Critical Review

The Tumor Radiobiology of SRS and SBRT: Are More Than the 5 Rs Involved?

J. Martin Brown, PhD,* David J. Carlson, PhD,[†] and David J. Brenner, PhD[‡]

*Department of Radiation Oncology, Stanford University School of Medicine, Stanford, California; [†]Department of Therapeutic Radiology, Yale University School of Medicine, New Haven, Connecticut, and [‡]Center for Radiological Research, Columbia University Medical Center, New York, New York

Stereotactic radiosurgery (SRS) and stereotactic body radiation therapy (SBRT), also known as stereotactic ablative radiation therapy (SABR), are rapidly becoming accepted practice for the radiation therapy of certain tumors. Typically, SRS and SBRT involve the delivery of 1 or a few large-dose fractions of 8 to 30 Gy per fraction: a major paradigm shift from radiation therapy practice over the past 90 years, when, with relatively large amounts of normal tissues receiving high doses, the goal was to maximize tumor response for an acceptable level of normal tissue injury. The development of SRS and SBRT have come about because of technologic advances in image guidance and treatment delivery techniques that enable the delivery of large doses to tumors with reduced margins and high gradients outside the target, thereby minimizing doses to surrounding normal tissues. Because the results obtained with SRS and SBRT have been impressive, they have raised the question whether classic radiobiological modeling, and the linear-quadratic (LQ) model, are appropriate for large doses per fraction. In addition to objections to the LQ model, the possibility of additional biological effects resulting from endothelial cell damage, enhanced tumor immunity, or both have been raised to account for the success of SRS and SBRT. In this review, we conclude that the available preclinical and clinical data do not support a need to change the LQ model or to invoke phenomena over and above the classic 5 Rs of radiobiology and radiation therapy, with the likely exception that for some tumors high doses of irradiation may produce enhanced antitumor immunity. Thus, we suggest that for most tumors, the standard radiobiology concepts of the 5 Rs are sufficient to explain the clinical data, and the excellent results obtained from clinical studies are the result of the much larger biologically effective doses that are delivered with SRS and SBRT. © 2014 Elsevier Inc.

Introduction

Stereotactic radiosurgery (SRS) and stereotactic body radiation therapy (SBRT), also known as stereotactic ablative radiation therapy (SABR), are novel and increasingly popular ways of delivering radiation therapy. SRS, usually limited to brain lesions, is an extreme example of SBRT in that the entire dose is typically given in a single fraction. SBRT is defined as treatment of tumors outside the brain with 1 to 5 dose fractions. The generally outstanding results already obtained with SRS and SBRT, together with certain preclinical data, have led to the suggestion that the large single doses of SRS, or high doses per fraction in SBRT, produce greater antitumor efficacy than would be predicted from the survival curves of the tumor cells or from the accumulated clinical experience with fractionated radiation therapy. We shall critically examine these claims using both preclinical and clinical data.

However, we must first consider why single-dose radiation therapy can even be considered, given that it is a major paradigm shift from the practice of radiation therapy that has developed over the past 90 years, when the goal was to maximize tumor response for an acceptable level of normal tissue injury. It is uncontested

Reprint requests to: J. Martin Brown, PhD, Division of Radiation and Cancer Biology, Department of Radiation Oncology, Stanford University, A246, 1050A Arastradero Rd, Palo Alto, CA 94304-1334. Tel: (650) 723-5881; E-mail: mbrown@stanford.edu

that fractionation of the radiation dose is superior to single doses in achieving such differential sparing of normal tissue compared with tumor. The reason why SRS and SBRT can essentially ignore this classic fractionation paradigm is the result of technologic advances in image guidance and treatment delivery techniques that enable the delivery of large doses to tumors with reduced margins with high gradients outside the target, thereby minimizing doses to relatively large volumes of surrounding normal tissue. This practice has now raised the question whether these large doses per fraction produce greater antitumor efficacy than predicted by classic radiobiology, or the 5 Rs.

Factors Affecting Tumor Response to Irradiation (the 5 Rs)

Loss of reproductive ability caused by double strand breaks (DSB) in DNA is the primary means by which radiation kills cells: any cell that is incapable of reproducing indefinitely is by definition considered dead, although it may still be metabolically active for some time. The response of tumors to radiation has therefore been largely characterized in terms of factors that influence the ability of radiation to damage DNA and that affect a population of cells in tumors to recover from such damage.

Almost a century of research on the biological basis of radiation therapy has revealed 5 factors that are critical in determining the net effect of radiation therapy on tumors. They are as follows:

1. Repair of sublethal cellular damage
2. Repopulation of cells after radiation
3. Redistribution of cells within the cell cycle
4. Reoxygenation of the surviving cells
5. Radiosensitivity (intrinsic)

The first 4 of these factors were described initially by Withers (1), but the list was subsequently increased to 5 by Steel et al (2) on the basis of emerging data that the responsiveness of tumors to radiation therapy correlated with the intrinsic radiosensitivity of the cells in vitro. These 5 factors can work in opposite directions depending on the particular tumor and the way in which the radiation is delivered. For example, if a given dose of radiation is divided into a set of (typically daily) fractions, redistribution and reoxygenation facilitate increased overall cell kill by redistributing the resistant survivors into more sensitive states over time. However, repair and repopulation produce increased cell survival by allowing for recovery of cells after individual radiation doses and by allowing proliferation between radiation doses. Modern fractionation schemes are based on manipulating these effects so as to maximize tumor cell kill while avoiding normal tissue toxicities, particularly those arising in late-responding tissues. Tumors are generally considered early-responding tissues, although, given the heterogeneity of neoplastic tissues, this is not universal.

For many years after the publication of the first mammalian radiation survival curve by Puck and Marcus (3) using colony formation as the criterion for cell survival, investigators fitted survival curve data using the multitarget model, which describes cell survival (S) in terms of the dose (D), a parameter D_0, representing the slope of the exponential portion of the curve, and the extrapolation number n as follows:

$$S = 1 - \left(1 - e^{-D/D0}\right)^n$$

This equation produced a good fit to most experimental data over a wide range of cell killing. However, the equation does not describe any realistic cell biology for cell killing by irradiation. Nonetheless, it is notable for the fact that experimental data obtained over 2 to 3 logs of survival can be readily extrapolated to the very high levels of cell kill expected for doses of 15 to 30 Gy given in SRS because the curve quickly becomes a straight line on a semilog plot, so extrapolation to high doses is simple and reasonable.

However, when investigators developed techniques to measure cell killing at low radiation doses (producing 80% to 90% survival), they quickly found that this equation, which predicts zero cell killing at small radiation doses (zero initial slope to the survival curve) did not fit the experimental data at these low doses. This, plus the fact that another model, the linear quadratic (LQ) model, which derives from biological considerations of how cells could be killed by ionizing radiation, did fit the data at low doses, led to the replacement of this equation by the LQ equation as follows:

$$S = e^{-\left(\alpha D + \beta D^2\right)}$$

This equation represents a model in which cell killing is caused by either 1 or 2 radiation tracks (4, 5), which is consistent with extensive experimental data showing that cells die by chromosome breakage producing a dicentric along with an acentric fragment (which require breaks in 2 adjacent chromosomes) or by terminal deletions. Furthermore, the model has become successfully used by the radiation oncology community to calculate changes in dose per fraction or in number of fractions to achieve the same radiation effects on normal tissues as a standard fractionation regimen. The only parameter needed to perform these calculations is the value of α/β, which, based on extensive preclinical and clinical data, is typically considered to be ~ 3 Gy for late-responding tissues and ~ 10 Gy for early-responding tissues, including most tumors, although especially for tumors there is much uncertainty in the values and α/β can be very high, particularly for lung tumors (6).

So successful has been the LQ model that it has been used as the basis of clinical trials of hyperfractionation based on the predicted superiority of regimens with small doses per fraction
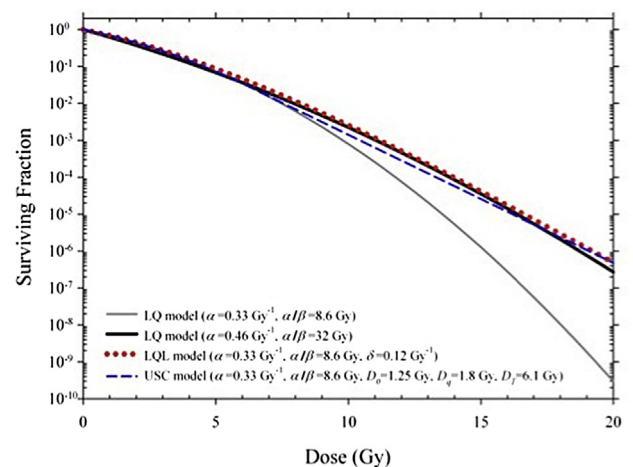
Fig. 1. The perceived overprediction of cell killing at high doses by the LQ model is resolved by assuming a higher α/β value Comparison of predictions of the linear quadratic (LQ), linear-quadratic-linear (LQL) (14, 22), and universal survival curve (USC) (17) models. LQL and USC model predictions are similar assuming an α/β of 8.6 Gy.
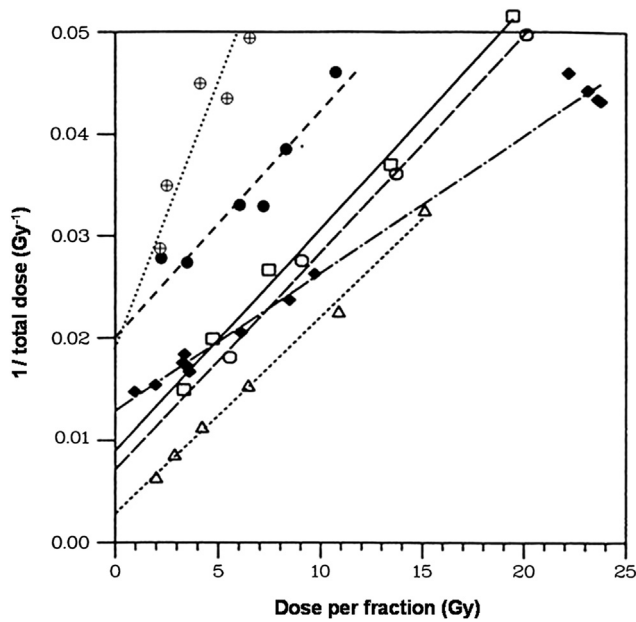
Fig. 2. Isoeffect data for response in normal tissues fit the linear quadratic model. Data for different regions (□, O, Δ) of the rat spinal cord (24), for acute skin reactions (◆) in mice (25), and for early (●) and late (O+) murine intestinal damage (26). The LQ model predicts straight lines for these plots. From (15) with permission.

(<2 Gy) in terms of reducing late effects for the same level of early effects (including tumor response) (7). Typical among such trials is a European head and neck trial (EORTC 22791), in which treatment with a 7-week course consisting of twice-daily 1.15-Gy fractions to a total dose of 80.5 Gy was compared with a conventional 7-week course administering 2 Gy daily for 5 days a week (8). As predicted by the LQ model, late effects between the

groups were comparable, whereas acute reactions were elevated (but manageable), and the 5-year local control rate was significantly higher in the hyperfractionated arm.

In general, clinical results have validated the LQ model over a modest dose range of 1 to 5 Gy per fraction and in particular the lower α/β ratio for late-responding normal tissues relative to acute effects and most tumors, with the likely exception of prostate cancer (9, 10). However, implicit in modeling of tumor response by the LQ equation is that full reoxygenation occurs between each dose fraction. It therefore seems difficult to justify large single doses of radiation because the LQ equation, and the detrimental effect of the lack of interfraction reoxygenation, would predict that such doses would produce far less tumor cell kill for the same level of normal tissue damage (11). However, there is little doubt that the clinical results with SBRT, particularly for early-stage non-small cell lung cancer (NSCLC), have been impressive (12). Possible reasons for the efficacy of high dose per fraction radiation therapy are these:

1. Advances in image guidance and dose delivery enable the delivery of large doses to tumors with much smaller volumes of normal tissue irradiated, thus overcoming the need in some situations to be concerned with normal tissue injury.
2. The LQ model may not accurately predict cell killing at high doses. It might be suggested that the model may overpredict cell killing at high doses, so the damage to late-responding normal tissues (which have smaller α/β values and therefore a more "curvy" dose−response curve) may be less than predicted by the model, thereby allowing bigger doses than predicted by the model to be used in practice.
3. There are antitumor effects of high radiation fractions that are not predicted by classic radiobiology, including enhanced antitumor immunity and secondary effects deriving from injured vasculature.
4. Many tumors may not be hypoxic, so there would be no benefit of reoxygenation between doses in a multifraction regimen.
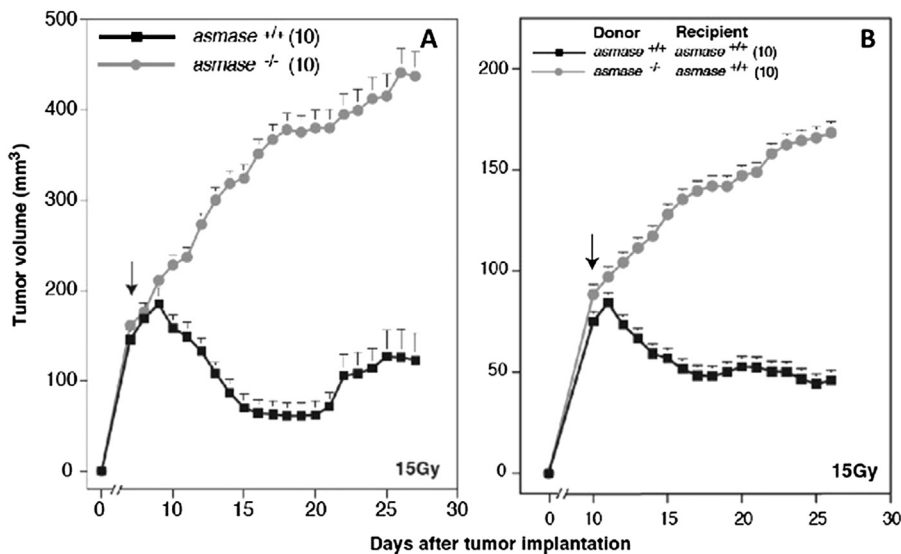
Fig. 3. Tumor response is affected by the genetics of the host. (A) Response of the MCA/129 fibrosarcoma to 15 Gy either in wild-type (asmase$^{+/+}$) (endothelial apoptosis-sensitive) or asmase$^{-/-}$ (apoptosis-resistant) mice. (B) Response of the MCA/129 fibrosarcoma in asmase$^{+/+}$ mice that had undergone transplantation with bone marrow from asmase$^{+/+}$ or asmase$^{-/-}$ mice. These data suggest that it may be the asmase$^{-/-}$ bone marrow, rather than the asmase$^{-/-}$ tumor endothelium, that confers tumor radioresistance. Adapted from (29) with permission.
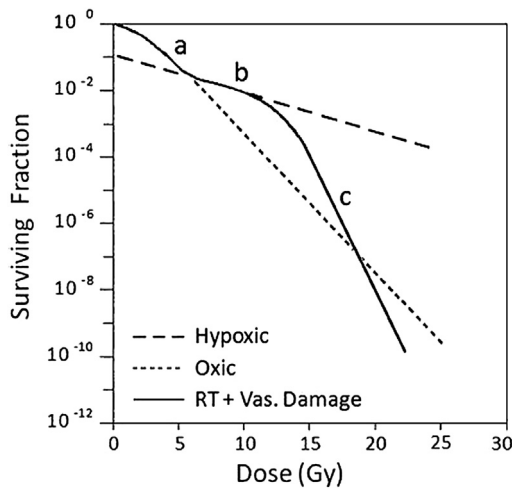
Fig. 4. Illustration of how indirect death due to vascular damage could contribute to total clonogenic cell kill in tumors irradiated with large single doses of radiation. The model assumes that 10% of the tumor cells are maximally radioresistant hypoxic cells. The dotted lines indicate the response of oxic (- - - - -) and hypoxic (− − − −) tumor cells. The response at doses 0 to 5 Gy is dominated by oxic cells (a), and that at 5 to 12 Gy is dominated by hypoxic cells (b). As radiation dose is increased above 12 Gy, it is suggested that indirect cell death due to vascular damage (c) can enhance total cell kill. From (60) with permission.

The first of these is undoubtedly true. Next we examine the evidence for the other possibilities.

Is the Linear-Quadratic Model Adequate to Describe Cell Killing at High Doses?

Clinical data from prospective randomized trials is of course the gold standard in medicine, but in the absence of good clinical outcome data, biological models should be exploited to carefully and systematically guide the selection of new or alternative treatment regimens. The ideal biological model should be accurate over the entire dose range of interest and have a small number of adjustable biological parameters that are well characterized.

The validity of the LQ model at high doses per fraction is controversial and has been critically examined by many investigators (13-18). It is well known that the LQ is only an approximation to more sophisticated kinetic reaction rate models (5, 19, 20), which, when fit only to the low-dose data, can provide a better prediction of in vitro clonogenic survival data at larger doses. LQ predictions begin to deviate, for example, from repair-misrepair and lethal−potentially lethal model predictions above ∼5 Gy for high dose rates (4, 14). However, the LQ model has been shown to fit experimental survival data well up to ∼10 Gy (14), and it may even be appropriate for single fraction doses as large as ∼15 to 20 Gy when fit over the entire dose range (15). Several empirical or semiempirical modifications have recently been proposed (14, 17, 21, 22) that introduce additional high-dose terms to synthetically straighten the survival curve at high doses. If that is done, any plausible underlying biological mechanisms of the original LQ model are lost. In a recent review of the history of the use of biologically effective dose (BED), Fowler (23) questioned the need for a straightening of the simple LQ curve beyond

an arbitrary threshold dose and suggested that instead this straightening could be achieved by assuming a higher α/β. There is a good biological rationale for higher α/β values in rapidly proliferating and hypoxic tumors. Figure 1 shows a visual comparison of model predictions for the LQ, linear-quadratic-linear (LQL) (14, 22), and universal survival curve (USC) (17) models. Substantial overlap in model predictions is achievable by simply assuming a higher α/β for the LQ model.

Although an alternative "high-dose" model may provide a superior visual fit to a specific in vitro cell survival dataset fit over a restricted dose range, it remains to be shown whether any of these models with additional high-dose terms can provide significantly better fits to multiple datasets. In addition, the utility of an empirical model decreases with the introduction of additional adjustable parameters. For example, the number of variable model parameters increases to 3 and 5 in the LQL and USC models, respectively, compared with only 2 parameters in the simpler LQ model. It would not be surprising at all if a 5-parameter model could provide a statistically superior fit to an in vitro dataset compared with a 2-parameter model. Yet, this has not been definitively demonstrated, to the knowledge of the authors. Two important questions remain unanswered at this time: (*1*) can an alternative high-dose model provide a statistically superior fit to the data considering an increase in the number of adjustable parameters; and (*2*) is there any evidence that any of these alternative models provide better estimates of clinically relevant endpoints than the conventional LQ model?

There is compelling in vitro and in vivo normal tissue evidence that the LQ model provides reasonable results at high doses (15). In particular, Figure 2 shows isoeffect results for late-responding damage to the rat spinal cord (24), for acute damage in mouse skin (25), and for early and late damage to the murine small intestine (26) up to very high single doses. All the quantitative in vivo endpoints are consistent with the LQ model, over a wide range of doses per fraction, including those of interest to SBRT, including the data for single fractions of ∼20 Gy. In addition, clinical outcome data for local tumor control can be used to compare biological models over a wide range of doses and fractionations. Recently, Mehta et al (27) analyzed the available local control data for patients with early-stage NSCLC undergoing 3-dimensional (3D) conformal radiation therapy (3D-CRT) and SBRT. They found that the clinical data could not distinguish between the LQ and USC models, suggesting that it may be difficult with tumor response to distinguish between the LQ model and LQ modification models.

We conclude therefore that the LQ model is reasonably predictive of in vitro and in vivo normal tissue dose−response relations in the dose per fraction range of 1.8 to 20 Gy, and it is not currently possible to identify an alternative high-dose model that performs better than the LQ for predicting cell killing. There is also insufficient clinical evidence at this time that the LQ needs to be modified or replaced at high doses. If alternative models do not provide a better fit to the clinical data (even with additional adjustable parameters), then there is no justification for using them regardless of how well they fit an in vitro cell survival curve. However, no model describing dose-time patterns can be fully complete or correct. Some of the main perceived mechanistic uncertainties of the LQ model have been discussed in detail by Brenner (15), so we will only summarize the conclusions reached in that review. Brenner concluded that although the mechanistic basis for the LQ model is usually attributed to pairwise production of chromosome aberrations, this does not have to be the case, and
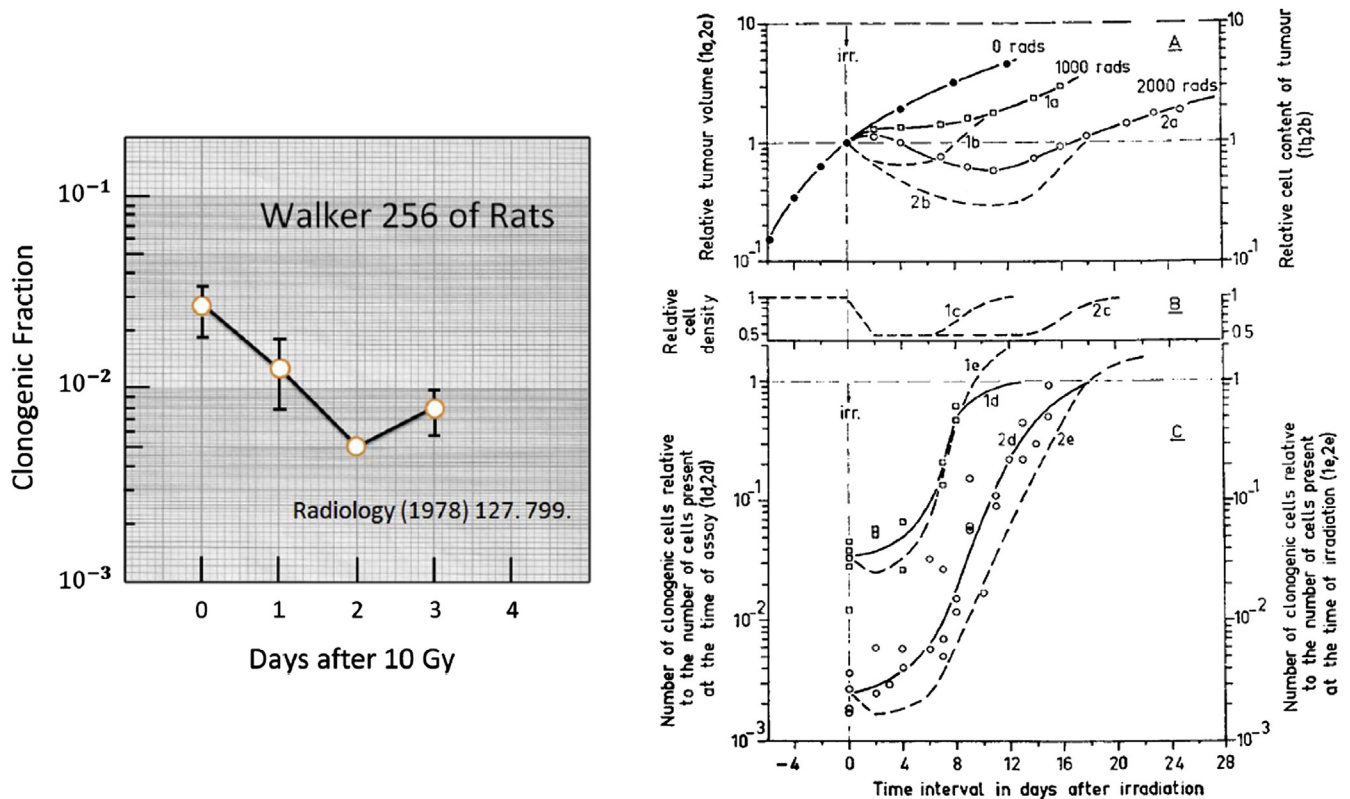
Fig. 5. Conflicting data on whether large single doses produce indirect cell kill. (A) Data on the Walker 256 tumor showing falling cell survival after a single dose of 10 Gy (originally published in 1978 and reproduced recently (60) with permission) (B) Above, gross response of the rat rhabdomyosarcoma to 10 or 20 Gy. Below, data on the cell survival from the same tumors as a function of time after irradiation. Note that there is no evidence for a fall in cell survival over the first 4 days after irradiation, before the rapid growth of the surviving cells. From (38) with permission.

the model does accommodate other cell killing mechanisms such as apoptosis and lethal mutations. One important objection often raised about the generality of the LQ model at high doses is whether repair might saturate at high doses. Two arguments suggest this is not the case. First, for normal tissues, as mentioned earlier, the dose–response curves fit the LQ model up to at least 20 Gy. Second, the rate and extent of DSB repair is similar in cells after 1 Gy (determined by γ-H2AX loss) and after 80 Gy (determined by pulsed field gel electrophoresis) (28). Thus, in the absence of any data to the contrary, it appears that saturation of repair up to doses that could conceivably be used in radiation therapy is not important.

However, given the preponderance of evidence that the LQ model fits both in vitro and in vivo normal tissue dose responses, there remains the major question of the response of tumors to irradiation, and it is this question that is the focus of the claims that high dose per fraction SBRT provides results superior to those expected from standard fractionation. We now review this question, which essentially comprises 3 major challenges to the validity of standard radiobiology to high-dose irradiation of tumors.

Biological Challenges to the 5 Rs for SRS/SBRT

As noted earlier, several biological effects have suggested that doses per fraction above 10 Gy give greater antitumor efficacy than predicted from standard radiobiological modeling, as follows:

Endothelial cell damage may enhance the cytotoxic effect of irradiation on tumor cells

The joint laboratories of Zvi Fuks and Richard Kolesnick published in 2003 an influential article (29), and expanded later (30), proposing that the radiation sensitivity of tumors to dose fractions of 10 Gy or more was governed by the sensitivity of the tumor endothelial cells to apoptosis: the same tumors in mice sensitive to radiation-induced endothelial cell apoptosis were more sensitive to radiation than those in mice resistant to endothelial cell apoptosis (Fig. 3A). However, data in the same publication suggest that there could be another explanation, namely, that the composition of the bone marrow could have affected the radiation response. Figure 3B shows that the tumors in wild-type (amase$^{+/+}$) mice could be converted from sensitive to resistant by a bone marrow transplant from endothelial apoptosis resistant (amase$^{-/-}$) mice. The authors proposed that the endothelial cells in the mice undergoing bone marrow transplantation had derived from the new bone marrow, but more recent studies of others has cast doubt on this possibility (31, 32) or have suggested that incorporation of bone marrow cells into tumor endothelium is mostly very low (33). This suggests that the asmase$^{-/-}$ character of the bone marrow, not the endothelial cells of the tumor, is responsible for the tumor resistance in this model. Another challenge to the endothelial cell apoptosis theory is that no other laboratory has independently confirmed the data; rather, most publications have shown only modest changes to the vasculature

Combined Determinants of Radiocurability
(Clonogen number & clonogen radiosensitivity)
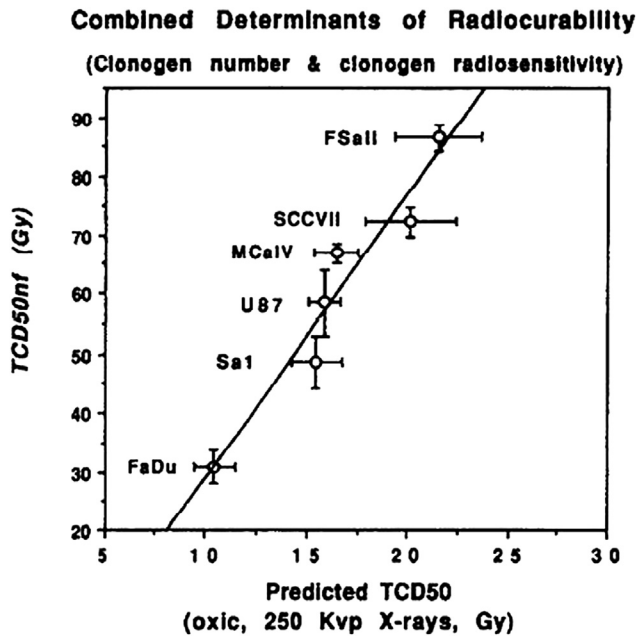
Fig. 6. The radiation dose to control 50% of the tumors (TCD50) is well predicted from the radiosensitivity of the cells in vitro and the number of cells needed to transplant the tumor (TD50). The observed TCD50 under air breathing conditions as a function of the predicted TCD50s, calculated from tumor cell radiosensitivity (in the 0- to 12-Gy range) and tumor clonogen number (from the TD50). Error bars are 1 standard deviation. From (48) with permission.

with a gradual loss of tumor endothelial cells after irradiation (34, 35). We therefore conclude that without further confirmation, the concept that rapid postirradiation endothelial damage amplifies tumor cell kill may not be generally applicable to SBRT.

Vascular damage at high doses produces secondary cell killing

This theory, suggested by Park et al (36), suggests that radiation doses higher than ~10 Gy induce vascular damage leading to indirect tumor cell death. The concept is illustrated in Figure 4. Although this is an attractive hypothesis, the data to support it are only fragmentary (37) (Fig. 5A). There are also extensive early data from Barendsen and Broese (38) on the survival of cells in a rat rhabdomyosarcoma as a function of time after single doses of both 10 and 20 Gy that show no evidence of this increasing cell kill as a function of time after irradiation (Fig. 5B). We thus conclude that there needs to be considerable more experimental evidence that this potential mechanism plays a role in the sensitivity of tumors after high dose per fraction radiation therapy.

Enhanced antitumor immunity after tumor irradiation

There is now clinical evidence that for melanoma, irradiation by SBRT of a tumor at 1 site contributes to an antitumor immunologic rejection of a metastatic lesion at a distant site—a so-called abscopal effect (39, 40). So far, the data have been reported for only 2 patients, so there are many questions to be resolved. These include whether this phenomenon is produced only at high single doses (or high doses per fraction) and whether other tumors besides melanoma experience this effect. On the first of these questions, the preclinical data suggest that although radiation enhances the antigenicity of tumors (41-43), it has been reported by Dewan et al (44) that this is greater for fractionated irradiation than for single doses. However, none of the radiation schedules tested in this study was comparable with standard fractionation: of the schedules tested (20 Gy × 1, 8 Gy × 3, and 6 Gy × 5 fractions in consecutive days), the fractionated 8 Gy was the most effective, with the 6 Gy intermediate and the 20 Gy the least effective. Thus, all these schedules could be considered to be similar to SBRT. Another preclinical study by Lee et al (45) has reported a similar enhancement of antitumor immunity by local tumor irradiation, but in this case there was a greater effect of 20 Gy × 1 than of 5 Gy × 4 over 2 weeks. Of interest is that the study in mice (44) and the clinical study with melanoma already mentioned (), the radiation was combined with anti-CTLA-4 antibody; in the case of the preclinical study there was no indication of enhanced antitumor immunity by the radiation alone, although in the study by Lee et al (45), antitumor immunity was achieved by irradiation alone. These data are clearly exciting and illustrate the fact that
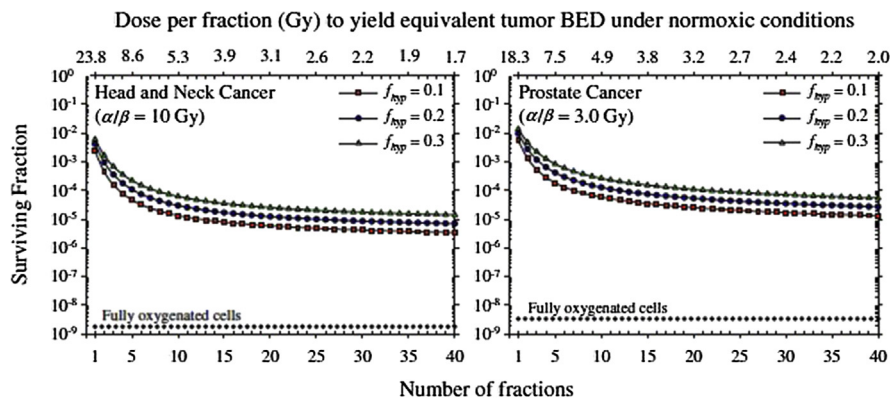
Fig. 7. Modeling (using the 5 Rs) predicts loss of efficacy of tumor cell kill for the same level of normal tissue toxicity as the dose per fraction increases. Predicted surviving fraction of tumor cells for different size dose fractionations assuming full reoxygenation between fractions, Dependence of predictions on the assumed hypoxic fraction of the tumor, f_{hyp}, is shown. It is evident that there is less cell kill predicted for very few fractions compared with standard fractionation for the same biologically effective dose (BED) (response of well-oxygenated normal tissues). From (11) with permission.
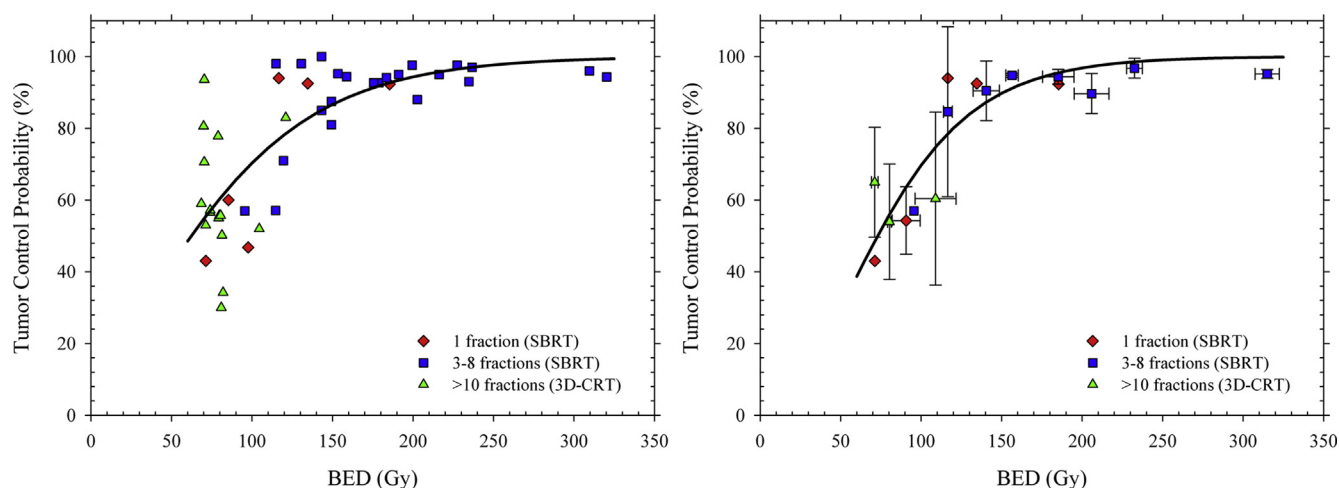
Fig. 8. Tumor control probability (TCP) as a function of biologically effective dose (BED) for stage I non-small cell lung cancer. Left, symbols show local control rates (≥2 years) from a pooled analysis reported by Mehta et al (27) with symbols distinguishing conventional and stereotactic body radiation therapy (SBRT) fractionations. Right, weighted mean TCP probabilities calculated to compensate for the different numbers of patients in each study. Solid lines show linear quadratic-based fits to the data showing that within the limits of clinical data, the efficacy of single doses, a few SBRT fractions, and conventional radiation therapy produce the same overall TCP for the same BED. From (58) with permission. 3D-CRT = 3-dimensional conformal radiation therapy.

much more information is needed in this field to enable recommendations of the best doses per fraction and timing of the radiation regimen to optimize this effect. Also of major importance is just how general the phenomena of enhanced antitumor immunity by high dose per fraction radiation therapy will be across the spectrum of tumors undergoing radiation therapy.

Preclinical Data With Tumors Do Not Support Enhanced Efficacy of High-Dose Radiation

Several investigators have addressed the question whether tumor control at high single doses can be predicted from in vitro survival curves obtained at low doses (46-48). In general these have been successful (ie, the dose to control 50% of the tumors [TCD50] is consistent with the sensitivity of the tumor cells determined at low to moderate doses). The most compelling of these data are from Gerweck et al (48), who determined the in vitro sensitivity of 6 tumor cell lines, and the number of cells needed to transplant the tumors (TD50), and showed that these 2 parameters could predict the in vivo TCD50 (Fig. 6). Importantly, 4 of the 6 tumors were from tumors that had originated spontaneously in mice and were transplanted into their respective hosts. Thus, an immunologic component could have been involved. The other 2 were human tumors transplanted into nude mice.

These data demonstrating that the TCD50 to large single doses (>20 Gy) can be predicted from the radiation survival curve at low doses (<10 Gy) do not support any extra cell kill caused by endothelial damage, vascular collapse, or enhanced immunity.

Tumor Hypoxia Is Likely to Be More Important for SRS/SBRT Than for Conventional Fractionation

It has been known for some 60 years that hypoxic cells are resistant to killing by ionizing radiation (49). A cell population

deprived of oxygen requires approximately a 3-fold larger radiation dose to produce the same amount of cell kill as a cell population exposed to physiological oxygen conditions. Hypoxia has been observed in many human cancers. Approximately 90% of all solid tumors have median oxygen concentrations less than the typical values of 40 to 60 mm Hg found in normal tissues, with many cancers having median oxygen levels below 10 mm Hg (50), which would make the cells more resistant than normal tissues to irradiation. Several investigators have now demonstrated unequivocally that the extent of tumor hypoxia has a negative impact on the ability of radiation therapy to locally control certain tumors (51, 52). This is despite the fact that fractionation of radiation mitigates the protection afforded by tumor hypoxia because of the phenomenon of reoxygenation (53), the process by which the hypoxic cells surviving a given radiation dose become oxygenated before the next radiation dose, most likely as a result of fluctuating tumor blood flow (54).

Given that tumor hypoxia has been demonstrated to have a negative impact on the efficacy of radiation therapy, at least for some tumors, it is reasonable to ask what impact it would have for SBRT and SABR. Both preclinical and modeling studies have demonstrated that tumor hypoxia will be an even greater detrimental factor for SRS.

First, the preclinical data. Some 30 years ago, Fowler and colleagues (55) measured control of transplanted mouse mammary tumors for the same level of skin damage (an early-responding normal tissue) for a variety of fractionation schemes, including single doses. They showed that large single doses of radiation were notably inferior in achieving tumor control for a given level of skin reaction (hence providing justification for fractionation in radiation therapy). Furthermore, this inferiority could be entirely overcome if the resistance of the hypoxic cells in the tumors was eliminated by pretreatment of the mice with a large dose of the hypoxic cell radiosensitizer misonidazole. In other words, fractionation is effective in improving tumor control for a given level of early-responding normal tissue damage because it partially overcomes the resistance of hypoxic cells caused by reoxygenation between doses. The LQ model predicts a further benefit of

fractionation for tumor response relative to late-responding tissues.

Second, up-to-date modeling of the influence of hypoxia on tumor response confirms that fractionation will produce more antitumor effect for the same biological effect on normal tissues (11). This is shown in Figure 7. In this study, the effect of tumor hypoxia, modeled as a continuous distribution of oxygen tensions (and radiosensitivity) from the blood vessels to the most hypoxic regions, on the survival of tumor cells is calculated for the same BED for oxygenated cells (or the cells of normal tissues). The reader is reminded that BED is an LQ model-based estimate of the effective biological dose that corrects for the effect of dose fractionation (56). The conclusions from this study are these:

1. Tumor hypoxia makes a large difference to the calculated level of cell killing even for highly fractionated irradiation, and the value of α/β or the "hypoxic fraction" has relatively less effect. This is consistent with the clinical data.
2. As the number of fractions decreases the expected tumor cell survival increases (ie, less tumor response) for the same BED or the same normal tissue damage. The worst situation is with a single dose, which gives a survival of approximately 10^{-2} compared with 10^{-5} with 30 daily fractions.
3. A logical consequence of the predicted increased importance of tumor hypoxia to the response of tumors to SBRT is that hypoxic cell radiosensitizers, which largely failed with fractionated irradiation, become a realistic option to improve the clinical outcome (11, 57).

Thus, both preclinical data and modeling studies show that tumor hypoxia is more of a detrimental factor for single dose treatments than for fractionated irradiation. Are there clinical data that can be used to address this question?

We recently analyzed tumor control data for NSCLC and for brain metastases treated both with single doses and fractionated SBRT and by conventional radiation therapy (Brenner et al, unpublished data). The results of this analysis suggest that tumor control was significantly less for single doses than fractionated irradiation for the same BED. This is consistent with the predicted loss of tumor response because of tumor hypoxia of single doses compared with fractionated radiation therapy for the same BED.

Clinical Data Suggest That Radiobiological Modeling With the Linear-Quadratic Equation Is Adequate to Explain the Efficacy of SRS and SBRT

In a recent editorial (58), we suggested that dose escalation, not "new biology," can account for the efficacy of SBRT with early-stage NSCLC. We used the term "new biology" to describe any of the already mentioned novel radiobiological mechanisms that could potentially make SBRT more effective than would be predicted from clinical experience with fractionated radiation therapy. Mehta and colleagues (27) recently reviewed the available local control data for early-stage NSCLC patients undergoing 3D-CRT and SBRT. Figure 8A shows the NSCLC tumor control probability (TCP) data as a function of BED, replotted in Brown et al (58) to clearly distinguish the data for single-fraction SBRT, multifraction SBRT, and conventional 3D-CRT. A monotonic relationship between TCP and BED is clearly observed for the 3D-CRT and SBRT data. Regardless of fractionation, higher TCPs are

obtained by delivering higher tumor BEDs. Thus, there is currently no evidence from the available NSCLC data in the literature that SBRT and 3D-CRT produce different probabilities of tumor control when corrected for tumor BED.

On the basis of the observations that: (1) TCP increases monotonically with BED and (2) the TCP versus BED relation is similar for 3D-CRT and for single-fraction and multifraction SBRT, we can say with some confidence that the great success of SBRT is due to the fact that the new stereotactic radiation therapy technologies provide dose distributions that permit the clinician to prescribe BEDs of 100 Gy or more (59). These high tumor BEDs are simply unachievable with conventional dose delivery techniques. The higher TCPs for SBRT can therefore be fully explained by the much higher tumor doses delivered, and they are entirely consistent with predictions of the LQ model. For NSCLC, there is no need to invoke a "new biology" to explain the high cure rates. We have also reached the same conclusions for brain metastases (Brenner et al, unpublished data, 2013).

References

1. Withers HR. The four R's of radiotherapy. In: Lett JT AH, editor. Advances in Radiation Biology, Vol 5. New York: Academic Press; 1975. p. 241-271.
2. Steel GG, McMillan TJ, Peacock JH. The 5Rs of radiobiology. *Int J Radiat Biol* 1989;56:1045-1048.
3. Puck TT, Marcus PI. Action of x-rays on mammalian cells. *J Exp Med* 1956;103:653-666.
4. Sachs RK, Hahnfeld P, Brenner DJ. The link between low-LET dose-response relations and the underlying kinetics of damage production/repair/misrepair. *Int J Radiat Biol* 1997;72:351-374.
5. Carlson DJ, Stewart RD, Semenenko VA, et al. Combined use of Monte Carlo DNA damage simulations and deterministic repair models to examine putative mechanisms of cell killing. *Radiat Res* 2008;169:447-459.
6. Thames HD, Bentzen SM, Turesson I, et al. Time-dose factors in radiotherapy: A review of the human data. *Radiother Oncol* 1990;19: 219-235.
7. Brenner DJ, Sachs RK, Peters LJ, et al. We forget at our peril the lessons built into the alpha/beta model. *Int J Radiat Oncol Biol Phys* 2012;82:1312-1314.
8. Horiot JC, Le Fur R, N'Guyen T, et al. Hyperfractionation versus conventional fractionation in oropharyngeal carcinoma: Final analysis of a randomized trial of the EORTC cooperative group of radiotherapy. *Radiother Oncol* 1992;25:231-241.
9. Brenner DJ, Hall EJ. Fractionation and protraction for radiotherapy of prostate carcinoma. *Int J Radiat Oncol Biol Phys* 1999;43:1095-1101.
10. Carlson DJ, Stewart RD, Li XA, et al. Comparison of in vitro and in vivo alpha/beta ratios for prostate cancer. *Phys Med Biol* 2004;49:4477-4491.
11. Carlson DJ, Keall PJ, Loo BW Jr., et al. Hypofractionation results in reduced tumor cell kill compared to conventional fractionation for tumors with regions of hypoxia. *Int J Radiat Oncol Biol Phys* 2011; 79:1188-1195.
12. Iyengar P, Timmerman RD. Stereotactic ablative radiotherapy for non-small cell lung cancer: Rationale and outcomes. *J Natl Compr Canc Netw* 2012;10:1514-1520.
13. Dutreix J, Cosset JM, Girinsky T. [Biological equivalency of high single doses used in intraoperative irradiation]. *Bull Cancer Radiother* 1990;77:125-134.
14. Guerrero M, Li XA. Extending the linear-quadratic model for large fraction doses pertinent to stereotactic radiotherapy. *Phys Med Biol* 2004;49:4825-4835.
15. Brenner DJ. The linear-quadratic model is an appropriate methodology for determining isoeffective doses at large doses per fraction. *Semin Radiat Oncol* 2008;18:234-239.

16. Kirkpatrick JP, Meyer JJ, Marks LB. The linear-quadratic model is inappropriate to model high dose per fraction effects in radiosurgery. *Semin Radiat Oncol* 2008;18:240-243.

17. Park C, Papiez L, Zhang S, et al. Universal survival curve and single fraction equivalent dose: Useful tools in understanding potency of ablative radiotherapy. *Int J Radiat Oncol Biol Phys* 2008;70:847-852.

18. Hanin LG, Zaider M. Cell-survival probability at large doses: An alternative to the linear-quadratic model. *Phys Med Biol* 2010;55: 4687-4702.

19. Tobias CA. The repair-misrepair model in radiobiology: Comparison to other models. *Radiat Res Suppl* 1985;8:S77-S95.

20. Curtis SB. Lethal and potentially lethal lesions induced by radiation: A unified repair model. *Radiat Res* 1986;106:252-270.

21. Astrahan M. Some implications of linear-quadratic-linear radiation dose-response with regard to hypofractionation. *Med Phys* 2008;35: 4161-4172.

22. Guerrero M, Carlone M. Mechanistic formulation of a lineal-quadratic-linear (LQL) model: Split-dose experiments and exponentially decaying sources. *Med Phys* 2010;37:4173-4181.

23. Fowler JF. 21 years of biologically effective dose. *Br J Radiol* 2010; 83:554-568.

24. van der Kogel AJ. Chronic effects of neutrons and charged particles on spinal cord, lung, and rectum. *Radiat Res Suppl* 1985;8:S208-S216.

25. Douglas BG, Fowler JF. The effect of multiple small doses of x rays on skin reactions in the mouse and a basic interpretation. *Radiat Res* 1976;66:401-426.

26. Peck JW, Gibbs FA. Mechanical assay of consequential and primary late radiation effects in murine small intestine: Alpha/beta analysis. *Radiat Res* 1994;138:272-281.

27. Mehta N, King CR, Agazaryan N, et al. Stereotactic body radiation therapy and 3-dimensional conformal radiotherapy for stage I non-small cell lung cancer: A pooled analysis of biological equivalent dose and local control. *Pract Radiat Oncol* 2012;2:288-295.

28. Rothkamm K, Kruger I, Thompson LH, et al. Pathways of DNA double-strand break repair during the mammalian cell cycle. *Mol Cell Biol* 2003;23:5706-5715.

29. Garcia-Barros M, Paris F, Cordon-Cardo C, et al. Tumor response to radiotherapy regulated by endothelial cell apoptosis. *Science* 2003; 300:1155-1159.

30. Fuks Z, Kolesnick R. Engaging the vascular component of the tumor response. *Cancer Cell* 2005;8:89-91.

31. Ahn GO, Brown JM. Matrix metalloproteinase-9 is required for tumor vasculogenesis but not for angiogenesis: Role of bone marrow-derived myelomonocytic cells. *Cancer Cell* 2008;13:193-205.

32. Gothert JR, Gustin SE, van Eekelen JA, et al. Genetically tagging endothelial cells in vivo: Bone marrow-derived cells do not contribute to tumor endothelium. *Blood* 2004;104:1769-1777.

33. Duda DG, Cohen KS, Kozin SV, et al. Evidence for incorporation of bone marrow-derived endothelial cells into perfused blood vessels in tumors. *Blood* 2006;107:2774-2776.

34. Kioi M, Vogel H, Schultz G, et al. Inhibition of vasculogenesis, but not angiogenesis, prevents the recurrence of glioblastoma after irradiation in mice. *J Clin Invest* 2010;120:694-705.

35. Kozin SV, Duda DG, Munn LL, et al. Neovascularization after irradiation: What is the source of newly formed vessels in recurring tumors? *J Natl Cancer Inst* 2012;104:899-905.

36. Park HJ, Griffin RJ, Hui S, et al. Radiation-induced vascular damage in tumors: Implications of vascular damage in ablative hypofractionated radiotherapy (SBRT and SRS). *Radiat Res* 2012;177:311-327.

37. Clement JJ, Tanaka N, Song CW. Tumor reoxygenation and post-irradiation vascular changes. *Radiology* 1978;127:799-803.

38. Barendsen GW, Broerse JJ. Experimental radiotherapy of a rat rhabdomyosarcoma with 15 MeV neutrons and 300 kV x-rays. I. Effects of single exposures. *Eur J Cancer* 1969;5:373-391.

39. Hiniker SM, Chen DS, Knox SJ. Abscopal effect in a patient with melanoma. *N Engl J Med* 2012;366:2035. author reply 2035-2036.

40. Postow MA, Callahan MK, Barker CA, et al. Immunologic correlates of the abscopal effect in a patient with melanoma. *N Engl J Med* 2012;366:925-931.

41. Lugade AA, Moran JP, Gerber SA, et al. Local radiation therapy of B16 melanoma tumors increases the generation of tumor antigen-specific effector cells that traffic to the tumor. *J Immunol* 2005; 174:7516-7523.

42. Apetoh L, Ghiringhelli F, Tesniere A, et al. Toll-like receptor 4-dependent contribution of the immune system to anticancer chemotherapy and radiotherapy. *Nat Med* 2007;13:1050-1059.

43. Matsumura S, Wang B, Kawashima N, et al. Radiation-induced CXCL16 release by breast cancer cells attracts effector T cells. *J Immunol* 2008;181:3099-3107.

44. Dewan MZ, Galloway AE, Kawashima N, et al. Fractionated but not single-dose radiotherapy induces an immune-mediated abscopal effect when combined with anti-CTLA-4 antibody. *Clin Cancer Res* 2009;15:5379-5388.

45. Lee Y, Auh SL, Wang Y, et al. Therapeutic effects of ablative radiation on local tumor require CD8+ T cells: Changing strategies for cancer treatment. *Blood* 2009;114:589-595.

46. Reinhold HS, De Bree C. Tumour cure rate and cell survival of a transplantable rat rhabdomyosarcoma following x-irradiation. *Eur J Cancer* 1968;4:367-374.

47. Baumann M, Dubois W, Suit HD. Response of human squamous cell carcinoma xenografts of different sizes to irradiation: Relationship of clonogenic cells, cellular radiation sensitivity in vivo, and tumor rescuing units. *Radiat Res* 1990;123:325-330.

48. Gerweck LE, Zaidi ST, Zietman A. Multivariate determinants of radiocurability. I: Prediction of single fraction tumor control doses. *Int J Radiat Oncol Biol Phys* 1994;29:57-66.

49. Gray LH, Conger AD, Ebert M, et al. Concentration of oxygen dissolved in tissues at the time of irradiation as a factor in radiotherapy. *Br J Radiol* 1953;26:638-648.

50. Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. *Nat Rev Cancer* 2004;4:437-447.

51. Nordsmark M, Overgaard M, Overgaard J. Pretreatment oxygenation predicts radiation response in advanced squamous cell carcinoma of the head and neck. *Radiother Oncol* 1996;41:31-40.

52. Brizel DM, Dodge RK, Clough RW, et al. Oxygenation of head and neck cancer: changes during radiotherapy and impact on treatment outcome. *Radiother Oncol* 1999;53:113-117.

53. Kallman RF, Dorie MJ. Tumor oxygenation and reoxygenation during radiation therapy: their importance in predicting tumor response. *Int J Radiat Oncol Biol Phys* 1986;12:681-685.

54. Brown JM. Evidence for acutely hypoxic cells in mouse tumours, and a possible mechanism of reoxygenation. *Br J Radiol* 1979;52:650-656.

55. Fowler JF, Sheldon PW, Denekamp J, et al. Optimum fractionation of the C3H mouse mammary carcinoma using x-rays, the hypoxic-cell radiosensitizer Ro-07-0582, or fast neutrons. *Int J Radiat Oncol Biol Phys* 1976;1:579-592.

56. Fowler JF. The linear-quadratic formula and progress in fractionated radiotherapy. *Br J Radiol* 1989;62:679-694.

57. Brown JM, Diehn M, Loo BW Jr. Stereotactic ablative radiotherapy should be combined with a hypoxic cell radiosensitizer. *Int J Radiat Oncol Biol Phys* 2010;78:323-327.

58. Brown JM, Brenner DJ, Carlson DJ. Dose escalation, not "new biology," can account for the efficacy of stereotactic body radiation therapy with non-small cell lung cancer. *Int J Radiat Oncol Biol Phys* 2013;85:1159-1160.

59. Lo SS, Fakiris AJ, Chang EL, et al. Stereotactic body radiation therapy: A novel treatment modality. *Nat Rev Clin Oncol* 2010;7: 44-54.

60. Song CW, Cho LC, Yuan J, et al. Radiobiology of stereotactic body radiation therapy/stereotactic radiosurgery and the linear-quadratic model. *Int J Radiat Oncol Biol Phys* 2013;87:18-19.