microvascular integrity and function (66). In early-stage SSc, pericytes are activated and express receptors for PDGF and THY-1, all of which are features of wound healing (65). Under certain conditions, epithelial cells can undergo transformation to fibroblasts (67). Epithelial-mesenchymal transition (EMT) has

**Figure 3**
Profibrotic signaling by TGF-β through SMAD-dependent pathways. The ECM serves as a reservoir for latent TGF-β, which is maintained in an inactive form by latent TGF-β binding proteins (LTBPs). Upon activation, TGF-β binds to its cell surface receptors and triggers SMAD-mediated intracellular signaling. Activated SMAD proteins accumulate in the nucleus and bind to conserved SBE regulatory elements in target genes, recruit coactivators and chromatin-modifying enzymes such as p300/CBP to the DNA, and induce mRNA synthesis and cellular responses. Inhibitory SMAD7 blocks ligand-induced SMAD protein phosphorylation and shuts down SMAD-mediated signaling.
a vital role during embryonic development. The process of EMT can be induced by TGF-β in cultured alveolar epithelial cells and suppressed by bone morphogenetic protein-7. Pathological EMT has been documented in cancer as well as in renal and pulmonary fibrosis. To date, the role of EMT in the pathogenesis of tissue fibrosis in SSc has not been examined.

The BM can serve as a source of fibroblast progenitors. Circulating BM-derived mesenchymal progenitors continuously replenish the resident fibroblast population as part of normal tissue homeostasis and might have a role in fibrosis (61). Fibrocytes are CD34+ BM-derived precursors normally present in small numbers in the peripheral blood (68). Fibrocytes synthesize collagen and express CD14 (a monocyte marker) as well as chemokine receptors CC chemokine receptor 3 (CCR3), CCR5, and CX3C chemokine receptor 4 (CXCR4). The cell surface expression of chemokine receptors allows fibrocytes to respond to chemokine gradients and to traffic to and accumulate in specific tissues. Stromal cell–derived factor-1 (SDF-1, also known as CXCL12) is an important chemokine regulator of progenitor trafficking, and its expression, as well as that of its receptor, CXCR4, is elevated in lesional skin in early SSc (69). The pathogenic role for fibrocyte trafficking into lesional tissue has been established in animal models using neutralizing antibodies and in mice genetically deficient in CXCR4 (70). Other studies have identified multipotent monocyte-derived mesenchymal progenitors in peripheral blood (71). The roles of pericytes, fibrocytes, and other monocyte-derived circulating fibroblast progenitors in the pathogenesis of fibrosis in SSc remain to be fully elucidated.

**Molecular determinants of fibrosis: TGF-β.** The expression of ECM genes is normally tightly regulated by paracrine and/or autocrine actions of soluble mediators as well as by cell-cell contact, hypoxia, and contact with the surrounding ECM (59). Of the multiple cytokines implicated in SSc, TGF-β is considered to be the master regulator of both physiological (wound healing and tissue repair) and pathological (scar) fibrogenesis (60). In addition, TGF-β has essential roles in normal tissue repair, angiogenesis, immune regulation, cell proliferation, and cell differentiation (72). TGF-β is secreted by platelets, monocytes/macrophages, T cells, and fibroblasts. Most cell types express specific cell surface receptors for TGF-β and secrete TGF-β as a latent complex that is sequestered in the ECM, in part by an interaction with fibrillin-1 (73). Activation of latent TGF-β to its biologically active form capable of inducing cellular responses can be mediated by integrins, thrombospondins, THY-1, or plasmin (72).

The responses elicited by TGF-β are context dependent and specific for target cell lineage. In mesenchymal cells, TGF-β functions as a potent fibrogenic stimulus by enhancing collagen synthesis, proliferation, migration, adhesion, and transdifferentiation into myofibroblasts (72). Binding of TGF-β to the TGFBR1 triggers an intracellular signal transduction cascade that leads to the induction of target genes (74). The evolutionarily conserved canonical TGF-β signal transduction pathway involves phosphorylation of TGFBR1 (also known as ALK5), a transmembrane serine-threonine kinase that in turn phosphorylates SMAD proteins (Figure 3). Ligand-induced phosphorylation of SMAD2 and SMAD3 allows them to form heteromers with SMAD4 and translocate from the cytoplasm into the nucleus, where they recognize and bind to a cis-acting DNA sequence (CAGAC) that defines the consensus SMAD-binding element (SBE). Upon SBE binding, activated SMAD proteins recruit transcriptional cofactors to the DNA, resulting in gene transcription. The conserved SBE is found in the promoters of many TGF-β–inducible genes, including type I collagen, plasminogen activator inhibitor-1, α-SMA, and CTGF. Ligand-induced signal transduction through the SMAD proteins is tightly controlled by endogenous inhibitors such as SMAD7. Deregulated expression or function of activating and inhibitory SMAD proteins and their cofactors has been documented in SSc fibroblasts (1, 58, 60) and might contribute to the initiation or propagation of the abnormal fibrogenic response (Table 2).

Although the SMAD pathway is considered to be the central conduit for signals from the TGF-β receptors, emerging evidence highlights the importance of non-SMAD pathways (75). Non-SMAD molecules activated by TGF-β include protein kinases (the MAPKs p38 and JNK, focal adhesion kinase [FAK], and TGF-β activated kinase 1), lipid kinases (such as PI3K and its downstream target, AKT), and the calcium-dependent phosphatase calcineurin (Figure 4). Recent studies indicate novel roles for the tyrosine kinase c-ABL (76) and early growth response 1 (77) in mediating the stimulation of profibrotic responses induced by TGF-β in fibroblasts. These novel non-SMAD pathways interact with each other and with SMAD proteins in complex, cell lineage–specific signaling networks.

**Molecular effectors of fibrosis: cytokines, growth factors, and chemokines.** In addition to TGF-β, an expanding list of cytokines, growth factors, and chemokines that regulate mesenchymal cell function has been found to be overexpressed or abnormally regulated in patients with SSc. These soluble factors contribute to the pathogenesis of fibrosis and represent attractive potential targets for therapy. CTGF is a cysteine-rich modular protein belonging to the Cyr61/CTGF/NOV (CCN) family of matricellular growth factors, all of which function as adaptor molecules connecting the cell surface to the ECM (78). The CCN proteins have adhesive abilities that control cell attachment and migration, modulate the activi-

### Table 2

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Alteration in SSc fibroblast</th>
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<tr>
<td>SMAD2/3</td>
<td>Constitutive nuclear accumulation and phosphorylation</td>
</tr>
<tr>
<td>SMAD7</td>
<td>Reduced expression and/or function</td>
</tr>
<tr>
<td>SP1</td>
<td>Constitutive phosphorylation</td>
</tr>
<tr>
<td>Fli-1</td>
<td>Reduced expression</td>
</tr>
<tr>
<td>p380/CBP</td>
<td>Elevated expression, constitutive binding to SMAD2/3</td>
</tr>
<tr>
<td>PPARγ</td>
<td>Reduced expression</td>
</tr>
<tr>
<td>TGF-β receptors</td>
<td>Elevated expression of TGFβRI and/or TGFβRII; altered ratio of TGFβRI and TGFβRII</td>
</tr>
<tr>
<td>PDGF receptors</td>
<td>Both PDGFR-α and PDGFR-β elevated</td>
</tr>
<tr>
<td>Integrin αvβ3</td>
<td>Elevated</td>
</tr>
<tr>
<td>Integrin αvβ5</td>
<td>Elevated</td>
</tr>
<tr>
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<td>Elevated</td>
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<tr>
<td>ERK</td>
<td>Constitutively active</td>
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<td>FAK1</td>
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ties of TGF-β and other cytokines, and regulate cell differentiation, proliferation, apoptosis, and ECM synthesis (78). Although expressed at only very low levels in normal tissues, expression of CTGF is induced by TGF-β as well as ET-1 and hypoxia (78). The levels of CTGF are markedly elevated in lesional tissues from patients with SSc (79) and in mouse models of scleroderma (19). Because many activities of CTGF parallel those induced by TGF-β, it has been suggested that at least some TGF-β responses are mediated through endogenous CTGF (78). The identity of the fibroblast receptor for CTGF and the mechanism of action underlying profibrotic responses elicited by this matricellular protein are still incompletely characterized. PDGF, which is produced by macrophages, endothelial cells, and fibroblasts in addition to platelets, is a potent mitogen and chemoattractant for fibroblasts and can induce them to synthesize collagen, fibronectin, and proteoglycans and to secrete TGF-β1, monocyte chemoattractant protein 1 (MCP-1), and IL-6. The expression of PDGF and its receptors is elevated on SSc fibroblasts and in lesional tissues (80) and in bronchoalveolar lavage fluid of SSc patients (81). Autoantibodies specific for the PDGF receptor can be detected in serum from patients with SSc (51). These antibodies have been shown to induce activation of normal fibroblasts in vitro through PDGF receptor-mediated activation of the intracellular Ha-Ras–ERK1/2 cascade and generation of ROS (51).

The cytokines IL-4 and IL-13 have major roles in the pathogenesis of Th2 cell–mediated diseases such as asthma and in the pathogenesis of fibrotic disorders (40). In normal fibroblasts, IL-4 stimulates proliferation, chemotaxis, and collagen synthesis and enhances the production of TGF-β and CTGF (82). Levels of IL-4 are elevated in the sera of patients with SSc (83), and the number of IL-4–producing T cells is increased in their peripheral blood (84, 85). Expression of IL-4 protein and mRNA is markedly elevated in SSc lesional skin and cultured fibroblasts (86). The profibrotic activities of IL-13 involve both direct fibroblast activation and indirect mechanisms due to stimulation of TGF-β (87, 88). Although levels of IL-13 are elevated in the serum of patients with SSc, its role in inducing and maintaining tissue fibrosis in SSc remains to be determined. Chemokines have a broad range of cellular targets and biological activities and are increasingly implicated in fibrosis (60). In vitro, MCP-1 stimulates collagen production directly or indirectly through the induction of endogenous TGF-β and IL-4 (89). Levels of MCP-1 and of multiple other chemokines are elevated in the serum and bronchoalveolar lavage fluid from patients with SSc and correlate with the severity of fibrosis (90, 91). The expression of MCP-1 and MCP-3 and of their receptors is markedly elevated in lesional skin and cultured fibroblasts from patients with SSc, particularly in early-stage disease (92, 93).

Figure 4
TGF-β signaling through non-SMAD pathways. Receptor activation by TGF-β can cause activation of non-SMAD pathways involved in regulating cell proliferation, cytoskeletal rearrangement, ECM synthesis, and apoptosis. Activation of intracellular protein and lipid kinase cascades results in activation of DNA-binding transcription factors and regulation of gene expression. These signal transduction pathways might converge on or interact with the canonical SMAD pathway or operate completely independent of SMAD pathways. AP-1, activator protein 1; EGR-1, early growth response 1; PAK2, p21-activated kinase 2; ROCK, Rho-associated, coiled-coil containing protein kinase 1; TAB1/2, TAK1-binding protein 1/2; TAK1, TGF-β activated kinase 1.
Endogenous mechanisms for attenuating the fibrotic response. To avoid excessive scarring following injury, redundant biological mechanisms have evolved to suppress ECM synthesis and fibroblast proliferation and differentiation and thereby slow the fibrotic process. Appropriate temporospatial utilization of these mechanisms allows repair of damaged tissue without the development of fibrosis. The prototypic Th1 cytokine IFN-γ suppresses the expression of the genes encoding the collagen proteins and abrogates the stimulatory effects of TGF-β (94, 95). IFN-γ is also a potent inhibitor of fibroblast proliferation, ECM contraction, and fibroblast transdifferentiation to myofibroblasts (60). It has been shown that IFN-γ suppresses fibrotic responses in vivo and in vitro (96) and therefore might have a physiological function as an endogenous natural antifibrotic mediator. Although some studies suggest that IL-10 (alone or in combination with IFN-γ) also functions as a repressor of the fibrotic process (97), the role of IL-10 in SSc remains to be clarified.

Fibroblasts are equipped with endogenous molecules that can suppress the stimulation of ECM gene expression. For example, by blocking SMAD-mediated TGF-β signal transduction, SMAD7 limits the intensity and duration of the TGF-β response and prevents prolonged stimulation of collagen synthesis by normal fibroblasts (98). Impaired SMAD7 expression or function has been demonstrated in SSc fibroblasts (99). Additional molecules that function as intrinsic repressors of basal or inducible collagen synthesis include the transcription factors SP3, FLI-1, p53, RAS, and the nuclear hormone receptor PPARγ (100–103). Diminished expression or function of these endogenous inhibitors, or their impaired responsiveness to extracellular ligands, could contribute to excessive and sustained ECM upregulation in fibrosis.

SSc fibroblasts: autonomously activated effector cells in fibrosis. Over 3 decades ago, E.C. LeRoy demonstrated that fibroblasts explanted from lesional skin or the fibrotic lungs of patients with SSc displayed an abnormal activated phenotype that persisted for several passages in vitro (104). This seminal observation focused attention on the pivotal role of the fibroblast in the pathogenesis of fibrosis and spawned extensive research into the underlying mechanisms (105). Persistent fibroblast activation in the absence of the fibrotic environment (106, 107), indicates autonomous, signal-independent alterations in cell function. The SSc phenotype is characterized by enhanced ECM synthesis, constitutive secretion of cytokines and chemokines, and increased expression of cell surface receptors for fibrogenic signaling mediators (60). Moreover, SSc fibroblasts show evidence of myofibroblast transdifferentiation, due in part to constitutive activation of FAK (108). Elevated expression of the prosurvival factors BCL2 and AKT in fibroblasts from patients with SSc might account for their relative resistance to apoptosis (109–111). Because most of the SSc fibroblast characteristics can be induced in normal fibroblasts by incubation with TGF-β, it has been proposed that the SSc phenotype is due to autocrine TGF-β signaling (60). The levels of receptors for TGF-β are elevated on SSc fibroblasts, permitting them to mount a robust response to endogenously produced TGF-β or to subthreshold levels of exogenous TGF-β in their environment (112, 113). Furthermore, thrombospondin and αβ integrins, both of which can mediate latent TGF-β activation, are elevated on SSc fibroblasts (114, 115). Consistent with the autocrine TGF-β hypothesis, SSc fibroblasts show evidence of ligand-independent intracellular TGF-β signaling, with elevated expression and nuclear accumulation of activated SMAD3 (99, 116) and its constitutive interaction with the p300/CREB coactivator (117, 118). Other studies have demonstrated defective expression of function of endogenous suppressors of TGF-β signaling such as FLI-1 and SMAD7 (27, 98, 119), and of PPARγ (J. Varga and M. Whitfield, unpublished observations).

However, fibroblast activation mediated by an autocrine TGF-β–SMAD signaling pathway cannot fully account for all of the phenotypic hallmarks of SSc fibroblasts, such as constitutive production of CTGF and ET-1 (120). Furthermore, small molecule inhibitors of TGFβRI-dependent SMAD signaling fail to fully normalize SSc fibroblasts in vitro (113, 121, 122). These recent observations therefore imply that SMAD-independent TGF-β signaling mechanisms, TGF-β–independent activation events, or both also have a role in inducing or maintaining the SSc phenotype. The autonomous phenotype of SSc fibroblasts might also result from altered integrin signaling from the surrounding ECM (123). Recent evidence suggests that epigenetic regulation in SSc fibroblasts might contribute to their persistent dysfunction (119). For example, the gene encoding FLI-1, an important negative regulator of collagen synthesis, seems to be constitutively silenced in SSc fibroblasts through a mechanism that involves DNA methylation or chromatin histone deacetylation (119). Silencing the expression of this negative regulator is associated with increased collagen synthesis (119).

Conclusions and perspectives

Vascular injury and damage, along with autoimmunity and inflammation, are prominent early manifestations of SSc that precede the onset of fibrosis and contribute to its pathogenesis in poorly understood ways. Vascular damage and tissue fibrosis are widespread in SSc and largely account for the protean clinical manifestations and substantial morbidity and mortality. Current therapies such as cyclophosphamide, azathioprine, mycophenolate, d-penicillamine, and methotrexate are primarily immunomodulatory in nature. These agents have shown little or no effect on reversing fibrosis or slowing its progression or in altering the natural progression of the disease. Effective therapy will require directly targeting the fibrotic process and will necessitate an improved understanding of the roles of individual cell types and their products in the development of fibrosis. Despite substantial progress during the past decade in uncovering the molecular and cellular mechanisms underlying pathological fibrogenesis, translation of basic research findings into more effective management of patients with SSc has been slow. To identify novel therapeutic targets, additional studies are necessary to delineate the intracellular signaling pathways regulating the physiologic expression of ECM genes and their dysregulation in SSc as well as the factors governing mesenchymal cell transdifferentiation into activated myofibroblasts and accumulation of circulation-derived progenitors in the lesional tissues during the fibrotic process. Genetic analysis to identify determinants that influence disease susceptibility and clinical phenotypes will also be needed. Clinical evaluation of targeted drugs and interventions for SSc is critically dependent on the availability of simple and reliable biomarkers for assessing disease severity and progression and therapeutic response. In addition to novel therapies that block key mediators in fibrosis, such as TGF-β, CTGF, ET-1, and chemokines, or that inhibit the activities or interactions of intracellular signaling molecules such as TGFβRI, FAK, and p300/CBP, several drugs already in use for the treatment of other conditions might turn out to have substantial unexpected utility in SSc. Such drugs include the c-ABL kinase inhibitor ima-
tininb (used to treat chronic myelogenous leukemia), statins (used to treat hyperlipidemia and cardiovascular disease), the angioten-
sin II receptor blocker losartan (used to treat hypertension), and the insulin-sensitizing thiazolidinedione PPARγ agonists (used to treat type 2 diabetes). Each of these agents has been shown to have vasculoprotective, antiinflammatory, and antifibrotic activities over and above their kinase-blocking, lipid-lowering, hypotensive, and insulin-sensitizing activities, respectively, and therefore should undergo rigorous evaluation for the treatment of SSc.

Acknowledgments
This work was supported by grants from the NIH (AR42509 and AR49025).

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